THE RELATIONSHIP BETWEEN SALT-AFFECTED AGRICULTURAL SOILS AND THE POTENTIAL ROLE OF EARTHWORMS IN RESTORATION: A REVIEW

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Abstract. Agricultural soil salinization has negative impacts on food production, environmental health and socio-economic wellbeing. The restoration of salt-affected agricultural soils focuses not only on the mediation of physicochemical habitats for plant establishment and growth but also on biological habitats for ecological functions. Earthworms are recognized to be essential components of soil biota but have been neglected to a great extent in the restoration practice of salt-affected agricultural land. Here we review current restoration of salt-affected agricultural soils, how soil salinity affects earthworms, how earthworm may contribute to the restoration of salt-affected agricultural soils and how earthworm populations may be improved in salt-affected agricultural soils. The major points are the following (1) The restoration of salt-affected agricultural soils focused on not only the "bioindicators" of earthworms but also the "facilitators" of it, which may contribute to the sustainable improvement of restored soils. (2) Soil salinity decreased biomass, abundance and diversity of earthworms in different degree depending on earthworm categories and salt ion composition. (3) Earthworm could improve soil properties (e.g. soil salt content, pH and physical structure), which would contribute to the restoration of the salt-affected agricultural soils. (4) The application of high-quality organic residues enhances establishment and development of earthworm population in salt-affected agricultural soils, which are conducive to implementation of the role of earthworms.

Keywords: *ecological restoration, ecological engineer, soil salinity, organic residues application*

Introduction

Salt-affected soils are generally defined as that electrical conductivity of soil is greater than 4 dS/m. Soil salinization arising from either natural or human-induced causes leads to an increase in concentration of dissolved salts in the soil profile. Moreover, the growing world population add to the competition for fresh water and global warming had risen soil evaporation and rising seawater level may further exacerbate soil salinization in some coastal regions (Rozema and Flowers, 2008; Eckelmann et al., 2006; Wang and Li, 2012; Tsanis et al., 2016). More than 3% of total soil resources are currently salt-affected (FAO, 2020), which figure will continue growing at a rate of up to 2 Mha yr^{-1} (Singh, 2018). Currently, about 33% of irrigated agricultural land is already affected by salinization as a result of poor agricultural practices. It is estimated that 50% of the croplands worldwide will become salinized by

2050 (Nachshon, 2018). The soil salinization has set serious risks to long-term agricultural sustainability (Pulido-Bosch et al., 2018). In salt-affected agricultural soils, salts (mainly Na⁺ ions) result in the dispersion of clay particles leading directly to the breakup of aggregates and to poor soil structure (*Fig. 1*) (Tejada and Gonzalez, 2006; Choudhary et al., 2006), which limits water infiltration and salt leaching (Gharaibeh et al., 2010; Yu et al., 2010). The formation of salt-affected agricultural soils is considered the main factor leading to soil degradation (*Fig. 1*) which impairs food production, environmental health and socio-economic wellbeing (Rozema and Flowers, 2008; Ondrasek et al., 2011). Salinization effects on agriculture including decreased productivity, an increased risk for less tolerant species, and the loss of biodiversity (Church et al., 2013).

Figure 1. Conceptual graph describing soil salinization change soil physical chemical and biological properties and consequently affect soil function productivity and environment. Earthworm applied into salt-affected soils improve soil structure leaching of salt and soil organism activity and consequently accelerate salt-affected soil restoration

Soil organisms are essential to functioning of the soil ecosystems because they participate in key ecological processes like organic decomposition, nutrient cycling and maintenance of soil structure (Briones, 2014; Cohan et al., 2022). Soil fauna in various terrestrial ecosystems have received considerable attention in recent years due to their role in sustainable ecosystem functioning and services (Snyder and Hendrix, 2008; Blouin et al., 2013; van Groenigen et al., 2014; Bertrand et al., 2015). Earthworms are essential components of the fauna in temperate climates and are commonly regarded as ecosystem engineers (Lavelle et al., 1997). There is growing interest in how earthworms contribute to the improvement of the physical chemical and biological properties of the soil (*Fig. 1*) (Baker et al., 2006; Eriksen-Hamel and Whalen, 2007; De Oliverira et al., 2012) and the increase of plant production (Fonte et al., 2023). What roles earthworms might play in the ecological restoration of various degraded soils (Snyder and Hendrix, 2008; Butt, 2008; Boyer and Wratten, 2010; McDaniel et al., 2013; Duarte et al., 2014). The potential of earthworms to restore ecosystem function in salt-affected agricultural soils by improving soil and plant growth is an important future research topic.

In this review we focus on the restoration of salt-affected agricultural soils and the interaction between soil salinization and earthworms. This review first discusses recent

research trends in the field of salt-affected land restoration. Second, we discuss the effect of soil salinization on earthworm abundance, biomass and ecological categories. We then discuss how the roles of earthworms in soils would contribute to the restoration of salt-affected agricultural soil and improve plant productivity. Finally, we also discuss how to increase earthworm abundance and biomass in salt-affected agricultural soil.

Restoration of salt-affected agricultural soils

Soil salinization causes a series of ecological and environmental issues such as soil deterioration rapid nutrient cycling and vegetation destruction which need to be addressed. Therefore, environmental criteria for restoration success after soil salinization are traditionally related to improvements in soil chemistry nutrient availability and crop yield (Tejada et al., 2006). Improvements to soil physical and chemical properties are essential for the restoration of most degraded agricultural land ecosystems and soil organisms can be manipulated to increase the speed of restoration (Harris, 2009). In earlier research most soil organism communities as "bioindicators" are gradually used to indicate the restoration state of the ecosystem (Callaham et al., 2008) and are usually well studied as "facilitators" in the recent decade especially soil fauna (Snyder and Hendrix, 2008; Boyer and Wratten, 2010). Attempts to restore other aspects of soil ecosystem function cannot succeed without a functioning soil ecological community (Heneghan et al., 2008). Therefore, increasing populations or addition of groups of soil fauna may be necessary to accelerate the restoration of soil ecosystem function in the restoration soils (Snyder and Hendrix, 2008) and in agricultural and reclaimed soils (Baker et al., 2006; Boyer and Wratten, 2010).

In salt-affected agricultural soils, one of the main soil environmental issues is the destruction of the soil structure which regulates water infiltration percolation and retention and material transfer (Romero-Aranda et al., 2001; Choudhary et al., 2006; Bottinelli et al., 2015). Moreover, improvements in porosity and aggregation can increase the soil's capacity to retain moisture which reduces soil osmotic stress and consequently provides favorable conditions for the proliferation of plant roots and the activity of soil microorganisms (Paranychianakisa et al., 2004; Mavi and Marschner, 2012). For this reason, studies of salinized soil restoration are common (Tejada et al., 2006; Clark et al., 2007; Lakhdar et al., 2009) and often have the main aim of restoring soil structure (Snyder and Hendrix, 2008). Soil fauna are critical to the formation and stabilization of soil aggregates; they also create burrows and macropores that greatly facilitate the movement of water in the soil (Bottinelli et al., 2015). Many studies point to the importance of single dominant keystone organisms in determining soil aggregation and pore structure (Bastardie et al., 2003; Capowiez et al., 2012; Bottinelli et al., 2017). Additionally, many methods are used for the remediation of salt-affected agricultural soils which can improve soil physical and chemical properties and crop growth (Raychev et al., 2001; Rietz and Haynes, 2003; Tao et al., 2014; Tao et al., 2019). Moreover, the further improvement and sustainability of salt-affected soils may be obtained by increasing key soil organism population and diversity such as earthworms. It is well recognized that earthworms play an important role in mediating soil processes and functioning (Edwards, 2004; Blouin et al., 2013) by acting as soil engineers in soils (Jones et al., 1994; Lavelle et al., 1997). They can increase the decomposition and mineralization of organic residues by comminution incorporation and gut passage. Their burrowing and mixing activity results in the formation of macropores and aggregations influencing soil aeration and movement of water (Brown et al., 2000).

Interaction between salt-affected soil properties and earthworm ecotype

The impact of soil salinity on earthworms

Salt-affected agricultural soils in which salt concentrations are normally considered safe for many plants are known to have detrimental effects on soil earthworms (Owojori et al., 2009). Salinity as an individual parameter negatively affected earthworm survival and led to changes in earthworm individual biomass (McDaniel et al., 2013). The effects of soil salinity on earthworms could be attributed to the direct effects of the osmotic potential and the indirect effects of the food resources of earthworms (plantderived materials and microorganisms) on earthworm abundance and biomass (Yuan et al., 2007; Wichern et al., 2006). A previous study in laboratory experiments showed that soil salinity (EC of 3.35 dS m⁻¹) decreased the dry biomass of earthworms (*Aporrectodea trapezoides)* and that soil salinity (EC of 7.35 dS m-1) had adverse effects on the survival of earthworms in a salt-affected agricultural soil (Tao et al., 2012). In grassland, soil salinity (EC of 1.05 dS m-1) decreased the abundance and biomass of earthworms under field conditions (Owojori and Reinecke, 2010). These results indicated that the changes of earthworm biomass were a more sensitive measure than the number of earthworms in the assessment of detrimental effects of soil salinity on earthworms in the short term. Additionally, soil salinity (EC of 0.52 dS m⁻¹) could reduce not only earthworm abundance and but also diversity of earthworm communities in meadows of the delta in the long term (Ivask et al., 2012).

Soil salinity also affected cocoon production. The study of Owojori et al. (2008) found that the salt content influencing cocoon production is lower than that influencing earthworm growth. Owojori et al. (2009) suggested that soil salinity caused a greater influence on earthworm survival than on earthworm reproduction at same salinity levels. A previous study showed that soil salinity (EC of 5.26 and 7.35 dS m^{-1}) had a significant effect on earthworm (*Aporrectodea trapezoides*) reproduction and caused the total cessation of reproduction in a coastal salt-affected agricultural soil (Tao et al., 2012). These results suggested that earthworm cocoon production of earthworms may be more sensitive to salinity than earthworm survival and biomass change of earthworms in the salt-affected agricultural soils (Raiesi et al., 2020). These results reinforce the idea that the increase in soil salinization likely led to a delayed effects on earthworm abundance (Pereira et al., 2019), which suggested a potential long-term impairment of the soil functions mediated by them. However, cocoons are commonly considered salt-resistant as well as drought- and stress-resistant and these characteristics maybe help to maintain earthworm survival with the decreasing of soil salinity (Holmstrup, 2001).

The impact of salt ion composition on earthworms

The effects of salinity on earthworms may be dependent on soil salt ion composition (Riet and Haynes, 2003; Owojori and Reinecke, 2010; Chen et al., 2017). In most studies on salinity the salt content of substrates is believed to be the main predictor of effects on earthworms but it is often measured without taking the soil salt composition into account (Owojori and Reinecke, 2014). In most studies on salinity the electrical

conductivity (EC) of substrates is believed to be the main predictor of effects on soil organisms. However some studies have shown that salinity as measured by EC could not be used to correctly predict the effects of salt on soil organisms when comparing different saline areas (Riet and Haynes, 2003; Mavi et al., 2012). This limitation may be due to differences in anion and cation composition between different salt-affected agricultural soils. In saline soils from specific areas Ca was the dominant cation (Riet and Haynes, 2003). Ca salts led to obviously higher decreases in earthworm (*Eisenia fetida*) survival and weight than Na salts (Owojori and Reinecke, 2014). In greenhouse soil excessive water and fertilizer applications caused soil salinity with $NO₃$ (Darwish et al., 2005; Shi et al., 2009) which may result in different effects on earthworms than soil salinity with Cl (Owojori and Reinecke, 2014). In future studies it should be considered how salt ion composition affects earthworm parameters.

The responses of earthworm ecological categories to soil salinity

Epigeic earthworms live close to the soil surface (topsoil); endogeic earthworms inhabit the deeper layers and consume large amounts of mineral soil; anecic earthworms feed on large organic material at the soil surface and inhabit the subsoil (Edward and Bohlen, 1996). Thus, the effects of soil salinization on earthworms would depend on ecological categories (Ivask et al., 2012). With respect to salt effects and distribution epigeic species may be more sensitive to the effects of salinity accumulating in the surface soil while endogeic species which feed and live within the soil would be less sensitive to the effects of surface salinity and anecic species might experience intermediate impacts. Epigeic earthworms were more tolerant to the limiting effects of salinity accumulated in the surface layer compared to other earthworm ecological categories (Ivask et al., 2007, 2012). The epigeic species *Dendrobaena octaedra Eiseniella tetraedra Dendrodrilus rubidus* and *Lumbricus castaneus* were present in subsaline and saline coastal zones (Keplin and Broll, 2010). However, a few studies suggested that epigeic earthworms (*Eisenia fetida*) appeared to be more sensitive responses to salinity than endogeic earthworms since the survival of epigeic earthworms was seriously affected (Owojori and Reinecke, 2009; Owojori et al., 2009) whereas no significant effect on survival was found for *Aporrectodea caliginosa* at this salinity level in natural saline soil (Owojori et al., 2009). However, the abundance of the endogeic species *Aporrectodea caliginosa, Aporrectodea rosea* and *Octolasion lacteum* were negatively affected by soil salinity in the coastal salt-affected agricultural soil of the Baltic Sea (Ivask et al., 2012). The growth of endogeic earthworms (*A. caliginosa*) was significantly affected by a lower salinity than the value that affected the growth of epigeic earthworms. These results suggest that the community tolerance of earthworms to soil salinity also depends on its ecological categories (Van Leeuwen and Vermeire, 2007; Pereira et al., 2019).

The role of earthworms in the restoration of salt-affected agricultural soils

Impact on soil salinity

The main objective of salt-affected soils is to reduce soil salinity, which alleviated the effects of it on plant growth, soil organism activity and soil structure. As some of the most important soil-dwelling organisms earthworms play a significant role in soil structure and water holding capacity improvement through their burrowing and casting

activities (Görres et al., 2001; Capowiez et al., 2012; Bottinelli et al., 2017), which can influence the movement of soil water-salt. Salt in salt-affected agricultural soils leaches into deeper soil depths as earthworm activities improve the soil structure (Zhang et al., 2015). Earthworm casts also significantly altered the soil structure, which facilitate the leaching of salt ions, reducing the salt content of the topsoils (Li et al., 2021). The application of earthworms (*Eudrilus eugeniae*) combined with organic material (compost and vermicompost) could reduce soil salinity which may be because that the increasing Ca^{2+} and Mg^{2+} concentration in the soil solution permitted greater leaching of exchanged Na⁺ in percolating water (Oo et al., 2015). The studies of Zhang et al. (2016, 2018) also showed that epigeic earthworms (*Eisenia fetida*) could decrease soil salt concentration through the improvement of soil structure and water movement drainage in saline alkali soil. A previous study also showed that the presence of endogeic earthworms significantly increased leaching of salt from salt-affected agricultural soils under rainfall conditions (Wang et al., 2016). They speculated that earthworm burrowing activity created channels and improve the leaching of saline water (*Fig. 2*). The anecic species tend to inhabit semi-permanent vertical burrows in the soil which may have strong effects on soil water-salt movement. Different earthworm ecological categories have distinct burrow networks and consequently result to different patterns of soil water-salt movement. Moreover, the interactions between different ecological groups could alter the movement of solutes (Shuster et al., 2002; Ernst et al., 2009) and thereby create distinct patterns in water-salt transport.

Figure 2. Conceptual graph describing earthworm activities (casting and burrowing) promote the leaching of water and salt from salt-affected agricultural soil (photographs by Tao Jun). Additionally, earthworm decrease the water content of the upper soil layer (earthworm mainly live in this soil layer). Consequently, this process may also affect upward movement of water and salt from the subsoil layer without earthworms

In addition, changes in soil structure due to earthworm burrowing and casting activity affected not only soil water infiltration and solution transfer but also soil evaporation and the upward movement of the soil solution (*Fig. 2*). The study of Lipiec et al. (2015) demonstrated that pore size distribution was different between earthworm casts and the surrounding soil. They found that the median and average pore radius were greater in earthworm casts than in the surrounding soil aggregates. The results of a

previous study indicated that soil porosity was significantly decreased in earthworm casts compared to that in the surrounding soil aggregates (Jouquet et al., 2008). These differences may contribute to the significantly higher salt content in casts under evaporation than in the surrounding soils (Wang et al., 2016). The burrowing activities of anecic and endogeic earthworm species might result in stronger aeration of the topsoil. Consequently, larger biopores in the soil increased the gas exchange and thus increased the loss of water vapor from the soil (Ernst et al., 2009). A previous study indicated that earthworms decreased the water content of salt-affected agricultural soils under evaporation conditions (Wang et al., 2016). However, the study of Li et al. (2021) found that Earthworm casts increased the total porosity and capillary porosity of the soil, which likely increased the soil's water-holding capacity and lowered evaporation. These soil water processes may also affect upward movement of salts in salt-affected agricultural soil (*Fig. 2*). Therefore, at present the related systematic research on the effects of earthworms on salt distribution under evaporation conditions is rather limited.

Impact on soil pH

Soil pH has a considerable impact on controlling plant nutrients particularly the availability of micronutrients (Grattan and Grieve, 1992). Saline soil pH is significantly increased by salt leaching in salt-affected agricultural soil (Murtaza et al., 2006; Choudhary et al., 2006; Wang et al., 2012). These studies showed that the soil desalinization process of salt leaching by watering could cause soil alkalinization problems. Although most studies found that the pH in earthworm casts is higher than that in the surrounding soil (Versteegh et al., 2014; Vos et al., 2014) the pH in earthworm casts is lower than in the surrounding soils when the original pH of the soils is higher than 7 (Van Groenigen et al., 2019). A few studies have indicated that earthworms have the potential to decrease soil pH in salt-affected agricultural soils (Oo et al., 2015; Zhang et al., 2016, 2018). The effect of earthworms might be a result of the production of various organic acids by their gut microbes and the pH-homeostasis mechanisms of their gut tissue (Ravindran et al., 2014). These observations indicated that the presence of earthworms may mitigate negative salinity-pH relationships and thus reduce the risk of soil secondary alkalization.

Impact on plant growth

Soil salinization is one of the major causes of yield reduction in modern agriculture (Rozema and Flowers, 2008). Salt accumulation in the root zone affects plant performance directly through the development of osmotic potential and the disruption of cell ion homeostasis (Zhu, 2001; Munns, 2002) and indirectly through the limiting soil nutrient availability in the rhizosphere (Farooq et al., 2015). Earthworms are thought to have positive effects on plant growth in salt-affected soils (Zhang et al., 2016, 2018). The positive effects of earthworms on plant growth may be due to several possible mechanisms; these mechanisms included the enhancement of nutrient availability the improvement of soil structure the production of plant growth-regulating substances and the stimulation of microbial plant symbionts (Fonte et al., 2023). The first mechanism was firstly mentioned in previous studies. Especially in the salt-affected soils the decreases in the size and activity of soil microorganisms had a negative effect on microbiological processes and consequently impacted organic matter decomposition and nutrient availability (Qadir and Schuber, 2002; Yuan et al., 2007; Yan and

Marschner, 2012). Earthworms can consume organic substrates and release nutrients into soil through metabolic processes and consequently affect soil nutrient cycling (Zhang et al., 2016). They found that earthworms significantly increased the abundance of *Pontibacter* in saline soil that contributed to the promotion of available P which was a contributing factor for plant growth (Dasager et al., 2011). Earthworms also increased the abundances of Trichoderma (plant-promoted fungi) in the saline soils (Zhang et al., 2016) which enhanced nitrogen uptake and promoted crop growth (Tanwar et al., 2013). Additionally, Oo et al. (2015) and Zhang et al. (2016) showed that earthworms enhanced soil microbial biomass catalase activity and basal respiration in salt-affected soils through their gut processes (Svensson and Friberg, 2007) which may increase total soil nutrient turnover and prevent crop from oxidative damages under salinity conditions. In salt-affected soils earthworm could affect plant pathogens (decreasing *Methylobacterium* abundance) and consequently promote crop growth as well in nonsaline soils (Zhang et al., 2016).

In the salt-affected agricultural soils, organic residues can serve as food for earthworms (Tao et al., 2012; Chen et al., 2017) and earthworms can increase crop production by increasing N mineralization (Zhang et al., 2016). Earthworms could significantly increase nutrient and mineral elements uptake into plant shoot in saltaffected soils to improve crop salt tolerance which promote crop growth under saline stress (Rabie and Almadini, 2005; Shahzad et al., 2012). For example, an increase in the uptake of exchangeable K and Ca (in competition with sodium) by earthworms could increase crop biomass in saline soil (Zhang et al., 2018). Earthworm could improve crop photosynthetic and osmoprotectants by increase the Mg and N content of shoot in saltaffected soils which may promote crop growth under saline conditions (Anjum et al., 2014; Porcel et al., 2015). Additionally, the beneficial effects of earthworms on plant growth may be due to the restoration of demolished soil structure destroyed by salinity. Some previous studies suggested that the presence of earthworms increased plant biomass in the disturbed soils compared to that in the undisturbed soils (Spurgeon et al., 2013; Van Groenigen et al., 2014). In salt-affected soils earthworms could increase soil macroaggregates proportions, stability of aggregates and macropores numbers under mesocosm experiment which contributed to enhancement of crop growth (Kou et al., 2023). However, it may be difficult to distinguish the effects of the improved soil structure from those of increased nutrient availability when earthworms are present in the salt-affected agricultural soils. Apart from the previously mentioned mechanisms the effects of earthworms on plant growth in salt-affected agricultural soil may be partly attributed to the reductions in the root zone soil salt concentration which is leached downward to deeper soil layers (Kou et al., 2023).

Enhancement of earthworm populations in salt-affected agricultural soils

Earthworms that are able to recolonize or enhance in a salt-affected agricultural soil are important for soil restoration. Numerous studies have shown accelerated the recolonization and enhancement of earthworms following organic matter amendments of base restoration soil (Lowe and Butt, 2002; Hurisso et al., 2011; Wu et al., 2013). As discussed in the previous studies (Eijsackers, 2011) an important factor for earthworm recolonization and population growth in these various degraded soil habitats is soil organic carbon which enables earthworm survival and growth. However, in salt-affected arable land the amounts of organic carbon in the soil tend to decrease which does not

support earthworm population growth. Various organic residues (e.g., plant residues, manures, biosolids and composts) have been applied to salt-affected agricultural soil to maintain soil organic carbon and improve soil physical chemical and microbial properties (Tejada et al., 2006; Liang et al., 2005; Lakhdar et al., 2009). The extent to which earthworm population growth and species diversity increases in a salt-effected agricultural soil as well as in other restored soils would be likely to depend on agricultural management practices such as organic matter application (Wu et al., 2001). The study of Qiao et al. (2001) concluded that management practices (mainly the input of organic material) which gradually reduces the salt concentrations are conditions suitable for earthworm recolonization and population growth in field.

The application of organic residues increased the growth of endogeic earthworms in salt-affected agricultural soil and significantly promoted their reproduction in the soils with the low salinity (Tao et al., 2012). Therefore, they explained that the amendment with organic residues might improve soil quality and food sources for earthworms including the available substances and microbial biomass. In addition, the quality and composition of organic residues applied to the soil influences soil organic carbon and consequently benefits earthworms. The biochemical composition of organic residues such as the carbon to nitrogen ratio of residues could be considered as a qualitative indicator of the food resources supplied to earthworms (Cesarz et al., 2016). The study of Chen et al. (2017) suggested that the input of low C/N residues (clover plants) into salt-affected agricultural soil increased earthworm growth and reproduction compared to that under high C/N residues (wheat plants) in the coastal region of Bohai Bay in China. The results of this experiment are in agreement with those of a previous study (Fraser and Piercy, 1998) in which amendment with clover residues significantly increased earthworm population. This may be because that the low C/N residue application has a significantly higher decomposition rate than the high C/N residues. On the other hand, the decomposition of low C/N residues could produce a higher concentration and longer lasting source of water-extractable organic C (Hasbullah and Marschner, 2015) which can be easily taken up by soil microbes and earthworms (Hasbullah and Marschner, 2016). Moreover, the higher N concentration in residues could improve the production of osmolytes (such as proline and glycine betaine) by providing N for their synthesis (Saum and Müller, 2007) which consequently supported earthworm growth and reproduction (Whalen and Parmelee, 1999; Curry and Schmidt, 2007). The decomposition of high C/N residues requiring greater energy for synthesis and release of enzymes (Uchida et al., 2012) is slow in a salt-affected agricultural soil (Setia and Marschner, 2013). In field conditions, the study of Chen et al. (2022) also observed a higher earthworm biomass in salt-affected agricultural soil with clover residues than that in the same soils without clover residues. Moreover, the latest research suggested that animal residue applied into a salt-affected agricultural soil significantly enhanced earthworm (endogeic species) population and biomass compared to plant residues in mesocosms of field (Chen et al., 2022). They explained that the application animal residue obtained a higher soil available resources and nutrients than clover residues (Nguyen et al., 2018; Naveed et al., 2021), which contribute to the increase in earthworm growth in the salt-affected soils. This result showed that to promote field survival and biomass of earthworms in a salt-affected agricultural soil, animal manure application may be a critical important amendment.

In addition to organic residue inputs for the benefit of earthworms, soil moisture is also considered to be a key environmental factor for determining earthworm numbers

growth reproduction and activity (Holmstrup, 2001; Perreault and Whalen, 2006) especially in salt-affected agricultural soils (Tao et al., 2013). Soil moisture conditions can change the response of earthworm growth and reproduction to soil salinity (Ivask et al., 2012). High soil moisture content contributed to higher earthworm biomass and numbers (Eriksen-Hamel and Whalen, 2006). For example, endogeic species preferred high moisture conditions and were present in high numbers in floodplains (Zorn et al., 2008). High soil moisture content (95% field capacity) significantly increased the dry weight and numbers of earthworm compared to the low soil moisture content (75% field capacity) in a salt-affected agricultural soil in a laboratory experiment (Chen et al., 2017). This result may have occurred for several reasons (1) the decrease in the osmotic potential of soil solution (Setia and Marchner, 2013) could have benefited earthworm growth; (2) the increase in transport of soluble substrates (Poll et al., 2008) contributed to higher earthworm consumption and assimilation; or (3) other available organic matter increased earthworm growth (Leroy et al., 2008; Butt, 2011). Additionally, the high soil moisture content increased the number of cocoons compared to those at a low moisture content in a salt-affected agricultural soil (Chen et al., 2017). This result indicated that high soil moisture content promoted earthworm reproduction, which might affect earthworm population over a period of time.

Moreover, the increase in earthworm populations as a restoration agent requires careful consideration of the location of the restoration project. Restoration practitioners must ask themselves "Whether there are earthworms in the restoration region", "Which earthworm species should be supported?" Meanwhile restoration goals for earthworm populations must take into account the location's native earthworm species. A previous study found that the earthworm *Aporrectodea trapezoides* appears to be the dominant species in salt-affected agricultural soils in the coastal region of China's Bohai Bay (Tao et al., 2013) and has the greatest potential for colonization of the salt-affected agricultural soils in this region (Tao et al., 2012). Further research should focus on the potential enhancement of native earthworm species in salt-affected agricultural sites (Lavelle et al., 1999).

Conclusions

Earthworms are a key functional group of soil organisms that are affected by soil salinity depending on their ecological categories and stages of development in saltaffected agricultural soils. In the salt-affected soils much previous research has focused on the changes of earthworm communities as bio-indicators of soil improvement. Although this function as an indicator is related to their potential use in restoration it is not strictly an ecosystem service. Future research should focus on understanding the use of earthworms to modify soil function and provide ecosystem services in the saltaffected agricultural soils. For example, the study of the relationship between earthworm burrows and water-salt movement would help to explain soil salt leaching and upward movement and the resulting distribution of salt in the soil profile. These soil processes are important for plant growth and successful ecosystem restoration in saltaffected lands. In addition, the colonization or recovery of earthworm population numbers and communities is the foundation for restoration of soil and the reestablishment of ecosystem function. Therefore, future research should therefore advance more management practices (e.g., various organic residue applications) that increase earthworm populations and growth in the salt-affected agricultural soil systems

or that help earthworm return to the disturbed lands because the presence of earthworms stimulates plant production and will help to narrow the productivity gap between saltaffected agricultural soils and non-salt-affected agricultural soils.

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REFERENCES

- [1] Anjum, N. A., Aref, I. M., Duarte, A. C., Pereira, E., Ahmad, I., Iqbal, M. (2014): Glutathione and proline can coordinately make plants withstand the joint attack of metal (loid) and salinity stresses. – Frontiers in Plant Science 5: 662. https://doi.org/10.3389/fpls.2014.00662.
- [2] Baker, G. H., Brown, G., Butt, K., Curry, J., Scullion, P. J. (2006): Introduced earthworms in agricultural and reclaimed land, their ecology and influences on soil properties, plant production and other soil biota. – Biology Invasions 8: 1301-1316. https://doi.org/10.1007/s10530-006-9024-6.
- [3] Bastardie, F., Capowiez, Y., de Dreuzy, J. R., Cluzeau, D. (2003): X-ray tomographic and hydraulic characterization of burrowing by three earthworm species in repacked soil cores. – Applied Soil Ecology 24: 3-16. https://doi.org/10.1016/s0929-1393(03)00071-4.
- [4] Bertrand, M., Barot, S., Blouin, M., Whalen, J., de Oliveira, T., Estrade, J. R. (2015): Earthworm services for cropping systems. A review. – Agronomy for Sustainable Development 35: 553-567. https://doi.org/10.1007/s13593-014-0269-7.
- [5] Blouin, M., Hodson, M. E., Delgado, E. A., Baker, G., Brussaard, L., Butt, K. R., Dai, J., Dendooven, L., Peres, G., Tondoh, J. E., Cluzeau, D., Brun, J. J. (2013): A review of earthworm impact on soil function and ecosystem services. – European Journal of Soil Science 64: 161-182. https://doi.org/10.1111/ejss.12025.
- [6] Bottinelli, N., Jouquet, P., Capowiez, Y., Podwojewski, P., Grimaldi, M., Peng, X. (2015): Why is the influence of soil macrofauna on soil structure only considered by soil ecologists? – Soil Tillage Research 146: 118-124. https://doi.org/10.1016/j.still.2014.01.007.
- [7] Bottinelli, N., Zhou, H., Capowiez, Y., Zhang, Z. B., Qiu, J., Jouquet, P., Peng, X. H. (2017): Earthworm burrowing activity of two non-Lumbricidae earthworm species incubated in soils with contrasting organic carbon content (Vertisol vs. Ultisol). – Biology and Fertility of Soils 53: 951-955. https://doi.org/org/10.1007/s00374-017-1235- 8.
- [8] Boyer, S., Wratten, S. D. (2010): The potential of earthworms to restore ecosystem services after opencast mining—a review. – Basic and Applied Ecology 11: 196-203. https://doi.org/10.1016/j.baae.2009.12.005.
- [9] Briones, M. J. I. (2014): Soil fauna and soil functions: a jigsaw puzzle. Frontiers in Environmental Science 22. https://doi.org/10.3389/fenvs.2014.00007.
- [10] Brown, G. G., Barois, I., Lavelle, P. (2000): Regulation of soil organic matter dynamics and microbial activity in the drilosphere and the role of interactions with other edaphic functional domains. – European Journal of Soil Biology 36: 177-198. https://doi.org/10.1016/S1164-5563(00)01062-1.
- [11] Butt, K. R. (2008): Earthworms in soil amelioration, Lessons learned from United Kingdom case studies of land reclamation. – Restoration Ecology 16: 637-641. https://doi.org/10.1111/j.1526-100X.2008.00483.x.
- [12] Butt, K. R. (2011): Food quality affects production of Lumbricus terrestris (L.) under controlled environmental conditions. – Soil Biology and Biochemistry 43: 2169-2175. https://doi.org/10.1016/j.soilbio.2011.06.021.
- [13] Callaham, M. A., Rhoades, C. C., Heneghan, L. (2008): A striking profile, soil ecological knowledge in amelioration management and science. – Restoration Ecology 16: 604-607. https://doi.org/org/10.1111/j.1526-100X.2008.00490.x.
- [14] Capowiez, Y., Stéphane, C., Pierre, B., Jean, R. E., Guy, R., Hubert, B. (2009): Experimental evidence for the role of earthworms in compacted soil regeneration based on field observations and results from a semi-field experiment. – Soil Biology and Biochemistry 41: 711-717. https://doi.org/10.1016/j.soilbio.2009.01.006.
- [15] Capowiez, Y., Stéphane, S., Stéphane, C., Pierre, B., Guy, R., Hubert, B. (2012): Role of earthworms in regenerating soil structure after compaction in reduced tillage systems. – Soil Biology and Biochemistry 55: 93-103. https://doi.org/10.1016/j.soilbio.2012.06.013.
- [16] Cesarz, S., Craven, D., Dietrich, C., Eisenhauer, N. (2016): Effects of soil and leaf litter quality on the biomass of two endogeic earthworm species. – European Journal of Soil Biology 77: 9-16. http://doi.org/10.1016/j.ejsobi.2016.09.002.
- [17] Chen, J., Gu, W., Tao, J., Xu, Y. J., Wang, Y., Gu, J. Y. (2017): The effects of organic residue quality on growth and reproduction of Aporrectodea trapezoides under different moisture conditions in a salt-affected agricultural soil. – Biology and Fertility of Soils 53: 103-113. https://doi.org/10.1007/s00374-016-1158-9.
- [18] Chen, J., Tao, J., Zhang, H., Gu, W. (2022): Effects of residue types and plastic mulch on earthworm Aporrectodea trapezoides (Duges, 1828) within mesocosms at a salt-affected soil. – Archives of Agronomy and Soil Science 69(7): 1055-1070. https://doi.org/10.1080/03650340.2022.2052050.
- [19] Choudhary, O. P., Ghuman, B. S., Josan, A. S., Bajwa, M. S. (2006): Effect of alternating irrigation with sodic and non-sodic waters on soil properties and sunflower yield. – Agricultural Water Management 85: 151-156. https://doi.org/10.1016/j.agwat.2006.03.017.
- [20] Church, J. A. et al. (2013): Sea level change. In Climate Change 2013: The Physical Science Basis. – In: Stocker, T. F. et al. (eds.) Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, UK, New York, NY.
- [21] Clark, G. J., Dodgshun, N., Sale, P. W. G., Tang, C. (2007): Changes in chemical and biological properties of a sodic clay subsoil with addition of organic amendments. – Soil Biology and Biochemistry 39: 2807-2817. https://doi.org/10.1016/j.soilbio.2007.06.003.
- [22] Coban, O., De Deyn, G.B., van der Ploeg, M. (2022): Soil microbiota as game-changers in restoration of degraded lands. – Science 375: 990.
- [23] Curry, J. P., Schmidt, O. (2007): The feeding ecology of earthworms—a review. Pedobiologia 50: 463-477. https://doi.org/10.1016/j.pedobi.2006.09.001.
- [24] Darwish, T., Atallah, T., Moujabber, M. E., Khatib, N. (2005): Salinity evolution and crop response to secondary soil salinity in two agro-climatic zones in Lebanon. – Agricultural Water Management 78: 152-164. https://doi.org/10.1016/j.agwat.2005.04.020.
- [25] Dastager, S. G., Deepa, C. K., Pandey, A. (2011): Plant growth promoting potential of Pontibacter niistensis in cowpea (Vigna unguiculata (L) Walp.). – Applied Soil Ecology 49: 250-255. https://doi.org/10.1016/j.apsoil.2011.04.016.
- [26] De Oliveira, T., Bertrand, M., Estrade, J. R. (2012): Short-term effects of ploughing on the abundance and dynamics of two endogeic earthworm species in organic cropping systems in northern France. – Soil Tillage Research 119: 76-84. https://doi.org/10.1016/j.still.2011.12.008.
- [27] Duarte, A. P., Melo, V. F., Brown, G. G., Pauletti, V. (2014): Earthworm (Pontoscolex corethrurus) survival and impacts on properties of soils from a lead mining site in

Southern Brazil. – Biology and Fertility of Soils 50: 851-860. https://doi.org/10.1007/s00374-014-0906-y.

- [28] Eckelmann, W., Baritz, R., Bialousz, S., Bielek, P., Carre, F., Houskova, B., Jones, R. J. A., Kibblewhite, M. G., Kozak, J., Le Bas, C., Tóth, G., Tóth, T., Várallyay, G., Yli Halla, M., Zupan, M. (2006): Common Criteria for Risk Area Identification According to Soil Threats. – European Soil Bureau Research Report no. 20. Office for Official Publications of the European Communities, Luxembourg.
- [29] Edwards, C. A. (2004): Earthworm ecology. Second Ed. CRC Press, Boca Raton.
- [30] Edwards, C. A., Bohlen, P. J. (1996): Biology and Ecology of Earthworms. Chapman and Hall, London.
- [31] Eijsackers, H. (2011): Earthworms as colonizers of natural and cultivated soil environments. – Applied Soil Ecology 50: 1-13. https://doi.org/10.1016/j.apsoil.2011.07.008.
- [32] Eriksen-Hamel, N. S., Whalen, J. K. (2007): Impacts of earthworms on soil nutrients and plant growth in soybean and maize agroecosystems. – Agriculture Ecosystems and Environment 120: 442-448. https://doi.org/10.1016/j.agee.2006.11.004.
- [33] Ernst, G., Felten, D., Vohland, M., Emmerling, C. (2009): Impact of ecologically different earthworm species on soil water characteristics. – European Journal of Soil Biology 45: 207-213. https://doi.org/10.1016/j.ejsobi.2009.01.001.
- [34] FAO (2020): Management of Salt Affected Soils: 'Soil Management' under 'FAO SOILS PORTAL'. – Food and Agriculture Organization of the United Nations, Rome. http://www.fao.org/soils-portal/soil-management/management-of-some-problemsoils/salt-affected-soils/more-information-on-salt-affected-soils/en/ (accessed 9 April 2020).
- [35] Farooq, M., Hussain, M., Wakeel, A., Siddique, K. H. (2015): Salt stress in maize, effects, resistance mechanisms, and management. A review. – Agronomy for Sustainable Development 35: 461-481. https://doi.org/10.1007/s13593-015-0287-0.
- [36] Fonte, S. J., Hsieh, M., Mueller, N. D. (2023): Earthworms contribute significantly to global food production. – Nature Communications 14: 5713.
- [37] Fraser, P. M., Piercy, J. E. (1998): The effects of cereal straw management practices on lumbricid earthworm populations. – Applied Soil Ecology 9: 369-373. https://doi.org/10.1016/S0929-1393(98)00091-2.
- [38] Gharaibeh, M., Eltaif, N., Shra'ah, S. (2010): Reclamation of a calcareous saline sodic soil using phosphoric acid and by product gypsum. – Soil Use and Management 26: 141- 148. https://doi.org/org/10.1111/j.1475-2743.2010.00260.x.
- [39] Görres, J. H., Savin, M. C., Amador, J. A. (2001): Soil micropore structure and carbon mineralization in burrows and casts of an anecic earthworm (Lumbricus terrestris). – Soil Biology and Biochemistry 33: 1881-1887. https://doi.org/10.1016/S0038-0717(01)00068- \mathfrak{D}
- [40] Grattan, S. R., Grieve, C. M. (1992): Mineral element acquisition and growth response of plants in saline environments. – Agriculture, Ecosystems Environment 38: 275-300. https://doi.org/10.1016/0167-8809(92)90151-Z.
- [41] Harris, J. (2009): Soil microbial communities and amelioration ecology, facilitators or followers? – Science 325: 573. https://doi.org/10.1126/science.1172975.
- [42] Hasbullah, H., Marschner, P. (2015): Residue properties influence the impact of salinity on soil respiration. – Biology and Fertility of Soils 51: 99-111. https://doi.org/10.1007/s00374-014-0955-2.
- [43] Hasbullah, H., Marschner, P. (2016): Multiple additions of rapidly decomposable residue alleviate the negative impact of salinity on microbial activity. – Soil Research 54: 692- 699. https://doi.org/10.1071/SR15103.
- [44] Heneghan, L., Miller, S. P., Baer, S., Callaham, M. A., Montgomery, J., Pavao-Zucherman, M., Rhoades, C. C., Richardson, S. (2008): Integrating soil ecological

knowledge into amelioration management. – Restoration Ecology 16(4): 608-617. https://doi.org/org/10.1111/j.1526-100X.2008.00477.x.

- [45] Holmstrup, M. (2001): Sensitivity of life history parameters in the earthworm Aporrectodea caliginosa to small changes in soil water potential. – Soil Biology and Biochemistry 33: 1217-1223. https://doi.org/10.1016/s0038-0717(01)00026-8.
- [46] Hurisso, T. T., Davis, J. G., Brummer, J. E., Stromberger, M. E., Stonaker, F. H., Kondratieff, B. C., Booher, M. R., Goldhamer, D. A. (2011): Earthworm abundance and species composition in organic forage production systems of northern Colorado receiving different soil amendments. – Applied Soil Ecology 48: 219-226. https://doi.org/10.1016/j.apsoil.2011.03.003.
- [47] Ivask, M., Truu, J., Kuu, A., Truu, M., Leito, A. (2007): Earthworm communities of flooded grasslands in Matsalu, Estonia. – European Journal of Soil Biology 43: 71-76. https://doi.org/10.1016/j.ejsobi.2006.09.009.
- [48] Ivask, M., Meriste, M., Kuu, A., Kutti, S., Sizov, E. (2012): Effect of flooding by fresh and brackish water on earthworm communities along Matsalu Bay and the Kasari River. – European Journal of Soil Biology 53: 11-15. https://doi.org/10.1016/j.ejsobi.2012.08.001.
- [49] Jones, C. G., Lawton, J. H., Shachak, M. (1994): Organisms as ecosystem engineers. Oikos 69: 373-386. https://doi.org/10.2307/3545850.
- [50] Jouquet, P., Bottinelli, N., Podwojewski, P., Hallaire, V., Thu, T. D., Duc, T. T. (2008): Chemical and physical properties of earthworm casts as compared to bulk soil under a range of different land-use systems in Vietnam. – Geoderma 146: 231-238. https://doi.org/10.1016/j.geoderma.2008.05.030.
- [51] Keplin, B., Broll, G. (2010): Earthworm Coenoses in Wet Grassland of Northwest-Germany. Effects of Restoration Management on a Histosol and a Gleysol. – In: Broll, G., Merbach, W., Pfeiffer, E.-M. (eds.) Wetlands in Central Europe. Soil Organisms, Soil Ecological Processes and Trace Gas Emission, Springer, Berlin, pp. 11-34.
- [52] Kou, X. C., Chen, J., Tao, Y., Tao, J. (2023): Soil structure shifts with earthworms under different organic fertilization in salt-affected soils. – Land Degradation & Development 1-9. https://doi.org/10.1002/idr.4966.
- [53] Lakhdar, A., Rabhi, M., Ghnaya, T., Montemurro, F., Jedidi, N., Abdelly, C. (2009): Effectiveness of compost use in salt-affected agricultural soil. – Journal of Hazard Mater 171(1-3): 29-37. https://doi.org/10.1016/j.jhazmat.2009.05.132.
- [54] Lavelle, P., Bignell, D., Lepage, M. (1997): Soil function in a changing world, The role of invertebrate ecosystem engineers. – European Journal of Soil Biology 33: 159-193.
- [55] Lavelle, P., Brussaard, L., Hendrix, P. (1999): Earthworm Management in Tropical Agroecosystems. – CABI, New York.
- [56] Leroy, B. L. M., Schmidt, O., Van den Bossche, A., Reheul, D., Moens, M. (2008): Earthworm population dynamics as influenced by the quality of exogenous organic matter. – Pedobiologia 52: 139-150. https://doi.org/10.1016/j.pedobi.2008.07.001.
- [57] Li, Y.P., Wang, J., Shao, M. A. (2021): Effects of earthworms casts on water and salt movement in typical Loess Plateau soils under brackish water irrigation. – Agricultural Water Management 252: 106930.
- [58] Liang, Y. C., Si, J., Nikolic, M., Peng, Y., Chen, Y., Chen, W., Jiang, Y. (2005): Organic manure stimulates biological activity and barley growth in soil subject to secondary salinization. – Soil Biology and Biochemistry 37: 1185-1195. https://doi.org/10.1016/j.soilbio.2004.11.017.
- [59] Lipiec, J., Turski, M., Hajnos, M., Świeboda, R. (2015): Pore structure, stability and water repellency of earthworm casts and natural aggregates in loess soil. – Geoderma 243-244: 124-129. https://doi.org/10.1016/j.geoderma.2014.12.026.
- [60] Lowe, C. N., Butt, K. R. (2002): Influence of organic matter on earthworm production and behaviour, a laboratory-based approach with applications for soil amelioration. –

http://www.aloki.hu ● ISSN 1589 1623 (Print) ● ISSN 1785 0037 (Online)

DOI: http://dx.doi.org/10.15666/aeer/2202_12471265

European Journal of Soil Biology 38: 173-176. https://doi.org/10.1016/s1164- 5563(02)01141-x.

- [61] Mavi, M. S., Marschner, P. (2012): Drying and wetting in saline and saline-sodic soils effects on microbial activity, biomass and dissolved organic carbon. – Plant Soil 355: 51- 62. https://doi.org/10.1007/s11104-011-1078-2.
- [62] Mavi, M. S., Marschner, P., Chittleborough, D. J., Cox, J. W., Sanderman, J. (2012): Salinity and sodicity affect soil respiration and dissolved organic matter dynamics differentially in soils varying in texture. – Soil Biology and Biochemistry 45: -13. https://doi.org/10.1016/j.soilbio.2011.10.003.
- [63] McDaniel, J. P., Stromberger, M. E., Barbarick, K. A., Cranshaw, W. (2013): Survival of Aporrectodea caliginosa and its effects on nutrient availability in biosolids amended soil. – Applied Soil Ecology 7: 11-6. https://doi.org/10.1016/j.apsoil.2013.04.010.
- [64] Munns, R. (2002): Comparative physiology of salt and water stress. Plant Cell Environment 25: 39-250.
- [65] Murtaza, G., Ghafoor, A., Qadir, M. (2006): Irrigation and soil management strategies for using saline-sodic water in a cotton-wheat rotation. – Agricultural Water Management 81: 8-114. https://doi.org/10.1046/j.0016-8025.2001.00808.x.
- [66] Nachshon, U. (2018): Cropland soil salinization and associated hydrology: trends, processes and examples. – Water 10: 1030.
- [67] Naveed, M., Ditta, A., Ahmad, M., Mustafa, A., Ahmad, Z., Conde-Cid, M., Tahir, S., Shah, S. A. A., Abrar, M. M., Fahad, S. (2021): Processed animal manure improves morpho-physiological and biochemical characteristics of Brassica napus L. under nickel and salinity stress. – Environ Science and Pollution Research 28: 45629-45645.
- [68] Nguen, B. T., Trinh, N. N., Thi Le, C. M., Nguyen, T. T., Tran, T. V., Thai, B. V., Le, T. V. (2018): The interactive effects of biochar and cow manure on rice growth and selected properties of salt-affected soil. – Arch. Agron. Soil Sci. 64: 1744-1758.
- [69] Ondrasek, G., Rengel, Z., Veres, S. (2011): Soil Salinisation and Salt Stress in Crop Production. – In: Shanker, A., Venkateswarlu, B. (eds.) Abiotic Stress in Plants, Mechanisms and Adaptations. – InTech, London, pp. 171-190.
- [70] Oo, A., Iwai, C., Saenjan, P. (2015): Soil properties and maize growth in saline and nonsaline soils using cassava-industrial waste compost and vermicompost with or without earthworms. – Land Degradation Development 26: 300-310. https://doi.org/ 10.1002/ldr.2208.
- [71] Owojori, O. J., Reinecke, A. J. (2009): Avoidance behaviour of two eco-physiologically different earthworms (Eisenia fetida and Aporrectodea caliginosa) in natural and artificial saline soils. – Chemosphere 75: 279-283. https://doi.org/10.1016/j.chemosphere.2008.12.051.
- [72] Owojori, O. J., Reinecke, A. J. (2010): Effects of natural (flooding and drought) and anthropogenic (copper and salinity) stressors on the earthworm Aporrectodea caliginosa under field conditions. – Applied Soil Ecology 44: 156-163. https://doi.org/10.1016/j.apsoil.2009.11.006.
- [73] Owojori, O. J., Reinecke, A. J. (2014): Differences in ionic properties of salts affect saline toxicity to the earthworm Eisenia fetida. – Applied Soil Ecology 83: 247-252. https://doi.org/10.1016/j.apsoil.2013.05.019.
- [74] Owojori, O. J., Reinecke, A. J., Rozanov, A. (2008): Effects of salinity on partitioning , uptake and toxicity of zinc in the earthworm Eisenia fetida. – Soil Biology and Biochemistry 40: 2385-2393. https://doi.org/10.1016/j.soilbio.2008.05.019.
- [75] Owojori, O. J., Reinecke, A. J., Voua-Otomo, P., Reinecke, S. A. (2009): Comparative study of the effects of salinity on life-cycle parameters of four soil dwelling species (Folsomia candida, Enchytraeus doerjesi, Eisenia fetida and Aporrectodea caliginosa). – Pedobiologia 52: 351-360. https://doi.org/10.1016/j.pedobi.2008.12.002.
- [76] Paranychianakis, N. V., Aggelides, S., Angelakis, A. N. (2004): Influence of rootstock, irrigation level and recycled water on the growth and yield of Soultanina grapevines. – Agricultural Water Management 69: 13-27. https://doi.org/10.1016/j.agwat.2004.03.012.
- [77] Pereira, C. S., Lopes, I., Abrantes, I., Sousa, J. P., Chelinho, S. 2019 Salinization effects on coastal ecosystems: a terrestrial model ecosystem approach. – Phil. Trans. R. Soc. B 374: 20180251. http://dx.doi.org/10.1098/rstb.2018.0251.
- [78] Perreault, J. M., Whalen, J. K. (2006): Earthworm burrowing in laboratory microcosms as influenced by soil temperature and moisture. – Pedobiologia 50: 397-403. https://doi.org/10.1016/j.pedobi.2006.07.003.
- [79] Porcel, R., Redondo-Gómez, S., Mateos-Naranjo, E., Aroca, R., Garcia, R., Ruiz-Lozano, J. M. (2015): Arbuscular mycorrhizal symbiosis ameliorates the optimum quantum yield of photosystem II and reduces non-photochemical quenching in rice plants subjected to salt stress. – Journal of Plant Physiology 185: 75-83. https://doi.org/10.1016/j.jplph.2015.07.006.
- [80] Pulido-Bosch, A., Rigol-Sanchez, J. P., Vallejos, A., Andreu, J. M., Ceron, J. C., Molina-Sanchez, L., Sola, F. (2018): Impacts of agricultural irrigation on groundwater salinity. – Environment Earth Science 77: 197.
- [81] Qadir, M., Schubert, S. (2002): Degradation processes and nutrient constraints in sodic soils. – Land Degradation & Development 13: 275-294. https://doi.org/10.1002/ldr.504.
- [82] Qiao, Y. H., Wu, W. L. (2001): Relationship between production input and secondary succession of earthworm population in salinity transforming region of North China- A case study in Quzhou County. – Chinese Journal of Applied Ecology 12: 414-416.
- [83] Rabie, G., Almadini, A. (2005): Role of bioinoculants in development of salt-tolerance of Vicia faba plants under salinity stress. – African Journal of Biotechnology 4: 210. https://doi.org/10.1016/j.jbiotec.2004.09.017.
- [84] Raiesi, F., Motaghian, R. H., Nazarizadeh, M. (2020): The sublethal lead (Pb) toxicity to the earthworm Eisenia fetida (Annelida, Oligochaeta) as affected by NaCl salinity and manure addition in a calcareous clay loam soil during an indoor mesocosm experiment. – Ecotoxicology and Environmental Safety 190: 110083. https://doi.org/10.1016/j.ecoenv.2019.110083.
- [85] Ravindran, B., Contreras-Ramos, S. M., Wong, J. W. C., Selvam, A., Sekaran, G. (2014): Nutrient and enzymatic changes of hydrolysed tannery solid waste treated with epigeic earthworm Eudrilus eugeniae and phytotoxicity assessment on selected commercial crops. – Environmental Science and Pollution Research 21: 641-651. crops. – Environmental Science and Pollution Research 21: 641-651. https://doi.org/10.1007/s11356-013-1897-1.
- [86] Raychev, T., Popandova, S., Józefaciuk, G., Hajnos, M., Sokoowska, Z. (2001): Physicochemical reclamation of saline soils using coal powder. – International Agrophysics 15: 51-54.
- [87] Rietz, D. N., Haynes, R. J. (2003): Effects of irrigation-induced salinity and sodicity on soil microbial activity. – Soil Biology and Biochemistry 35: 845-854. https://doi.org/10.1016/s0038-0717(03)00125-1.
- [88] Romero-Aranda, R., Soria, T., Cuartero, J. (2001): Tomato plant–water uptake and plant– water relationships under saline growth conditions. – Plant Science 160: 265-272. https://doi.org/10.1016/s0168-9452(00)00388-5.
- [89] Rozema, J., Flowers, T. (2008): Ecology, Crops for a Salinized World. Science 322: 1478-1480. https://doi.org/10.1126/science.1168572.
- [90] Saum, S. H., Müller, V. (2007): Salinity-dependent switching of osmolyte strategies in a moderately halophilic bacterium, Glutamate induces proline biosynthesis in Halobacillus halophilus. – Journal of Bacteriology 189: 6968-6975. https://doi.org/10.1128/JB.00775- 07.
- [91] Setia, R., Marschner, P. (2013): Carbon mineralization in saline soils as affected by residue composition and water potential. – Biology and Fertility of Soils 49: 71-77. https://doi.org/10.1007/s00374-013-0797-3.
- [92] Shahzad, M., Witzel, K., Zörb, C., Mühling, K. (2012): Growth-related changes in subcellular ion patterns in maize leaves (Zea mays L.) under salt stress. – Journal of Agronomy and Crop Science 198(1): 46-56. https://doi.org/10.1111/j.1439- 037X.2011.00487.x.
- [93] Shi, W. M., Yao, J., Yan, F. (2009): Vegetable cultivation under greenhouse conditions leads to rapid accumulation of nutrients, acidification and salinity of soils and groundwater contamination in South-Eastern China. – Nutrient Cycling in Agroecosystems 83: 73-84. https://doi.org/10.1007/s10705-008-9201-3.
- [94] Shuster, W. D., Subler, S., McCoy, E. L. (2002): The influence of earthworm community structure on the distribution and movement of solutes in a chisel-tilled soil. – Applied Soil Ecology 21: 159-167. https://doi.org/10.1016/s0929-1393(02)00063-x.
- [95] Singh, A. (2018): Managing the environmental problems of irrigated agriculture through the appraisal of groundwater recharge. – Ecological Indication 92: 388-393.
- [96] Snyder, B. A., Hendrix, P. F. (2008): Current and potential roles of soil macroinvertebrates (earthworms, millipedes, and isopods) in ecological amelioration. – Restoration Ecology 16: 629-636. https://doi.org/10.1111/j.1526-100X.2008.00484.x.
- [97] Spurgeon, D. J., Keith, A. M., Schmidt, O., Lammertsma, D. R., Faber, J. H. (2013): Land use and land-management change, relationships with earthworm and fungi communities and soil structural properties. – BMC Ecology 13: 46. https://doi.org/10.1186/1472-6785-13-46.
- [98] Svensson, K., Friberg, H. (2007): Changes in active microbial biomass by earthworms and grass amendments in agricultural soil. – Biology and Fertility of Soils 44: 223-228. https://doi.org/10.1007/s00374-007-0200-3.
- [99] Tanwar, A., Aggarwal, A., Kaushish, S., Chauhan, S. (2013): Interactive effect of AM fungi with Trichoderma viride and Pseudomonas fluorescens on growth and yield of broccoli. – Plant Protection Science 49: 137-145. https://doi.org/10.1016/j.biortech.2013.05.002.
- [100] Tao, J., Gu, W., Griffiths, B., Liu, X. J., Xu, Y. J., Zhang, H. (2012): Maize residue application reduces negative effects of soil salinity on the growth and reproduction of the earthworm Aporrectodea trapezoides, in a soil mesocosm experiment. – Soil Biology and Biochemistry 49: 46-51. https://doi.org/10.1016/j.soilbio.2012.02.010.
- [101] Tao, J., Wu, L. H., Liu, X. J., Zhang, H., Gu, W. (2014): Application of brackish ice on the salt content and nutrient levels of saline soil in FGD gypsum-amended, man-made, raised bed agroecosystem. – Soil Science Society of America Journal 78: 1734-1740. https://doi.org/10.2136/sssaj2014.03.0095.
- [102] Tao, J., Wu, L. H., Gu, W., Zhang, H. (2019): Effect of continuous application flue-gas desulfurization gypsum and brackish ice on soil chemical properties and maize growth in saline soil in coastal area of China. – Soil Science and Plant Nutrient 65: 82-89. https://doi.org/10.1080/00380768.2018.1531355.
- [103] Tao, Y., Gu, W., Chen, J., Tao, J., Xu, Y. J., Zhang, H. (2013): The influence of land use practices on earthworm communities in saline agriculture soils of the west coast region of China's Bohai Bay. – Plant Soil and Environment 59: 8-13. https://doi.org/10.17221/374/2012-PSE.
- [104] Tejada, M., Garcia, C., Gonzalez, J. L., Hernandez, M. T. (2006): Use of organic amendment as a strategy for saline soil remediation, influence on the physical, chemical and biological properties of soil. – Soil Biology and Biochemistry 38: 1413-1421. https://doi.org/10.1016/j.soilbio.2005.10.017.
- [105] Tripathi, S., Kumari, S., Chakraborty, A., Gupta, A., Chakrabarti, A., Bandyapadhyay, K. B. (2006): Microbial biomass and its activities in salt-affected coastal soils. – Biology and Fertility of Soils 42: 273-277. https://doi.org/10.1007/s00374-005-0037-6.
- [106] Tsanis, I. K., Daliakopoulos, I. N., Koutroulis, A. G., Karatzas, G. P., Varouchakis, E., Kourgialas, N. (2016): Soil Salinization. – In: Stolte, J. et al. (eds.) Soil Threats in

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Europe: Status, Methods, Drivers and Effects on Ecosystem Services. Review Report, RECARE Project. European Union, Luxembourg, pp. 104-117.

- [107] Uchida, Y., Nishimura, S., Akiyama, H. (2012): The relationship of water-soluble carbon and hot-water-soluble carbon with soil respiration in agricultural field. – Agricultural Ecosystems and Environment 156: 116-122. https://doi.org/10.1016/j.agee.2012.05.012.
- [108] Van Groenigen, J. W., Lubbers, I. M., Vos, H. M. J., Brown, G. G., De Deyn, G. B., Van Groenigen, K. J. (2014): Earthworms increase plant production, a meta-analysis. – Scientific Reports 4: 6365. https://doi.org/10.1038/srep06365.
- [109] Van Groenigen, J. W., Van Groenigen, K. J., Koopmans, G. F., Stokkermans, L., Vos, H. M. J., Lubbers, I. M. (2019): How fertile are earthworm casts? A meta-analysis. – Geoderma 338: 525-535. https://doi.org/10.1016/j.geoderma.2018.11.001.
- [110] Van Leeuwen, C. J., Vermeire, T. G. (2007): Risk Assessment of Chemicals: An Introduction. 2nd Ed. – Springer, Dordrecht.
- [111] Versteegh, E. A. A., Black, S., Hodson, M. E. (2014): Environmental controls on the production of calcium carbonate by earthworms. – Soil Biology and Biochemistry 70: 159-161. https://doi.org/10.1016/j.soilbio.2013.12.013.
- [112] Vos, H. M. J., Ros, M. B. H., Koopmans, G. F., van Groenigen, J. W. (2014): Do earthworms affect phosphorus availability to grass? A pot experiment. – Soil Biology and Biochemistry 79: 34-42. https://doi.org/10.1016/j.soilbio.2014.08.018.
- [113] Wang, X., Zhao, Q., Hu, Y., Zheng, Y., Wu, X., Wu, H. (2012): An alternative water source and combined agronomic practices for cotton irrigation in coastal saline soils. – Irrigation Science 30: 221-232. https://doi.org/10.1007/s00271-011-0277-1.
- [114] Wang, Y., Li, Y. (2012): Land exploitation resulting in soil salinization in a desertoasis ecotone. – Catena 100: 50-56. https://doi.org/10.1016/j.catena.2012.08.005.
- [115] Wang, Y., Chen, J., Gu, W., Xu, Y. J., Gu, J. Y., Tao, J. (2016): Earthworm activities increase the leaching of salt and water from salt-affected agricultural soil during the wetdry process under simulated rainfall conditions. – Biology and Fertility of Soils 52: 323- 330. https://doi.org/10.1007/s00374-015-1078-0.
- [116] Whalen, J. K., Parmelee, R. W. (1999): Quantification of nitrogen assimilation efficiencies and their use to estimate organic matter consumption by the earthworms Aporrectodea tuberculata (Eisen) and Lumbricus terrestris L. – Applied Soil Ecology 13: 199-208. https://doi.org/10.1016/S0929-1393(99)00033-5.
- [117] Wichern, J., Wichern, F., Joergensen, R. G. (2006): Impact of salinity on soil microbial communities and the decomposition of maize in acidic soils. – Geoderma 137: 100-108. https://doi.org/10.1016/j.geoderma.2006.08.001.
- [118] Wu, W. L., Qiao, Y. H., Xu, Q., Chen, K. (2001): An ecological study on earthworm in farmland of salinity transforming area in north China plain. – Acta Ecologia Sinica 21: 1109-1113.
- [119] Wu, Y. P., Li, Y. F., Zheng, C. Y., Zhang, Y. F., Sun, Z. J. (2013): Organic amendment application influence soil organism abundance in saline alkali soil. – European Journal of Soil Biology 54: 32-40. https://doi.org/10.1016/j.ejsobi.2012.10.006.
- [120] Yan, N., Marschner, P. (2012): Response of microbial activity and biomass to increasing salinity depends on the final salinity, not the original salinity. – Soil Biology and Biochemistry 53: 50-55. https://doi.org/10.1016/j.soilbio.2012.04.028.
- [121] Yu, J., Wang, Z., Meixner, F. X., Yang, F., Wu, H., Chen, X. (2010): Biogeochemical characterizations and reclamation strategies of saline–sodic soil in Northeastern China. – Clean- Soil Air Water 38: 1010-1016. https://doi.org/10.1002/clen.201000276.
- [122] Yuan, B. C., Li, Z. Z., Liu, H., Gao, M., Zhang, Y. Y. (2007): Microbial biomass and activity in salt affected soils under and conditions. – Applied Soil Ecology 35(2): 319- 328. https://doi.org/10.1016/j.apsoil.2006.07.004.
- [123] Zhang, T., Li, S. Y., Sun, X. Y., Zhang, Y., Gong, X. Q., Fu, Y., Jia, L. M. (2015): The earthworm Eisenia fetida can help desalinate a coastal saline soil in Tianjin, North China. – Plos One 10: 1-15. https://doi.org/10.1371/journal.pone.0144709.
- [124] Zhang, W., Cao, J., Zhang, S., Wang, C. (2016): Effect of earthworms and arbuscular mycorrhizal fungi on the microbial community and maize growth under salt stress. – Applied Soil Ecology 107: 214-223. https://doi.org/10.1016/j.apsoil.2016.06.005.
- [125] Zhang, W., Wang, C., Lu, T., Zheng, Y. (2018): Cooperation between arbuscular mycorrhizal fungi and earthworms promotes the physiological adaptation of maize under a high salt stress. – Plant and Soil 423: 125-140. https://doi.org/10.1007/s11104-017- 3481-9.
- [126] Zhu, J. K. (2001): Plant salt tolerance. Trends Plant Science 6: 66-71. https://doi.org/10.1016/S1360-1385(00)01838-0.
- [127] Zorn, M. I., Van Gestel, C. A. M., Morrien, E., Wagenaar, M., Eijsackers, H. (2008): Flooding responses of three earthworm species, *Allolobophora chlorotica*, *Aporrectodea caliginosa* and *Lumbricus rubellus*, in a laboratory-controlled environment. – Soil Biology & Biochemistry 40: 587-593. https://doi.org/ 10.1016/j.soilbio.2007.06.028.