INFLUENCE OF SALINITY ON ALIPHATIC AND INDOLE GLUCOSINOLATES IN BROCCOLI
(Brassica oleracea var. italica)

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Abstract. Broccoli (Brassica oleracea var. italica) synthesizes sulphur containing plant secondary metabolites known as glucosinolates that are often associated with bioactive properties upon hydrolysis. Glucosinolates are part of plant defence system and the glucosinolate content in plants is usually influenced by several factors including the genetic background of plants, developmental stage, environmental factors and soil conditions. The evaluation of factors affecting glucosinolate content in plants is important to obtain plants with improved bioactive properties. The objective of the present study was to determine the effect of NaCl (0, 40, 80, 100mM) on aliphatic and indole glucosinolate content of broccoli grown in greenhouse conditions. For this purpose, broccoli seedlings at 5-6 leaf were irrigated with 0 (control), 40mM, 80mM and 100mM NaCl. Glucosinolates were determined at three different time points (1, 3, 6 days upon treatment). According to the findings, the amount of individual and total aliphatics and indoles were lower at 40, 80 and 100mM NaCl applications compared to control plants determined at three different time points (P<0.001) suggesting the break down of glucosinolates due to stress conditions. The findings of the study revealed the glucosinolate profile and content of broccoli seedlings under saline conditions.

Keywords: brassica, stress, salinity, secondary metabolites

Introduction

Plants are exposed to various environmental conditions including climate and soil factors during their growth cycle. Unfavourable conditions such as high or low temperatures, drought, salinity and mineral nutrient deficiency in soil may limit the plant growth and development. Salinity is one of the major abiotic stress factors affecting plant physiology, growth and development (Zhu, 2001). Plants respond to such limiting conditions through the activation of their primary and secondary metabolism in order to overcome stress conditions and survive (Khan et al., 2009; Velasco et al., 2007). Under salinity stress, plants undergo an osmotic adjustment. In addition, in order to maintain the water balance, cells accumulate secondary metabolites. Glucosinolates, ascorbic acid, phenolic compounds and anthocyanin are the major secondary metabolites involved in plant defense against stress conditions (López-Berenguer et al., 2008).

Glucosinolates are plant secondary metabolites mainly present in the Brassicaceae and known to act as a plant defence system against insect and pathogen attacks (Cole, 1997) as well as environmental stress factors such as salinity (Qasim et al., 2003). Glucosinolates have attracted a great deal of attention delivering antimicrobial (Aires et al. 2009), antioxidant (Nilsson et al., 2006) and anticarcinogenic properties (Ávila et al., 2013). These
beneficial effects are attributed to isothiocyanates which are the hydrolysis products of glucosinolates (Navarro et al., 2011). Glucoraphanin (4-methylsulphinylbutyl) in broccoli has been of interest as the precursor of sulforaphane 1-isothiocyanato-4-methylsulphinylbutane.

Salinity has been shown to influence glucosinolate content in plants. However, the effect is dependent on many factors including the developmental stages of the plant, the tissue and plant part analyzed, the cultivar effect and salinity doses applied. In addition variation exists in terms of aliphatic and indole glucosinolates. Total glucosinolate content in broccoli leaves showed a slight increase after 1 day of saline treatment (40 and 80mM), but with no significant differences with regard to the controls. However, at 15 days, the total glucosinolates content was found higher in control leaves than in 40 mM NaCl-treated plants, with a marked increase at 80 mM NaCl (López-Berenguer et al., 2008). Hassini et al. (2016) reported that salinity (150mM) did not change glucosinolate content in broccoli.

The amount and pattern of glucosinolates may vary according to the genetic background of individuals as well as various environment and ontogenic factors as reviewed (Sarıkamış, 2009). Similarly, plant tolerance to stress factors may vary accordingly.

The objective of the present study is to assess the influence of salinity on aliphatic and indole glucosinolate content in young broccoli seedlings in order to evaluate the status of these health promoting compounds under stress conditions at earlier developmental stages.

**Material and Methods**

**Plant Material**

The experiment was conducted at the Department of Horticulture, Faculty of Agriculture, Ankara University. Broccoli (*Brassica oleracea* var. *italica* ‘Marathon’) seeds were sown in plug trays (5.5 cm width and 6 cm depth) containing a mixture of peatmoss (Plantaflor-Humus, Verkaufs-GmBH, Germany) and perlite (1:1). Seedlings at 2-3 leaf stage were transferred to 7 L containers filled with peat moss:perlite:cocopeat (1:1:1) mixture and maintained in the greenhouse. Daily temperature and humidity were recorded from measurements taken every 20 min using dataloggers in greenhouse (Fig. 1). Daily mean temperature were averaged between 21.72-28ºC and relative humidity between 53.2-63.11% in the greenhouse during the experiment.

**Salinity Treatments**

Broccoli seedlings at 5-6 leaf stage (2 weeks after transplanted to the plastic containers) were irrigated with 0 (control), 40mM, 80mM and 100mM NaCl. The pH of the growing media measured as 6.5-7.5. Leaf samples were collected at 1, 3, 6 days after NaCl treatment.

**Analysis of Glucosinolates**

The glucosinolates were extracted from two upper young leaves. Leaf samples were freeze-dried prior to the analysis. Extraction of glucosinolates, conversion to desulfoglucosinolates and analysis by HPLC was as described previously by (Sarikamis et al., 2006). Samples were analysed and separated by HPLC-UV (Shimadzu®) in the HPLC laboratory at Ankara University, Department of Horticulture. A volume of 50 μl from the extract was injected onto a Waters Spherisorb 5μM ODS 2, 4.6x250mm analytical cartridge. Analysis was carried out on a gradient of 99% water and 1%
acetonitrile at a flow rate of 1ml/min for 24 min. The detection was carried out at a wavelength of 229 nm. Glucotropaeolin (benzyl glucosinolate) was used as the internal standard for the quantification of the peaks. Glucosinolates were quantified according to internal standards and expressed as µmol g⁻¹ dry weight (DW). Correction factors were used during quantification for each compound as listed by (Brown et al., 2003).

Statistical Analysis

The experiment was conducted according to completely randomized design as three replicates each replicate contained four plants. Experimental data were expressed as the mean ± standard error of the mean with three replications (n = 3). One-way analysis of variance and Duncan’s multiple range test was used to determine the significance of differences using SPSS 18.0 (SPSS Inc., Chicago, IL). Significant differences were evaluated at P<0.001 error level.

Results and Discussion

The effect of NaCl treatments (0, 40, 80, 100mM) on aliphatic and indole glucosinolate content of broccoli seedlings were determined on leaf samples collected at three different time points (1, 3, 6 days upon treatment) in greenhouse conditions. Glucosinolates determined were glucoiberin (3-methylsulphinlypropyl), glucoraphanin (4-methylsulphynylbutyl), glucobrassicin (3-indolylglucobrassicin), 4-methoxyglucobrassicin (4-methoxy-3-indolylmethyl), neoglucobrassicin (1-methoxy-3-indolylmethyl) and 4-hydroxyglucobrassicin (4-hydroxy-3-indolylmethyl). The predominant glucosinolates were glucobrassicin of the indole group followed by glucoraphanin of the aliphatics (Table 1).
Table 1. Glucosinolate content of broccoli seedlings at 0, 40, 80, 100mM NaCl applications on 1, 3, 6 days

<table>
<thead>
<tr>
<th>Glucosinolates</th>
<th>Time</th>
<th>0mM</th>
<th>40mM</th>
<th>80mM</th>
<th>100mM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucoiberin</td>
<td>Day 1</td>
<td>0.890±0.118a</td>
<td>0.543±0.127b</td>
<td>0.467±0.114b</td>
<td>0.667±0.143b</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>1.003±0.173a</td>
<td>0.247±0.154b</td>
<td>0.82±0.09b</td>
<td>0.6825±0.0962b</td>
</tr>
<tr>
<td></td>
<td>Day 6</td>
<td>1.270±0.228a</td>
<td>1.060±0.348a</td>
<td>0.387±0.194b</td>
<td>0.870.478ab</td>
</tr>
<tr>
<td>Glucoraphanin</td>
<td>Day 1</td>
<td>4.640±0.386a</td>
<td>2.250±1.22ab</td>
<td>1.820±1.20b</td>
<td>3.817±0.556ab</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>5.260±0.460a</td>
<td>3.790±1.34b</td>
<td>2.060±0.960b</td>
<td>3.438±0.895b</td>
</tr>
<tr>
<td></td>
<td>Day 6</td>
<td>5.687±0.463a</td>
<td>4.730±2.11ab</td>
<td>2.630±1.13b</td>
<td>4.540±2.84ab</td>
</tr>
<tr>
<td>Glucobrassicin</td>
<td>Day 1</td>
<td>13.167±0.524a</td>
<td>8.840±3.57ab</td>
<td>6.910±2b</td>
<td>11.920±1.99a</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>15.293±0.695a</td>
<td>4.3367±0.0865b</td>
<td>9.83±5.13ab</td>
<td>8.190±0.817ab</td>
</tr>
<tr>
<td></td>
<td>Day 6</td>
<td>15.103±0.211a</td>
<td>9.300±2.11ab</td>
<td>5.28±1.21b</td>
<td>11.280±0.617ab</td>
</tr>
<tr>
<td>4-Methoxyglucobrassicin</td>
<td>Day 1</td>
<td>1.550±0.407a</td>
<td>0.630±1.092b</td>
<td>0.417±0.723b</td>
<td>0.491±0.851b</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>1.570±0.466a</td>
<td>0.8067±0.0841b</td>
<td>2.24±1.37a</td>
<td>1.480±0.225ab</td>
</tr>
<tr>
<td></td>
<td>Day 6</td>
<td>1.883±0.196ab</td>
<td>3.05±1.02a</td>
<td>0.853±0.267b</td>
<td>1.537±0.748ab</td>
</tr>
<tr>
<td>1- Methoxyglucobrassicin</td>
<td>Day 1</td>
<td>1.593±0.348a</td>
<td>2.61±1.29a</td>
<td>2.373±0.854a</td>
<td>1.510±1.04a</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>1.307±0.575a</td>
<td>2.810±0.361a</td>
<td>2.780±4.41a</td>
<td>1.950±0.682a</td>
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<tr>
<td></td>
<td>Day 6</td>
<td>1.350±0.617a</td>
<td>1.50±4.41a</td>
<td>1.92±1.75a</td>
<td>1.570±2.54a</td>
</tr>
<tr>
<td>4-Hydroxyglucobrassicin</td>
<td>Day 1</td>
<td>0.397±0.116a</td>
<td>0.206±0.0821a</td>
<td>0.176±0.0623a</td>
<td>0.396±0.0821a</td>
</tr>
<tr>
<td></td>
<td>Day 3</td>
<td>0.333±0.145a</td>
<td>0.086±0.0033a</td>
<td>0.280±0.190a</td>
<td>0.267±0.0377a</td>
</tr>
<tr>
<td></td>
<td>Day 6</td>
<td>0.413±0.139a</td>
<td>0.416±0.0769a</td>
<td>0.500±0.0987a</td>
<td>0.620±0.0850a</td>
</tr>
</tbody>
</table>
Total aliphatic glucosinolates at day 1 were determined as 5.53±0.40 µmol g⁻¹ DW, while total indoles were determined as 16.71±0.71 µmol g⁻¹ DW in control plants. The amount of total aliphatics and indoles were decreased at 40 and 80 mM NaCl, aliphatics determined as 2.79±1.20, 2.28±1.14 and indoles as 12.29±2.21, 9.88±0.35 µmol g⁻¹ DW respectively which was later slightly increased to 4.48±0.96 µmol g⁻¹ DW in aliphatics and 14.32±1.29 µmol g⁻¹ DW for indoles at 100 mM but still lower than control plants (P<0.001) (Fig. 2). On the third day of application, total aliphatic and indoles were decreased significantly at 40, 80, 100 mM doses (P<0.001). On the sixth day of application, similar to day 1, a decrease was observed compared to the control plants at 40, 80 mM and at 100 mM both in aliphatics and indoles (P<0.001) (Fig. 2).

Figure 2. Changes in total and aliphatic glucosinolates at 0 (control), 40, 80, 100 mM NaCl applications (days 1, 3, 6)

In terms of individual glucosinolates, glucoraphanin was the predominant aliphatic glucosinolate compared to glucoiberin. Glucoiberin content decreased at 40, 80, 100 mM NaCl applications at day 1 and 3, and decreased at 80 and 100 mM at day 6 (P<0.001). Similarly glucoraphanin content decreased with NaCl applications (P<0.001) (Table 1).

Glucoibassicin determined as the predominant indole glucosinolate. Glucobrassicin content decreased at 40, 80 NaCl concentrations. At day 3, the increase was highest at 80 mM NaCl (P<0.001). A decreasing trend was observed at 4-methoxyglucobrassicin except day 3 where an increase was observed at 80 mM and day 3, the highest level was reached at 40 mM NaCl. Slight increase was determined in 1-methoxyglucobrassicin at 40 and 80 mM NaCl however the increase was not statistically important (P>0.001). The amount of 4-hydroxyglucobrassicin was lowest among indoles and there was not significant variation among treatments (P>0.001).

Glucoisnolates are a group of secondary metabolites usually associated with health promoting properties (Dinkova-Kostova and Kostov, 2012; Verkerk et al., 2009). In a previous study, glucosinolates were shown to be influenced by salinity stress (Pang et al., 2012) suggesting that under low water potential they could be involved in osmotic adjustment and might be involved in salt tolerance in plants (Lopez Berenguer et al., 2009).
In the present study, the individual and total aliphatic glucosinolates as well as indoles of the broccoli seedlings were decreased significantly by NaCl application at 40 and 80 mM NaCl compared to control plants whereas a slight but insignificant increase was observed at 100 mM NaCl. A few studies report the influence of salinity on the content of glucosinolates in broccoli florets (Lopez Berenguer et al., 2009), radish sprouts (Yuan et al., 2010), broccoli sprouts (Guo et al., 2014) suggesting differential responses to salinity. The content of intact aliphatic and indole glucosinolates in the inflorescences of broccoli cultivars Nubia, Naxos and Parthenon has also revealed differential response to salinity treatments (Dominguez Perles et al., 2011). In another work aimed to improve health promoting glucosinolates in edible sprouts suggested that glucoraphanin content decreased significantly, indole glucosinolates neoglucobrassicin, 4-hydroxyglucobrassicin and glucobrassicin were reported to decrease by 40 and 80 mm NaCl and increase by 100 mM NaCl application in 7d old broccoli sprouts germinated under different NaCl doses (Guo et al., 2013). Different from above mentioned studies, our work aimed to determine the influence of salinity on glucosinolates in broccoli seedlings representing earlier developmental stages.

Although a few studies highlight the different responses of glucosinolates to salt stress, the mechanism behind is still not clear. The differential variation of glucosinolates is related to the strong metabolism alteration, turgor adjusting leading to a high growth reduction (López-Berenguer et al., 2009). On the other hand, exposure to stress may lead to cell membrane damage and the decrease in relative water content (Wang et al., 2013). Therefore, the decreases in glucosinolate content can be attributed to the break down of glucosinolates (hydrolysis) due to cellular damage caused by oxidative stress.

Conclusion

Broccoli contains sulforaphane that is often associated with health promoting activities including the anticancer properties. The precursor of sulforaphane is glucoraphanin which is an aliphatic glucosinolate and the amount of glucosinolates, glucoraphanin in particular, is important in order to gain health benefits. However, the amount of glucosinolates depend on several factors including the environmental and soil conditions during the growth period, stress factors and the developmental stage of the plant. Therefore in the present study, the effect of saline conditions on glucosinolate content of broccoli were determined in broccoli seedlings at 5-6 leaf stage grown in greenhouse conditions. The findings suggested an overall decrease in aliphatic glucosinolates and glucoraphanin as well as total glucosinolates compared to control plants. The decrease was attributed to the degradation of glucosinolates via the enzyme myrosinase also present in plant cells probably activated upon cellular damage caused by NaCl applications.

REFERENCES


