GENOME-WIDE IDENTIFICATION OF TCP TRANSCRIPTION FACTORS FAMILY IN *POPULUS* **SECT.** *TURANGA* **(***POPULUS PRUINOSA* **SCHRENK AND** *POPULUS EUPHRATICA* **OLIVE) REVEALED THE ROLES OF** *TCPS* **IN LEAF MORPHOLOGY**

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(Received $27th$ Oct 2022; accepted $20th$ Jan 2023)

Abstract. The TCP (TEOSINTE BRANCHED1/CYCLOIDEA/PROLIFERATING CELL FACTORS)

gene family is a plant-specific transcription factor family and plays an important regulatory role in plant growth and development. However, the identification, characterization, and expression levels of *TCPs* during leaf development in *Populus pruinosa* and *Populus euphratica* remain unclear. In this study, 33 and 34 non-redundant *TCPs* in *P. pruinosa* and *P. euphratica*, (*PpTCPs*/*PeTCPs*) are identified, respectively, which contain TCP-conserved domains and are unevenly distributed on 19 chromosomes. Among them, *PpTCP19*, *PpTCP27*, *PeTCP19,* and *PeTCP28* undergo positive selection in the *Populus* sect. *Turanga*. Furthermore, transcriptome data on different leaf morphologies of *P. pruinosa*/*P. euphratica* and available functional data of *TCPs* in *Arabidopsis thaliana* (*AtTCP*) support the involvement of the TCP gene family in the leaf development of the *Populus* sect. *Turanga. PpTCP7*, *PpTCP19*, *PeTCP7,* and *PeTCP19* may regulate leaf morphology by restricting cell division at the boundaries of leaves and sepals in *P. pruinosa* and *P. euphratica*, respectively. These results elucidate a foundation for an in-depth analysis of the correlation between the expression pattern of the *TCP* gene and leaf morphology changes in the *Populus* sect. *Turanga*, and provide valuable information for the functional study of *TCP* transcription factors in the *Populus* sect. *Turanga*.

Keywords: *desert poplar species, phylogenetic analysis, TCP gene family, heteromorphic leaves*

Introduction

Plant transcription factors (TFs) respond to various biotic and abiotic stresses by regulating the expression of target genes during plant growth and development (Huo et al., 2019; Jin et al., 2017). *TCPs* are one of the largest families that constitute plantspecific TF (Martín-Trillo et al., 2010). The TCP gene family is named after the earliest identified members of the TCP domain, including TB1 (teosinte branched1) from *Zea mays* L*.*, CYC (cycloidea) in *Antirrhinum majus* L., and PCF1 and PCF 2

(PROLIFERATING CELL FACTORS 1 and 2) in *Oryza sativa* L. (Luo et al., 1996; Doebley et al., 1997; Kosugi and Ohash, 1997). In addition, it contains approximately 59 amino acids and has an atypical basic helix (bHLH) structure (Cubas et al., 1999), and is located at the N-terminus, thereby allowing it to interact with DNA or proteins (Kosugi and Ohashi, 2002). Plant-specific *TCPs* play regulatory roles in different processes of plant growth and development, including the establishment of leaf morphology (Yu et al., 2022; Palatnik et al., 2003; Qi et al., 2019; Zhang et al., 2021; Li et al., 2022; Lin et al., 2016), the formation of trichomes (Vadde et al., 2018), and the floral organ morphogenesis as well as leaf growth (Nag et al., 2009; Koyama et al., 2011). Functional analysis of the CINCINNATA (CIN) subclass genes in *Arabidopsis* demonstrate that class II *TCP*s participate in plant leaf morphogenesis by inhibiting the proliferation of leaf margin cells (Palatnik et al., 2003). *TCP15* in *Arabidopsis* is involved in ROS-mediated signal transduction during exposure to high-light-intensity conditions (Viola et al., 2016). Moreover, *AtTCP14* and *AtTCP15* control the SPINDLY (SPY) sensitivity to cytokine (CK) and regulate the expressions of CK-responsive genes (Steiner et al., 2012). Furthermore, miR156 interacts with TCP through its target SPL9, the complex that promotes the complexity of leaves under the control of cup-shaped cotyledon (CUC), thereby suggesting that TCP may play an important role in the leaf morphology regulatory cascade center on the miR156 module (Wang et al., 2011; Yang et al., 2011).

Herein, *P. pruinosa* and *P. euphratica* have been studied, both belong to the sister species of the *Populus* sect. *Turanga* and are dominant tree species that occur commonly in the arid deserts of Central Asia (Gai et al., 2021). The natural distribution of *P. pruinosa* is limited to Western China and adjacent areas, while *P. euphratica* extends from Western China to Southern Morocco. They become natural protective barriers for desert forest ecosystems because of their high tolerance to salinity and drought stress (Sun et al., 2023; Wu et al., 2022; Han et al., 2013). A leaf is an organ with the largest area exposed to the atmosphere, and the change of its shape will reflect plants' adaptation to the environment. The morphological features of *P. pruinosa* and *P. euphratica* differentiate well, and they both have heteromorphic leaves. The leaf morphology of an adult *P. pruinosa* is oblong, round, and broad ovate from the bottom to the top of the canopy (Liu et al., 2016), meanwhile, adult *P. euphratica* is strip, lanceolate, ovate, and broad ovate from bottom to top (Wang et al., 1998; Hao, 2017; Zheng et al., 2007). Liu et al. (2016) studied the ontogeny process of the heteromorphic leaves of *P. pruinosa* and found that their ontogeny process appeared in sequence with oblong, round, and broad ovate leaves. Different photosynthetic areas and the accumulation of photosynthetic products respond to the constant demand for photosynthetic products and mineral nutrient metabolites in individual growth and development. Meanwhile, *P. euphratica* appears linear, lanceolate, ovate, and broad ovate leaves in turn with the process of individual development. Zeng et al. (Zeng, 2020) also studied the physiological and biochemical characteristics of typical heteromorphic leaves of *P. euphratica* and found that the specific leaf weight, dark respiration rate, and light saturation point of *P. euphratica* in linear, lanceolate, ovate, and broad ovate leaves gradually increased, which made it to have a different accumulation of photosynthetic products at different stages of development. Although various studies have reported the physiological and biochemical characteristics of heteromorphic leaves of *P. pruinosa* and *P. euphratica*, the role of *TCPs* in the leaf morphology of the *Populus* sect. *Turanga* species remains unknown.

This study conducted a systematic analysis of the TCP gene family of the *Populus* sect. *Turanga* including the identification of *TCPs*, physicochemical properties, phylogeny, and expression patterns during leaf morphology changes in the *Populus* sect. *Turanga.* Consequently, our data offer detailed information on *TCPs* in *P. pruinosa* and *P. euphratica* (*PpTCPs*/*PeTCPs*) character, which deeply improves our understanding of *TCPs* in the *Populus* sect. *Turanga* and elucidates the function of *PpTCPs/PeTCPs* in leaf morphology.

Materials and methods

Plant materials and experimental design

All samples of this experiment were sourced from the *P. pruinosa* and *P. euphratica* forest (40°32′36.90″N, 81°17′56.52″E) in Alar City, Xinjiang Province, China. *P. pruinosa* and *P. euphratica* leaves matured from the end of July to August. When there were 7–13 leaves on a bud, the first three fastest growing leaves in the branch of oblong, round, and broad ovate leaves of *P. pruinosa*, the linear, lanceolate, ovate, and broad ovate leaves of *P. euphratica* were selected as materials based on the variation law of *P.*hl (Zhao and Qin., 2017). All samples were quickly frozen in liquid nitrogen and stored at −80 °C for transcriptome sequencing. This experiment was performed in three biological replicates (three biological replicates here indicated three different plants).

Identification of TCPs in Populus sect. Turanga

PpTCPs/PeTCPs were analyzed based on *P. pruinosa* (https://www.ncbi.nlm.nih.gov/PRJCA006811)/*P. euphratica* genome numbers (https://ngdc.cncb.ac.cn/PRJCA005959/). The BLAST algorithm was applied to identify all potential *PpTCPs*/*PeTCPs* with TCP or TCP-like domains. BLASTP searches were performed to identify chromosome-level genomes in *P. pruinosa* and *P. euphratica* using amino acid sequences containing the TCP or TCP-like domains of *Arabidopsis* proteins (the genetic information of *P. pruinosa* can be obtained from the genome of *P. pruinosa* at the chromosome level. https://figshare.com/articles/online_resource/Pprgenome_fa/20705107/2). In addition, TCP proteins were identified using HMMER (http://hmmer.org/Download.html). Hidden Markov Model (HMM) profiles corresponding to the TCP-conserved domain (PF03634) downloaded from the Pfam protein family database (http://pfam.xfam.org/search) were scanned to identify the TCP proteins (Potter and Finn, 2018). Then, the same TCP-like domain sequences were verified using SMART (Simple Modular Architecture Research Tool, http://smart.embl-heidelberg.de/) and NCBI-CDD (https://www.ncbi.nlm.nih.gov/cdd). At E values $< 1e^{-5}$, the protein was identified as a member of the *PpTCPs/PeTCPs* transcription factor family. Finally, the physicochemical properties of *PpTCP/PeTCP* proteins, including amino acid length (aa), protein molecular weight (mw), and isoelectric points (pI) were obtained using ProtParam in the ExPASY website (https://web.expasy.org/protparam/) (Duvaud et al., 2021). Wolf PAORT (https://wolfpsort.hgc.jp/) was used to predict the subcellular localization of *PpTCP/PeTCP* proteins (Horton et al., 2007).

Gene duplications and repeats sequence of Populus sect. Turanga

Genome-wide replication modes were identified using MCScanX (https://megasoftware.net/), and replication patterns of *PpTCPs/PeTCPs* were counted. After being manually inspected, SMART was used to analyze the repeat amino acid sites in *PpTCP/PeTCP* proteins. Then, the alignments of full-length amino acid sequences of *PpTCPs/PeTCPs* were performed using the MUSCLE method of MEGA-X with default settings. Subsequently, after amino acid sequence alignment, gap trimming was performed using the multiple alignment trimming tool of TBtools software with a site coverage cutoff parameter of 0.95.

Phylogenetic relationship, gene structure and conserved motif composition of PpTCPs/PeTCPs

The gap trimming was conducted with MUSCLE method of MEGA-X with default settings and a parameter of 0.95 (Wang et al., 2010). The neighbor-joining method was used to build a phylogenetic tree (neighbor-joining), the bootstrapping repetition algorithm was set to 1000, and other parameters were set at default. The structure of *PpTCPs/PeTCPs* encoded proteins and the conserved motifs were screened and identified using the online website GSDS (http://gsds.cbi.pku.edu.cn/) and MEME (http://meme-suite.org/tools/meme). Based on this, the results of domain analysis were combined, and the structure and conserved motifs were drawn with TBtools (Chen et al., 2020).

Collinearity and phylogenetic analysis of TCPs in multispecies

Orthologous pairs of *P. pruinosa* and *Arabidopsis*, *Salix brachista,* and *P. euphratica* were aligned using the BLASTP. Then, collinear regions between *P. pruinosa* and *Arabidopsis*, *P. pruinosa* and *Salix brachista*, as well as *P. pruinosa* and *P. euphratica* were screened by MCscan and visualized using JCVI (https://zenodo.org/record/31631/). The SMART website was used to retrieve the domain coordinates in the TCP protein sequences of *P. pruinosa*/*P. euphratica* and *Arabidopsis*. Subsequently, the combined protein sequences were used to construct a phylogenetic tree between the two species using the EvolView (http://www.evolgenius.info/evolview/) and TBtools software was used to display the phylogenetic tree. Identical (Ka) and non-identical mutation frequency (Ks) values of the *PpTCPs* sequence were calculated using the TBtools software. $Ka/Ks < 1$, $Ka/Ks = 1$, and $Ka/Ks > 1$ indicated purification selection pressure, neutral evolution, and positive selection pressure, respectively.

Prediction of cis-acting elements in the promoters of TCPs of Populus sect. Turanga

The 2000 bp upstream sequence of the CDS transcription start site of *PpTCPs/PeTCPs* was extracted using TBtools, and Plant CARE (http://bioinformatics.psb.ugent.be/webtools/plantcare/html/) was used as a *cis*-acting element to predict the promoter region.

Transcriptome sequencing and data analysis of TCPs at different leaf morphology

Different leaf morphologies and their control samples of *P. pruinosa* and *P. pruinosa* leaves were selected for transcriptome sequencing. Moreover, RNA extraction, cDNA

library construction, RNA-seq, and raw data analysis were performed by Illumina novaseq 6000 (Frasergen, Wuhan, China). After the library was qualified, DNB (DNA Nano Ball) was prepared, then loaded onto the sequencing chip, and sequenced using a high-throughput sequencer made by MGI; the SOAPnuke (v2.1.0) software was used (Chen et al., 2018) to filter the original reads to obtain high-quality clean reads to process off-machine data. Afterward, the obtained high-quality Illumina clean reads were compared with *P. pruinosa* and *P. euphratica* reference genomes using the Hisat2 (v2.1.0) software (Kim et al., 2019). Meanwhile, the StringTie (v1.3.4d) (Pertea et al., 2016) software was used to carry out quantitative expression analysis of genes in each sample. Gene expression patterns were quantified using FPKM (fragments per kilobase per million), which essentially referred to fragments per kilobase as compared to exons of the reference genome per million reads. Among them, |log2 Fold Change| > 1.5 and *P* value < 0.05 were considered differentially expressed genes. The obtained data were subjected to cluster analysis and expression heat map drawing using the TBtools software.

Results

Identification and chromosomal classification of PpTCPs and PeTCPs

Herein, a total of 33 *PpTCPs* were identified in the *P. pruinosa* genome by HMMER and BLASTP, and they were named *PpTCP1*–*PpTCP33* based on the chromosomal arrangement of genes in the genome. A total of 34 *PeTCPs* were identified in the *P. euphratica* genome using the same methods, and they were named *PeTCP1*–*PeTCP34*. Chromosome mapping found that *TCPs* were unevenly distributed on the 19 chromosomes of the two species of the poplar. Moreover, *TCPs* were distributed on 17 out of the 19 *P. pruinosa* chromosomes in an uneven manner, with the number of *PpTCPs* per chromosome ranging from 0 to 5. Chromosomes 4 contained five genes, while no *TCP* was found on Chromosomes 17 and 18 (*Fig. 1A*). Meanwhile, a total of 34 *PeTCPs* were distributed across 19 chromosomes, except for Chr08 with different densities (*Fig. 1B*), these results suggested that each chromosome contributed differently to the evolution of *PpTCPs*/*PeTCPs*. The amino acid number, protein molecular weight (MW), and isoelectric point values of *PpTCP* proteins ranged from 176 (*PpTCP22*) to 572 (*PpTCP16*), from 19.32 kDa (*PpTCP22*) to 60.15 (*PpTCP16*), and from 5.44 (*PpTCP22*) to 9.96 (*PpTCP17*), respectively. Meanwhile, the amino acid number, MV, and the isoelectric point value of *PeTCP* proteins ranged from 120 (*PeTCP5*) to 661 (*PeTCP20*), from 12.96 kDa (*PeTCP5*) to 73.09 kDa (*PeTCP20*), and from 6.51 (*PeTCP29*) to 10.95 (*PeTCP5*), respectively (*Tables 1a, b, c* and *Electronic Appendix 1*). In addition, subcellular localization found that *PpTCP*/*PeTCP* proteins were primarily located in the nucleus.

TCP gene	Subcellular localization
PpTCP1	nucl: 14
PpTCP2	nucl: 13, golg: 1 PpTCP3 details nucl: 12, chlo: 1, extr: 1
P _p TCP ₃	nucl: 12, chlo: 1, extr: 1
PpTCP4	nucl: 12.5, cyto_nucl: 7.5, cyto: 1.5
PpTCP5	nucl: 13 , golg: 1
PpTCP6	nucl: 12.5, cyto_nucl: 7, plas: 1

Table 1a. The identification and character analysis of PpTCPs

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PpTCP7	nucl: 13, cyto: 1
PpTCP8	nucl: 13, chlo: 1
PpTCP9	nucl: 7, cyto: 5, extr: 1, cysk: 1
PpTCP10	nucl: 9, chlo: 1, cyto: 1, extr: 1, vacu: 1, pero: 1
PpTCP11	nucl: 14
PpTCP12	nucl: 10.5, nucl_plas: 6, mito: 2, cyto: 1
PpTCP13	chlo: 6, mito: 5, nucl: 3
PpTCP15	nucl: 10, mito: 2, plas: 1.5, golg_plas: 1.5
PpTCP16	nucl: 14
PpTCP17	nucl: 14
PpTCP18	nucl: 14
PpTCP19	nucl: 13, chlo: 1
PpTCP20	nucl: 12, chlo: 1, cyto: 1
PpTCP21	nucl: 13.5, cyto_nucl: 7.5
PpTCP22	nucl: 11, cyto: 2, cysk: 1
PpTCP23	nucl: 14
PpTCP24	nucl: 14 PpTCP14 details nucl: 12, chlo: 1, cyto: 1
PpTCP25	nucl: 13, plas: 1
PpTCP26	nucl: 14
PpTCP26	nucl: 14
PpTCP27	nucl: 10.5, cyto_nucl: 6, chlo: 1, extr: 1, vacu: 1
PpTCP28	nucl: 14
PpTCP29	chlo: 7, nucl: 3.5, mito: 3, cyto_nucl: 2.5
PpTCP30	nucl: 14
PpTCP31	nucl: 14
PpTCP32	nucl: 14
PpTCP33	nucl: 13.5, cyto_nucl: 7.5

Table 1b. The identification and character analysis of PpTCPs

APPLIED ECOLOGY AND ENVIRONMENTAL RESEARCH 21(2):1665-1696. http://www.aloki.hu ● ISSN 1589 1623 (Print) ● ISSN 1785 0037 (Online) DOI: http://dx.doi.org/10.15666/aeer/2102_16651696 © 2023, ALÖKI Kft., Budapest, Hungary

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PpTCP25	PprTF14G1024	417	44215.9	7.17
PpTCP26	PprTF15G0431	318	35593.78	8.23
PpTCP27	PprTF15G0488	357	37949.04	6.09
PpTCP28	PprTF15G1133	197	21251.72	6.59
PpTCP29	PprTF16G0759	302	32790.68	9.19
PpTCP30	PprTF19G0500	417	44537.21	6.52
PpTCP31	PprTF19G0575	472	51782.41	7.41
PpTCP32	PprTF19G0791	395	42682.97	7.01
PpTCP33	PprTF001Sca0109	389	44677.01	8.03

Table 1c. The identification and character analysis of PpTCPs

Sequences and protein structure analysis of TCPs in Populus sect. Turanga

The amino acid sites of TCP domains in *PpTCP* and *PeTCP* proteins were analyzed using SMART to study the domain sequence characteristics of TCP proteins (*Fig. 2A, B*). The results showed that all TCP protein sequences have a highly conserved domain (bHLH) consisting of 63–65 amino acids. Analysis of the conserved domains of the two poplars found that the basic region was the most conserved, consisting of 20 amino acids. Among them, *P. pruinosa* contained seven conserved amino acid residues (2D, 3R, 6K, 10R, 12R, 13R, and 15R), while *P. euphratica* contained five (6K, 10R, 12R, 13R, and 15R) (*Fig. A1A, B*). In addition, the helix regions of TCP proteins were relatively conserved in the two poplars, and the hydrophobic amino acids were widely distributed.

Figure 1. The chromosomal locations of Populus sect. Turanga. (A) The chromosomal locations of TCPs in P. pruinosa. (B) The chromosomal locations of TCPs in P. euphratica. Graphical representation of locations for TCPs on each chromosome

Figure 2. Alignment of TCP domains from 33 PpTCPs. (A) Amino acids are expressed in the standard single letter code. The size of the letters at each position represents their frequency. Numbers in the horizontal axis indicate the position of amino acids. (B) The conserved motif of PpTCPs predicted by MEME.5.4.1 online tools (http://meme-suite.org/tools/meme). The conserved amino acids are indicated with colored box. Numbers in the vertical axis indicate the total number of amino acids in this TCP domains

The secondary structure prediction results showed that 33 PpTCP proteins were primarily random coil in *P. pruinosa*, accounting for 51.26% (*PpTCP15*)–81.92% (*PpTCP10*); followed by α helix and extend strand, accounting for 9.04% (*PpTCP10*)– 39.34% (*PpTCP9*) and 6.17% (*PpTCP21*)–17.10% (*PpTCP12*); β turn accounted for the lowest proportion, ranging from 0.55% (*PpTCP9*)–6.25% (*PpTCP16*). Except for *PpTCP2* and *PpTCP28*, in which the secondary structure was random coil > extend strand $>\alpha$ helix $>\beta$ turn, the secondary structure of the remaining 31 PpTCP proteins was random coil > α helix > extend strand > β turn (*Table 2a*). The secondary structure prediction results showed that 34 PeTCP proteins were primarily random coil in *P. euphratica*, accounting for 50.80% (*PeTCP32*)–77.69% (*PeTCP31*); followed by α helix and extend strand, accounting for 10.74% (*PeTCP31*)–37.50% (*PeTCP27*), and 5.96% (*PeTCP1*)–18.33% (*PpTCP29*); and β turn accounted for the lowest proportion, ranging from 0.53% (*PeTCP9*)–8.33% (*PeTCP5*) (*Table 2b*).

Protein	Alpha helix/%	Extended strand/%	Beta turn/%	Random coil/%
PpTCP1	17.55%	9.27%	4.64%	
PpTCP2	11.62%	12.88%	5.30%	70.20%
PpTCP3	21.56%	8.12%	5.62%	64.69%
PpTCP4	13.60%	10.26%	1.43%	74.70%
PpTCP5	16.28%	10.47%	2.91%	70.35%
PpTCP6	20.20%	9.27%	4.30%	66.23%
PpTCP7	18.53%	11.17%	3.81%	66.50%
PpTCP8	15.04%	12.08%	2.12%	70.76%
PpTCP9	39.34%	8.20%	0.55%	51.91%
PpTCP10	9.04%	6.85%	2.19%	81.92%
PpTCP11	15.38%	11.36%	3.15%	70.10%
PpTCP12	20.82%	17.10%	5.95%	56.13%
PpTCP13	26.90%	8.12%	4.06%	60.91%
PpTCP14	21.38%	13.77%	5.43%	59.42%
PpTCP15	36.48%	10.69%	1.57%	51.26%
PpTCP16	22.73%	12.50%	6.25%	58.52%
PpTCP17	29.55%	14.21%	4.91%	51.33%
PpTCP18	19.24%	10.38%	4.81%	65.57%
PpTCP19	14.85%	13.18%	2.72%	69.25%
PpTCP20	14.00%	10.57%	3.19%	72.24%
PpTCP21	36.76%	6.17%	1.03%	56.04%
PpTCP22	17.66%	13.77%	3.89%	64.67%
PpTCP23	15.83%	10.07%	2.64%	71.46%
PpTCP24	13.01%	10.69%	2.89%	73.41%
PpTCP25	14.16%	12.35%	3.92%	69.58%
PpTCP26	35.99%	6.17%	1.80%	56.04%
PpTCP27	13.53%	10.88%	4.24%	71.35%
PpTCP28	13.45%	14.85%	4.76%	66.95%
PpTCP29	24.50%	17.00%	5.00%	53.50%
PpTCP30	19.90%	12.23%	3.84%	64.03%
PpTCP31	14.74%	11.85%	3.76%	69.65%
PpTCP32	14.74%	11.85%	3.76%	69.65%
PpTCP33	32.65%	9.25%	1.03%	57.07%

Table 2a. Secondary structure of PpTCPs

APPLIED ECOLOGY AND ENVIRONMENTAL RESEARCH 21(2):1665-1696. http://www.aloki.hu ● ISSN 1589 1623 (Print) ● ISSN 1785 0037 (Online) DOI: http://dx.doi.org/10.15666/aeer/2102_16651696 © 2023, ALÖKI Kft., Budapest, Hungary

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Replication modes and positive selection analysis of PpTCPs and PeTCPs

Gene duplication events play important roles not only in genomic rearrangement and expansion but also in the diversification of gene functions, thereby implicating them as the primary driving forces throughout the evolutionary process of genomes, including the WGD (whole genome duplication)/segmental duplication, tandem and dispersed (Wu and Poethig, 2006). The replication events between *P. pruinosa* and *P. euphratica* were analyzed. We found that the replication patterns of *PpTCPs* genes include 24 WGD/segmental duplication and 9 dispersed (*Fig. 3A; Table 3a*). The *PeTCPs* were mainly generated by WGD/segmental duplication (24), followed by dispersed (8) and tandem duplication (2) (*Fig. 3B; Table 3b*).

Gene ID	Dup_type	PpTCP gene
PprTF01G0485	WGD or segmental	PpTCP1
PprTF01G0945	WGD or segmental	PpTCP2
PprTF01G2752	WGD or segmental	PpTCP3
PprTF01G3161	WGD or segmental	PpTCP4
PprTF02G1396	WGD or segmental	PpTCP5
PprTF03G1426	WGD or segmental	PpTCP6
PprTF04G0329	WGD or segmental	PpTCP7
PprTF04G0501	Dispersed	PpTCP8
PprTF04G0806	WGD or segmental	PpTCP9
PprTF04G0910	Dispersed	PpTCP10
PprTF04G1758	Dispersed	PpTCP11

Table 3a. The genes duplication modes of PpTCPs

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PprTF05G0677	WGD or segmental	PpTCP12
PprTF06G1062	Dispersed	PpTCP13
PprTF07G0637	WGD or segmental	PpTCP14
PprTF08G1035	WGD or segmental	PpTCP15
PprTF09G0110	Dispersed	PpTCP16
PprTF10G1116	WGD or segmental	PpTCP17
PprTF11G0334	WGD or segmental	PpTCP18
PprTF11G0540	Dispersed	PpTCP19
PprTF11G0622	Dispersed	PpTCP20
PprTF12G0077	WGD or segmental	PpTCP21
PprTF12G0730	WGD or segmental	PpTCP22
PprTF13G0866	WGD or segmental	PpTCP23
PprTF13G0942	WGD or segmental	PpTCP24
PprTF14G1024	WGD or segmental	PpTCP25
PprTF15G0431	WGD or segmental	PpTCP26
PprTF15G0488	Dispersed	PpTCP27
PprTF15G1133	WGD or segmental	PpTCP28
PprTF16G0759	Dispersed	PpTCP29
PprTF19G0500	WGD or segmental	PpTCP30
PprTF19G0575	WGD or segmental	PpTCP31
PprTF19G0791	WGD or segmental	PpTCP32
PprTF001Sca0109	WGD or segmental	PpTCP33

Table 3b. Secondary structure of PeTCPs

APPLIED ECOLOGY AND ENVIRONMENTAL RESEARCH 21(2):1665-1696. http://www.aloki.hu ● ISSN 1589 1623 (Print) ● ISSN 1785 0037 (Online) DOI: http://dx.doi.org/10.15666/aeer/2102_16651696 © 2023, ALÖKI Kft., Budapest, Hungary

Wang et al.: Genome-wide identification of TCP transcription factors family in *Populus* sect. *Turanga* (*Populus pruinosa* Schrenk and *Populus euphratica* Olive) reveal the roles of *TCPS* in leaf morphology

Ka/Ks ratio was an important indicator for the evaluation of selection pressure during sequence evolution (Hurst, 2002). We determined the Ka, Ks, and Ka/Ks ratios of *PpTCPs* and *PeTCPs* to explore the evolutionary history of *TCPs*. The results showed that among the 26 pairs of homologous genes, only *PpTCP19* (homologous to *PeTCP19*) and *PpTCP27* (homologous to *PeTCP28*) had a Ka/Ks ratio greater than 1, and the Ka/Ks ratios of the remaining 24 pairs of homologous genes were less than 1. This result indicated that most *PpTCPs* have undergone purification selection, except *PpTCP19* and *PpTCP27*; *PeTCP19 and PeTCP28*, which have undergone positive selection.

Figure 3. Replication modes of TCP gene family in Populus sect. Turanga. (A) Replication modes of TCP gene family in P. pruinosa. (B) Replication modes of TCP gene family in P. euphratica

Analysis of cis-acting elements in the promoter region of PpTCPs and PeTCPs

Plant CARE was used to analyze *cis*-acting elements in the upstream 2 kb promoter regions of *PpTCPs*/*PeTCPs* to explore the potential functions of *PpTCPs*/*PeTCPs*. The results showed that the promoter regions of *PpTCPs*/*PeTCPs* contained a large number of DNA sequence elements such as CAT-box, TC-rich repeats, ABRE, and TGAelement (*Fig. 4A, B*). These promoter elements play important roles in meristem expression, defense and stress responses, abscisic acid responses, as well as auxin responses. *PpTCP4*, *PpTCP18*, *PeTCP3*, *PeTCP17,* and *PeTCP18* contain TGAelement cis-acting elements. *PpTCP1*, *PpTCP3*, *PpTCP4*, *PpTCP8*, *PpTCP13*, *PpTCP14*, *PpTCP19*, *PpTCP23, PpTCP24*, *PpTCP26, PpTCP30*; *PeTCP1*, *PeTCP2*, *PeTCP3*, *PeTCP5*, *PeTCP19*, *PeTCP21*, *PeTCP25*, *PeTCP27,* and *PeTCP34* contain CAT-box *cis*-acting elements, thereby indicating that these *TCPs* may play important roles in leaf morphology.

Figure 4. Cis-acting elements identified in the promoters of TCPs in Populus sect. Turanga. (A) Cis-acting elements identified in the promoters of TCPs in P. pruinosa. (B) Cis-acting elements identified in the promoters of TCPs in P. euphratica. The micro-segments with different function annotations are represented by different colors. The scale bar represents 100 bp

Collinearity and phylogenetic analysis of interspecific TCP gene family

Herein, we performed a collinear analysis of *TCPs* between *P. pruinosa* and other species, such as *P. euphratica*, *Salix brachista,* and *Arabidopsis* to explore the potential evolutionary process of *PpTCPs* (*Fig. 5*). First, a total of 34, 30, and 24 *TCP*s members were identified in *P. euphratica*, *Salix brachista*, and *Arabidopsis*, respectively (*Table 4a, b*). Next, the results of collinearity analysis showed that there were 32, 30, and 24 *PpTCPs* have collinearity between *P. euphratica*, *Salix brachista,* and *Arabidopsis TCP*s, respectively (*Electronic Appendix 2*). Among them, 22 *PpTCPs* (*PpTCP3*, *PpTCP4*, *PpTCP5*, *PpTCP8*, *PpTCP9*, *PpTCP10*, *PpTCP11*, *PpTCP12*,

PpTCP13, *PpTCP14*, *PpTCP17*, *PpTCP19*, *PpTCP20*, *PpTCP21*, *PpTCP23*, *PpTCP25*, *PpTCP26*, *PpTCP27*, *PpTCP28,* and *PpTCP30*) were collinear with three other species, thereby suggesting that these genes may have existed before their ancestors diverged. There were only three *PpTCPs* have a collinear relationship with *PeTCP*, including *PpTCP24*, *PpTCP31*, and *PpTCP32*, thereby suggesting that these *PpTCPs* may play important roles in the long-term adaptive evolution of *P. pruinosa*.

Figure 5. Collinear analysis of TCPs between P. pruinosa and three representative plant species (P. euphratica, S. brachista and A. thaliana). The gray line in the background indicates collinear blocks within the P. euphratica, S. brachista and A. thaliana genomes, while the red lines highlight the syntenic TCP gene pairs

A phylogenetic tree of *TCP*s in *P. pruinosa* and *Arabidopsis*, *P. euphratica* and *Arabidopsis* was constructed to study the evolutionary relationship between *PpTCPs*/*PeTCPs* (*Fig. 6A, B*), referring to *TCP*s in *Arabidopsis*. The interspecific phylogenetic tree of *P. pruinosa* was divided into two subgroups based on the topological structure of the interspecific tree (classes Ⅰ and Ⅱ). *TCPs* from these two species showed dispersed distribution in both classes I and II, with *PpTCPs* containing 18 and 15 members, and *PeTCPs* containing 20, 14, and 2 members. Previous studies showed that the TCP of *Arabidopsis* classes I and II CIN branches were involved in the regulation of leaf morphology, including *AtTCP3*, *AtTCP4*, *AtTCP5*, *AtTCP7*, *AtTCP9*, *AtTCP10*, *AtTCP11*, *AtTCP13*, *AtTCP14*, *AtTCP15, AtTCP20*, *AtTCP21*, *AtTCP23,* and *AtTCP24* (Li et al., 2005; Schommer et al., 2008; Herve et al., 2009; Koyama and Tomotsugu, 2010; Viola et al., 2011; Kieffer et al., 2011). The number of *PpTCPs* in class II was relatively small, which was consistent with the classification of *Arabidopsis*. *PpTCP1*, *PpTCP2*, *PpTCP3*, *PpTCP5*, *PpTCP6*, *PpTCP7*, *PpTCP11*, *PpTCP12*, *PpTCP13*, *PpTCP14*, *PpTCP16*, *PpTCP18*, *PpTCP22*, *PpTCP23*, *PpTCP25*, *PpTCP28*, *PpTCP29,* and *PpTCP30* were clustered with class I subgroup in *Arabidopsis*. Meanwhile, *PpTCP9*, *PpTCP15*, *PpTCP17*, *PpTCP21*, *PpTCP26,* and *PpTCP33* were clustered with class II CYC/TB1 branches in *Arabidopsis*. *PeTCP1*, *PeTCP2,* and other 20 *PeTCPs* were clustered with class I in *Arabidopsis*, *PeTCP9*, *PeTCP32,* and other five *PeTCPs* were clustered with the class II CYC/TB1 branch in *Arabidopsis*. Given that *AtTCPs* with similar functions often show a preference to belong to a subgroup, 21 *PpTCPs* such as *PpTCP6* and 25 *PeTCPs* such as *PeTCP1* may be involved in the regulation of leaf morphology in *P. pruinosa* and *P. euphratica*.

Table 4b. The identification of TCPs in A. thaliana

We performed an orthologous alignment between *PpTCPs*/*PeTCPs* and *AtTCPs* to explore the potential functions of *PpTCPs*/*PeTCPs* and found that 12 *PpTCPs* (*PpTCP3*, *PpTCP4*, *PpTCP5*, *PpTCP7*, *PpTCP10*, *PpTCP11*, *PpTCP12*, *PpTCP13*, *PpTCP19*, *PpTCP23*, *PpTCP27,* and *PpTCP28*) and 15 *PeTCPs* (*PeTCP2*, *PeTCP3*, *PeTCP7*, *PeTCP9*, *PeTCP10*, *PeTCP11*, *PeTCP12*, *PeTCP13*, *PeTCP16*, *PeTCP19*, *PeTCP23*, *PeTCP25*, *PeTCP26*, *PeTCP28,* and *PeTCP29*) were orthologous genes in *Arabidopsis*. These results supported the reliability of the evolutionary tree of *TCPs* between the *Populus* sect. *Turanga* and *Arabidopsis*.

Expression patterns of TCPs in different leaf morphology in Populus sect. Turanga

The gene expression levels of *PpTCPs/PeTCPs* were analyzed by RNA-Seq sequencing to explore the expression patterns of *PpTCPs/PeTCPs* in different leaf morphology. After removing low-quality reads, a total of 396,962,572 clean reads were obtained in *P. pruinosa*. The percentages of Q30 and GC were 91.7%–94.63% and 43.6%–45.6%, respectively. Meanwhile, a total of 681,315,674 clean reads were obtained in *P. euphratica*. The percentages of Q30 and GC were 93.49%–95.41% and

43.3%–44.8%, respectively. This indicated that the quality of the transcriptome sequencing data of *P. pruinosa* and *P. euphratica* was high enough for subsequent analysis.

Figure 6. A neighbor-joining phylogenetic tree of Populus sect. Turanga and A. thaliana. (A) A neighbor-joining phylogenetic tree of P. pruinosa and A. thaliana. (B) A neighbor-joining phylogenetic tree of P. euphratica and A. thaliana

The analysis of the expression patterns of *TCPs* in the *Populus* sect. *Turanga* showed that the expression levels of *PpTCP9*, *PpTCP15*, and *PpTCP27* (homologous to *AtTCP5*) in the oblong and round leaves were lower than broad ovate, and five *PpTCP*s expression levels in the round leaves were higher than that in the oblong and broad ovate leaves, such as *PpTCP12* (homologous to *AtTCP7*), *PpTCP23* (homologous to *AtTCP23*) (*Fig. 7C; Table 5a*). In *P. euphratica*, the expressions of 11 *PeTCPs*, such as *PeTCP11* (homologous to *AtTCP8*) and *PeTCP28* (homologous to *AtTCP5*), expression levels in broad ovate leaves were higher than that in oblong and ovate leaves (*Fig. 7D; Table 5b*). A total of 11 *PpTCPs* contained CAT-box, of which seven *PpTCPs* expression levels were upregulated in oblong leaves (e.g., *PpTCP1*, *PpTCP3*, *PpTCP8*, *PpTCP13*, *PpTCP14*, *PpTCP19,* and *PpTCP30*). In addition, 16 *PpTCPs* contained TC-rich repeats in *cis*-acting elements, six *PpTCPs* (*PpTCP3*, *PpTCP6*, *PpTCP8*, *PpTCP10, PpTCP13,* and *PpTCP20*) expression levels were upregulated in oblong leaves, and three *PpTCPs* (*PpTCP12*, *PpTCP22,* and *PpTCP31*) were upregulated in round leaves; 18 *PpTCPs* contained ABRE *cis*-acting elements, of which seven *PpTCPs* (*PpTCP1*, *PpTCP2*, *PpTCP6*, *PpTCP10*, *PpTCP16*, *PpTCP17,* and *PpTCP18*) expression levels were upregulated in oblong leaves. Moreover, two *PpTCPs* (*PpTCP21* and *PpTCP23*) expression levels were upregulated in round leaves, and two *PpTCPs* (*PpTCP9* and *PpTCP15*) expression levels were upregulated in broad ovate leaves (*Electronic Appendix 3a*). Furthermore, nine *PeTCPs* contained CAT-box, among which four *PeTCPs* expression levels were upregulated in broad ovate leaves (for example, *PeTCP1*, *PeTCP5*, *PeTCP25,* and *PeTCP34*). A total of 12 *PeTCPs* contained TC-rich repeats *cis*acting elements, of which two *PeTCPs* (*PeTCP2* and *PeTCP6*) expression levels were upregulated in linear leaves, three *PeTCPs* (*PeTCP20*, *PeTCP23,* and *PeTCP24*) expression levels were upregulated in broad ovate leaves, and 20 *PeTCPs* (*PeTCP20*, *PeTCP23,* and *PeTCP24*) contain ABRE *cis*-acting elements, of which two *PeTCPs* (*PeTCP6* and *PpTCP31*) expression levels were upregulated in linear leaves, one *PeTCP* (*PeTCP13*) expression level was upregulated in ovate leaves, and 10 *PeTCPs* (*PeTCP1*, *PeTCP4*, *PeTCP5*, *PeTCP7*, *PeTCP11*, *PeTCP14*, *PeTCP23*, *PeTCP24*, *PeTCP25,* and *PeTCP32*) expression levels were upregulated in broad ovate leaves (*Electronic Appendix 3b*). Among them, there are CAT-box and TC-rich repeats *cis*-acting elements in the promoters of *PpTCP8*, *PpTCP13,* and *PeTCP2*, ABRE and TGA-element *cis*-acting elements in the promoters of *PpTCP18*, CAT-box, and ABRE *cis*-acting elements in the promoters of *PpTCP1*, *PeTCP1*, *PeTCP5,* and *PeTCP25*. These results indicate that *PpTCPs*/*PeTCPs* have dynamic changes in the process of poplar leaf morphology. Moreover, *PpTCPs*/*PeTCPs* may be involved in the regulation of leaf morphology in the *Populus* sect. *Turanga*. In addition, some *PpTCPs* and *PeTCPs* lack expression information (e.g., *PpTCP4*, *PpTCP11,* and *PpTCP22*, etc.), which possibly indicates that these are pseudogenes or are expressed only under special conditions.

Table 5a. Expression of PpTCPs in three leaf shapes of P. pruinosa

PpTCP gene	Gene ID in genome	Pp LOF 1	Pp LOF 2	Pp LOF 3	Pp CF 1	Pp CF 2	Pp CF 3	Pp BOF 1	Pp BOF 2	Pp BOF 3
PpTCP1	PprTF01G0485	0.496527	0.304383	0.238924	0.573882	0.468835	0.368809	0.209621	1.355704	0.746733
PpTCP2	PprTF01G0945	8.100595	6.407289	13.83313	7.914284	8.822757	12.46217	8.703259	11.98129	18.01558
PpTCP3	PprTF01G2752	32.65648	14.62331	25.06645	24,00908	23.99753	26.20363	19.93702	23.94489	14.41347
PpTCP4	PprTF01G3161	118.0275	96.23908	70.7172	100.1984	81.42838	58.97735	121.8302	109.8492	105.9065
PpTCP5	PprTF02G1396	4.266036	9.185717	9.50161	1.274154	3.985363	3.827708	1.123666	0.604428	2.979659
PpTCP6	PprTF03G1426	7.628499	6.828698	9.582638	6.723079	6.656298	9.007969	6.579758	7.501992	9.439015
PpTCP7	PprTF04G0329	38.84488	38.96642	26.79684	34.76294	35.04028	21.74104	28.20439	19.6759	14.56901
PpTCP8	PprTF04G0501	43.30738	40.90691	53.88897	30.38738	31.20461	40.94036	28.85948	31.23158	36.62663

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Table 5b. Expression of PeTCPs in four leaf shapes of P. euphratica

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Figure 7. Expression profiles of TCP genes in the Populus sect. Turanga. (A) Photographs of leaf shape in P. pruinosa. (B) Photographs of leaf shape in P. euphratica. (C) Expression profiles of TCP genes in the P. pruinosa base on the transcriptome data. (D) Expression profiles of TCP genes in the P. euphratica base on the transcriptome data. The colour bar represents the normalized FPKM values as follows: red, high expression level; blue, low expression level; and white, no expression. Detail FPKM values is listed in Tables 5a, b

Discussion

P. pruinosa and *P. euphratica* as two sister plant species with morphologically well differentiated provide a good model for elucidating physiological and molecular mechanisms of tolerance in tree species. Plant leaves, as the main organ for photosynthesis and respiration, show continuous and discrete morphological changes during stage transitions: leaf size shows continuous changes, while several different shapes can be seen at different plant developmental stages (Wang and Wang, 1989). In addition to differences in types of habitat, they also occur in different shapes of leaves between *P. pruinosa* and *P. euphratica*. *P. pruinosa* shows three kinds of heteromorphic leaves, including oblong, round, and broad leaves, while *P. euphratica* have four kinds, which are linear, lanceolate, ovate, and broad ovate leaves. The TCP transcription factors play important roles in plant growth and development (Viola et al., 2011; Martín-Trillo and Cubas, 2010), and have been identified in various plants (Yao et al., 2007; Wei et al., 2016; Chen et al., 2016). However, the members of *TCPs* of *P.*

pruinosa remain unclear, and the comparative investigation of *TCPs* between *P. pruinosa* and its relatives has not been carried out.

In this study, we performed a comprehensive analysis of *PpTCPs*/*PeTCPs* using chromosome-level *P. pruinosa*/*P. euphratica* genome data, thereby identifying 33 and 34 *PpTCPs* and *PeTCPs*, respectively. Gene duplication events, such as tetraploidization (whole-genome duplication, WGD) and hexaploidization (wholegenome triplication, WGT), occur frequently in plants and are the major sources of evolutionary changes that enable rapid adaptation to different environments (Chen et al., 2016). Furthermore, WDG/segmental duplications are the main drivers of the expansion of *PeTCPs.* Identification and collinearity analysis of multispecies *TCPs* show that *TCPs* among Salicaceae species exhibit stronger collinearity as compared with *Arabidopsis*. Therefore, we speculate that the WGD event unique to Salicaceae causes the expansion of *PeTCPs* after differentiation from *Arabidopsis*.

Phylogenetic analysis and sequence alignment show that the genome of *P. euphratica* contains all the classes of *TCP*s, namely classes I and II. The alignment of these multiple sequences supports the classification of the phylogenetic tree, which is consistent with the *TCP* subclasses reported in plants, thereby indicating that this gene family is relatively conserved in plants and may have similar functions (Lin et al., 2016). Transcription factors from classes I and II are involved in the regulation of various growth and developmental processes in *Arabidopsis*. *AtTCP2*, *AtTCP3*, *AtTCP4*, *AtTCP10,* and *AtTCP24* were targeted by miR319 and involved in regulating leaf morphology (Palatnik et al., 2003). In addition, *AtTCP4* can regulate leaf margin serrations by affecting auxin distribution through the miR164-CUC pathway (Schommer et al., 2014), and its over-expression will lead to reduced cell division in *Arabidopsis*, thereby leading to smaller leaf size (Sarvepalli and Nath, 2011). In this study, *PpTCP19*, the orthologous gene of *AtTCP2*, is highly expressed in oblong leaves of *P. pruinosa*, and linear and lanceolate leaves of *P. euphratica*. Therefore, we speculate that *PpTCP19* and *PeTCP19* may affect the leaf morphology and "narrow leaves" (oblong, linear, and lanceolate) in the *Populus* sect. *Turanga* by participating in the regulation of leaf cell division. In addition, Ka/Ks analysis found that *PpTCP19* was a positively selected gene, and we speculated that the emergence of the oblong leaves of *P. pruinosa* may be the result of adaptive evolution.

The *AtTCP14* mutants show that *AtTCP14* is strongly expressed at the boundaries of leaves and sepals, wherein the cell division is highly restricted (Kieffer et al., 2011). The orthologous genes of *AtTCP14* in *P. pruinosa* and *P. euphratica* are *PpTCP7* and *PeTCP7*, respectively, which are lowly expressed in the broad ovate leaves of *P. pruinosa* and *P. euphratica*, respectively. Therefore, we speculate that *PpTCP7* and *PeTCP7* may participate in the regulation of "broad ovate leaves" of *P. euphratica* and *P. pruinosa* by inhibiting cell proliferation.

Conclusions

A total of 33 and 34 *PpTCPs* and *PeTCPs* were respectively identified based on the chromosome level genome of *P. pruinosa* and *P. euphratica,* and they were unevenly distributed on 19 chromosomes of *P. pruinosa* and *P. euphratica*. The main driving force of *TCPs* expansion in the *Populus* sect. *Turanga* was the WGD/fragmental duplication event. In addition, *PpTCP7*, *PpTCP19*, *PeTCP7,* and *PeTCP19* may play important roles in the leaf morphology of the *Populus* sect. *Turanga*. Overall, this study

is a valuable resource for the functional characterization of *TCPs* in the *Populus* sect. *Turanga* and lay further understanding of the structure-function relationship among these TCP members. Our study also provides comprehensive information and novel insights into the roles of TCP family genes in the regulation and heteromorphic leaves.

Availability of data and materials. The datasets supporting the results of *P. pruinosa* and *P. euphratica* in this article are available at the National Center for Biotechnology Information (*P. pruinosa*) and National Genome Data Center (*P. euphratica*) (NCBI, https://www.ncbi.nlm.nih.gov; NGDC, https://ngdc.cncb.ac.cn/) under project accession number PRJNA890019 and PRJCA005959.

Competing interests. The authors declared that they had no conflict of interests.

Funding. This work was supported by the Bintuan Science and Technology Program (grant number 2021BB010), the Xinjiang Production & Construction Corps Key Laboratory of Protection and Utilization of Biological Resources in Tarim Basin (grant number BRZD2003) and the Tarim University Natural Science Research Conditions Construction Project (grant number TDZKKY202202) and the Graduate Research Innovation Project of the Xinjiang Uygur Autonomous Region (XJ2020G268).

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APPENDIX
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Figure A1. Conserved domains of TCP gene family in P. euphratica. (A) Amino acids are expressed in the standard single letter code. The size of the letters at each position represents their frequency. Numbers in the horizontal axis indicate the position of amino acids. (B) The conserved motif of PeTCPs predicted by MEME.5.4.1 online tools (http://memesuite.org/tools/meme). The conserved amino acids are indicated with colored box. Numbers in the vertical axis indicate the total number of amino acids in this TCP domains

ELECTRONIC APPENDICES

Electronic Appendix 1. The identification and character analysis of PeTCPs Electronic Appendix 2. The collinearity analysis of TCPs between P. pruinosa and other three plants (P. euphratica, S. brachista and A. thaliana)

Electronic Appendix 3a. The cis-element analysis of the PpTCPs

Electronic Appendix 3b. The cis-element analysis of the PeTCPs