

# RAINFOREST EQUIVALENCE—A NEW APPROACH FOR COMPARATIVE ASSESSMENT OF ECOSYSTEM SERVICES OF HABITATS

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**Abstract.** To establish a framework of global agreements aimed at fostering a sustainable global community, it becomes imperative to possess the capability to measure the ecosystem services rendered by individual nations (or regions) to the entire planet in a manner that allows for meaningful comparisons. An effective valuation system should be capable of verification and recalculation by individuals at any time. It should rely solely on data that are consistently accessible in public databases, traceable, applicable across various spatial and temporal dimensions, and cumulative in nature. The lowland tropical rainforest is often regarded as the epitome of an ideal state of existence among the various biomes found on our planet. This particular biome stands out because of its exceptional levels of biomass, productivity, and biodiversity. The presence of liquid water diminishes considerably as individuals travel towards the northern or southern poles from the equator, and as they ascend higher altitudes in mountainous areas. Similarly, as individuals approach the Earth's poles, the amount of solar radiation suitable for utilization decreases, resulting in a decrease in annual heat input and a subsequent decrease in temperature. It is deemed suitable to quantify the worth of various communities in different habitats by using rainforest equivalents, taking into account their biomass, productivity, and the biodiversity they sustain. This approach to valuation enables quick and effective comparisons at a regional level, thus serving a crucial function in measuring adherence to multilateral international agreements, informing political and economic decisions, and evaluating changes in land use through monitoring.

**Keywords:** *biomes, biodiversity, climate, biomass, carbon*

## Introduction and objective

To develop a system of international conventions for a sustainable global society, it is crucial to quantify the ecosystem services (*Table 1*), provided by individual countries or regions. This quantification should enable meaningful comparisons at a global level. The valuation system must be verifiable and recalculable by anyone, at any time. It should be based only on data that are continuously available in public databases. The data must be traceable, applicable at different spatial and temporal scales, and cumulative.

The ecological conditions necessary for human survival include a climate regulated by biological processes, appropriate chemical composition of the atmosphere, and natural waters and soils. These conditions also involve biological materials used by humanity. Ecosystem services provide these conditions (Daily et al., 1997; Tallis and Kareiva, 2005; Costanza et al., 2017). The quantity and quality of these services depend on the condition, integrity, and health of the ecosystems that produce them (Carpenter et al., 2009). Different habitat types contribute differently to the quantity of services provided by the biosphere. This contribution per unit area (service value) is proportional to the total biomass of the biotic community. This can be expressed in terms of carbon stock as a

quantitative indicator. It also depends on the biological activity of the biotic community, which reflects annual productivity and can be expressed in terms of carbon sequestration. Additionally, it depends on the efficiency and reliability of the processes resulting from functional redundancy. This is an indicator of biodiversity and, on land, can be characterised by the number of vascular plant species.

**Table 1.** *Ecosystem services and functions Based on Newell and Roberts (2017)*

	<b>Ecosystem service</b>	<b>Ecosystem function</b>	<b>Example</b>
1.	Gas regulation	Regulating the chemical composition of the atmosphere	O <sub>2</sub> /CO <sub>2</sub> balance, O <sub>3</sub> for UVB protection, SO <sub>x</sub> levels
2.	Climate regulation	Temperature, rainfall regulation, and other biologically influenced processes	Dimethyl sulfoxide affects cloud formation
3.	Disruption and disaster recovery	Maintaining ecosystem integrity, response to fluctuations	Storm protection, flood mitigation
4.	Water control	Regulation of hydrological processes	Agriculture, industry, and transport
5.	Water supply	Water storage and retention	River basins, reservoirs, and catchment areas
6.	Erosion reduction and sediment retention	Maintaining the soil that forms the basis of the ecosystem	Preventing wind and flood erosion
7.	Soil development and maintenance	Regulation of biological processes that lead to soil formation	Weathering of rocks, accumulation of organic matter
8.	Regulation of nutrients	Regulation of C, N, and other cycles, processing, storage, and accumulation of nutrients	Nitrogen fixation, supply of nutrients to agriculture
9.	Contamination treatment	Removal or breakdown of toxic xenobiotic substances	Detoxification
10.	Pollination	Transport of pollen	Production of agricultural crops
11.	Biological control	Regulating populations	Pest control by predators
12.	Refugium	Habitat for local and migratory populations	Providing a protected site for species of economic importance
13.	Food production	A percentage of the ecosystem's production can be used as food	Fish, game, cereals and fruit
14.	Raw materials	A percentage of the ecosystem's production can be extracted as raw material	Timber, fuel and feed
15.	Genetic pool	Source of biological production	Genes to build resistance
16.	Recreation	Opportunity for recreational activities	Ecotourism, fishing
17.	Cultural	Providing opportunities for cultural activities, spiritual and scientific values	Art, aesthetics, education

Among the biomes of our planet, the lowland tropical rainforest is considered an “ideal state of existence” in terms of biomass, productivity, and biodiversity (Alexandrov and Matsunaga, 2008; Keith et al., 2009; Mics et al., 2013; Eiserhardt et al., 2017). In comparison, all other habitats are ‘struggle zones’ with much lower values (Hufnagel et al., 2018). This distinctive feature of tropical rainforests is due to high, stable temperatures, which are favourable for biological processes. It is also due to high solar radiation, high humidity, and good availability of liquid water (Park, 2003). Moving north

or south of the equator, or increasing altitude in mountains, reduces the availability of liquid water significantly. Moving towards the poles reduces the amount of usable solar radiation, decreases annual heat input, and lowers temperatures (Woodward et al., 2004).

It seems appropriate to express the value of communities in different habitats in terms of rainforest equivalents. This should be done in proportion to their biomass, productivity, and the biodiversity they support. This method of valuation allows for rapid and efficient regional comparisons. It can play an important role in quantifying commitments to international agreements. It can also assist in preparing political and economic decisions and in assessing land-use changes.

The aim of this paper is:

1. To develop a calculation procedure for a valuation system based on rainforest equivalency. This includes collecting the necessary baseline data and developing a global valuation of ecosystem services by biome and continent.
2. To conduct a global comparative valuation case study at the continental level. This serves as an example application and provides a conceptual basis for potential international agreements.

## Review of literature

### *Ecosystem services valuation methodologies*

Professionals are increasingly facing the challenge of estimating the value of specific areas, ecosystems, or species. Decisions are needed on how to manage an area and what should happen to its wildlife. For example, should a forest remain in its natural state or be converted for forestry? To answer this, the value of the forest must be assessed.

Two approaches to valuing natural ecosystems are found in the literature: anthropocentric and biocentric. The anthropocentric approach values nature based on its usefulness to humanity. The biocentric approach, however, considers everything in nature to have intrinsic value, regardless of its utility to humans (Daily, 1997). Proponents of the anthropocentric approach argue that since humans are the dominant species, they have the right to determine what is valuable for them (Daily, 1997).

Another approach views nature as having both direct (use value) and indirect (non-use value) value (de Groot et al., 2010). According to the Millennium Ecosystem Assessment, there are four categories of benefits from nature: provisioning (e.g., fisheries, timber), regulating (e.g., climate and river flow regulation), supporting (e.g., pollination, pest control), and cultural (e.g. tranquillity, inspiration) (Chan et al., 2006).

Since the 1960s, there has been increasing attention in the literature on assessing the value of ecosystems (Hein et al., 2006). Since the term “ecosystem services” was first mentioned in 1983, the number of related articles and citations has risen steeply (Costanza and Kubiszewski, 2012). Ecosystems provide a wide range of goods and services essential for human well-being (Nelson et al., 2009).

To protect ecosystems and their services, policymakers need to ensure that human activities are sustainable and that resources are allocated equitably and efficiently (Costanza and Folke, 1997). It must also be recognised that the value of nature and its contribution to quality of life cannot be separated from cultural and institutional contexts (Pascual et al., 2017). Political decisions and public opinion greatly influence the perceived value of services, making their valuation controversial (Loomis et al., 2000).

Some believe it is not possible or wise to value certain aspects of nature, as intangibles like aesthetics or long-term ecological benefits are hard to quantify (Costanza, 2000). As a result,

there can be significant contradictions between economic and ecological valuations (Farber et al., 2002). Particularly in Western countries, it is important to value natural ecosystems, where high productivity is prioritised in economic decisions (Gómez-Baggethun et al., 2009).

Expressing ecosystem services in monetary terms does not mean they can be treated as marketable products or private property (Costanza et al., 2014). For example, pollination and water control benefit everyone but cannot be owned (Wilson and Howarth, 2002). Policy decisions should consider this, though translating ecosystem service valuation into financial mechanisms is still unresolved (Daily et al., 2009). Ecosystem services are hard to compare to factory goods, which results in them being given little weight in policy decisions (Chee, 2004). However, economic valuation is crucial for monitoring services (Kumar and Kumar, 2008).

The attitude towards valuing services is illustrated by the water and diamond paradox. Water, essential to life, is valued little, while diamonds, which are not essential, have a high monetary value (Heal, 1999). Humanity benefits from natural ecosystems but also alters them. It is vital to monitor changes in ecosystem conditions, as their degradation affects human quality of life (Howarth and Farber, 2000). Ecological processes are under threat due to human activities. Habitat destruction, conversion, and pollution lead to the disappearance of natural ecosystems globally (Barbier, 2007). Despite environmental regulations, agricultural, industrial, and residential developments continue to degrade natural vegetation (National Research Council, 2005). These threats are expected to grow as demand for energy and raw materials increases (de Groot et al., 2012).

Most people today live disconnected from nature, and conservation is often seen as an obstacle to development. However, ecosystem services can shift this perspective, making conservation a driver of development (Gómez-Baggethun and Pérez, 2011). Valuing ecosystem services helps decision-makers choose between management options to achieve multiple objectives (Liu et al., 2010). This system links ecology with economics, so economic methods should be used to value ecosystem components (Chan et al., 2012). Many methods exist to determine the monetary value of services, though missing data make this challenging (Sherrouse et al., 2011).

## ***Direct market valuation methods***

### *Revealed preference methods*

#### Market price method

In some cases, the value of a service can be measured directly from the market price of its products, which can be sold in the market. In these cases, the value is based on the transaction price. This means no complex methods are needed. Examples include timber, firewood, fish, and other foodstuffs. The value of these commodities also reflects the value of the ecosystem service.

The advantage of this method is its simplicity. It uses available information on prices, quantities, and costs, and requires only simple assumptions. However, many services are not directly tradable, which can lead to false or distorted information that does not reflect the true value of the service. Additionally, it is difficult to use for large-scale changes that affect the supply and demand of the service (Koetse et al., 2015).

#### Production function method

This approach is used when a good or service is partly the result of human labour and partly due to ecosystem contributions. For example, many agricultural crops depend on

insect pollination, and the value of pollination can be estimated from the value and quality of these crops. It is a method developed to estimate indirect use values.

The difficulty lies in determining the relationship between ecosystem services and human contributions, making this method less commonly used. However, it has been applied to measure water quality or changes in water quality based on reduced treatment costs, improved crop production due to better pollination, or improved soil quality. In other words, the quality of a marketable commodity improves due to an ecosystem service.

A problem with this method is that researchers must consider both human and mechanical inputs, which can lead to overestimating the value of the ecosystem service. Despite this, it has the advantage of being theoretically suitable for valuing ecosystem services, as it assumes a close relationship between the service and its economic benefit (Pascual et al., 2009).

#### Cost-based methods

This method measures the value of an ecosystem service by estimating the damage that would occur if the service were not provided. It also considers the costs of replacing the service. It is commonly used to assess water quality, water purification costs, and protection against soil erosion, storms, and other natural disasters. It also measures the protection of natural habitats. In this case, the service is a non-marketable product and reflects the cost of producing the benefit, rather than the benefit itself. This method aligns with how the economy values and creates value. However, a disadvantage is that the cost of repairing the damage may not always reflect the actual benefits received (Daly, 2016).

#### Random utility and travel-cost methods

The travel-cost method and the random utility method are based on the assumption that people know their preferences, although researchers do not always know these preferences. Certain aspects of preferences can be determined using statistical methods. These methods are mostly used to evaluate recreational fishing in lakes, rivers, and the sea. They assess the value of a non-marketable ecosystem service by measuring how much money and time people spend to reach a fishing or swimming site. Time, money, and the number of visits represent the value of the site, fishing, and swimming (National Research Council, 2005).

#### Hedonic pricing method

This method measures the indirect value of an ecosystem service that cannot be sold but can be estimated through the value of a related good. To determine the value, two products are needed that are identical except for some environmental factors, such as traffic noise or distance from a park. The difference in their monetary value reflects people's willingness to pay for an ecosystem service. This method is often used to estimate the benefits or costs related to the quality of the natural environment, such as air pollution, water pollution, or noise. For example, the value of houses can reflect the quality of the environment. A house in a better environment is worth more, while a similar house in an area with higher air pollution may be worth less. This analysis helps determine whether a change in environmental factors affects the value of a marketable good (Bouma and van Beukering, 2015).

### *Stated preference methods*

#### Contingent valuation

This method uses questionnaires to assess the value of ecosystem services. The responses show how much people are willing to pay for certain services. In other words, it tries to understand how people would behave in specific situations. Since these services are not sold on the market, the questions ask how much the respondent would pay in a given situation. Options may include a new tax, an entrance fee to a national park, or an annual, monthly, or one-off maintenance fee. This method is widely used for assessing the value of community property. However, respondents often struggle to determine how much they would pay for the service. Many value the ecosystem highly but cannot assign it a monetary value, and responses also depend on their income level (Carson and Bergstrom, 2003).

#### Conjoint analysis

This is another popular method based on questionnaires. Respondents answer questions about the characteristics of a product or service. For example, they may choose between two options describing possible features of a park, such as distance from their home, size, vegetation, and accessibility. Statistical analysis then reveals the relative importance of these characteristics to the respondents. It also shows how far people are willing to travel to visit the park. Responses can be compared to those for other recreational opportunities (Bergkamp and Goldsmith, 2013).

### *Non-monetary valuation*

There are cases where monetary valuation methods cannot be used or are not suitable. This may be due to the nature of the ecosystem service, uncertainty about changes in natural factors, or the intentions of the participants. In such cases, other options are available (Hadley et al., 2011).

### *Interviews and focus groups*

Led by a moderator, 7-10 people who do not know each other discuss a topic in a structured manner. Although they are strangers, they share common characteristics or interests. During the discussion, participants express their opinions, and the moderator observes trends and patterns in their thinking. They are not pressured to vote, plan, or reach a consensus. In-depth interviews are similar to focus groups but aim to provide a detailed understanding of the research subject. In these interviews, experts are also involved (Erdoğan et al., 2016).

### *Citizen's jury*

This method involves a test with 12-25 people, guided by an independent moderator. The aim is to obtain an informed opinion from the community on a specific issue or when choosing between different decision options. Experts and stakeholders present their opinions and evidence. The jury considers this information carefully and forms an opinion, which is only a recommendation and not binding. Jury members are selected from all sections of society to represent the whole population and increase public participation in the democratic process (Wittmer and Gundimeda, 2012).

### *Health-based valuation approaches*

This approach is based on the impact on health, considering both the quality and length of life of the people studied. It measures how a person's health has improved or deteriorated over time, considering factors in the natural environment (Ten Brink, 2011).

### *Q-methodology*

This is a thorough and statistically reliable method for investigating people's subjective opinions. Participants complete a test called a Q-sort, where they rank various statements based on different criteria. This helps reveal the values and preferences that the community considers important. Participants associate negative, neutral, and positive indicators with different statements, and a ranking of importance is established. The results are then analysed using factor analysis (Armatas et al., 2014).

### *Delphi method*

The aim of this method is to collect and process the comments and opinions of experts in a particular field. By continuously reviewing and refining opinions, a consensus is eventually reached in the investigation. The collective input of several experts helps solve the problem more effectively. Originally developed for military purposes in the 1950s during the Cold War, this method is now used successfully in many disciplines (Boberg and Morris-Khoo, 1992).

### *Summary of literature*

Plants and plant ecosystems are essential in assessing land management practices, as they are significant natural and cultural assets (Wang et al., 2018). An important factor in this research is improving the objectivity and ease of conducting biological assessments. This can be done by developing formulas to quantify the environmental factors involved. Quantification is crucial, especially when working with planning authorities, who prefer numerical approaches.

Ecosystem valuation involves assigning a value—monetary, biophysical, or otherwise—to an ecosystem and its services. Quantifying vegetation's benefits, such as flood and erosion control, carbon storage, support for endangered species, and absorption of harmful chemicals, allows for monetizing these advantages. This approach helps policymakers and conservationists assess the impact of management strategies and conduct cost-benefit analyses to compare policies. Assigning monetary value makes evaluating management impacts more objective and aids decision-making. However, these valuations are only approximations, involving quantitative uncertainty and philosophical debate over non-market costs and benefits.

The current global challenge is to conserve natural resources, address climate change through mitigation and adaptation, and prevent the degradation of ecosystem services. Despite the growing importance of ecosystem services, their value remains poorly understood in economic markets and government policies (Hancock, 2010). The importance of biodiversity in supporting ecosystem services and the potential impacts of biodiversity decline are widely recognised (Feld et al., 2009). Preserving natural resources, addressing climate change, and preventing ecosystem degradation are the main challenges facing the global community today.

Estimating the monetary value of ecosystem services can be useful in various contexts. One such context is the use of decision-making tools like cost-benefit analysis and cost-effectiveness analysis, which rely heavily on monetary values. By incorporating these values, these tools provide a comprehensive evaluation of the significance of all aspects of ecosystem services, including both utilitarian and non-utilitarian aspects. This facilitates an objective assessment.

Despite many methodologies for classifying, quantifying, and valuing ecosystem services, choosing the most suitable one remains unclear and often depends on researchers' preferences. Valuing ecosystem services in different ecosystems, such as forests, grasslands, deserts, and wetlands, is challenging due to disputes over the methods used, the specific services valued, and the lack of comprehensive global coverage (Chan and Satterfield, 2020).

No monetary value is currently assigned to the rainforest equivalent, but it could be. The market value of available biomass (carbon stock) and biomass produced (carbon sequestration) is relatively easy to calculate. The value of biodiversity is more difficult to determine, but it is possible. Economic value can be defined as the monetary amount an individual is willing to pay to obtain an ecosystem service (willingness to pay, WTP). Alternatively, it can be the monetary compensation needed to persuade an individual to forgo the service (willingness to accept, WTA) (Loomis et al., 2014).

Market data consists of prices determined by supply and demand, which can include environmental goods. Restoration expenses or compensation payments are part of this category. The hedonic price approach and travel cost method can measure people's preferences for exploring natural landscapes. These techniques are indirect and use proxies to represent the value of the object being evaluated.

When market prices are not available, values can be obtained directly from individuals using stated-preference methods. The contingent valuation and choice modelling methods are used to determine prices for environmental goods based on their characteristics. Other methods include participatory models, which often start with social WTP, and system-dynamic approaches, which focus on the stability of the investigated ecosystem services (ESS).

In the above methods, the perspective of the people using the ecosystem service is relevant. However, it is also important to consider the characteristics of the vegetation, which are independent of users' subjective views. The chosen attributes (carbon stock, carbon sequestration, and biodiversity) influence the ecosystem services provided. Therefore, it is crucial to assess vegetation's intrinsic attributes. These two approaches—user perspectives and vegetation attributes—are complementary, not mutually exclusive. People's assessments can be subjective, so it is essential to also consider vegetation characteristics. Both approaches are important to accurately determine the environmental value of an area.

The rainforest equivalent is a proposed assessment based on vegetation characteristics. It considers both the basic values and their multiplication and sum. This allows for a combined estimation of the value of three types of ecosystem services and introduces a new perspective in valuing natural resources.

## Materials and methods

### *Basic data and their sources*

The extent of biomes on each continent is characterised by potential (pre-human activity) and current (turn of the millennium) values. For anthropogenic land use types, different period-specific values are provided, with data in *Table 2* and sources in *Table 3*.

The first number in each cell of *Table 2* shows the theoretically possible extent of vegetation in a biome without human activity. The second number shows the current extent by the turn of the millennium, reduced due to human activity.

For North America, Oceania, South America, and Asia, rainforest data are combined, as the original author treated these areas together for simplicity. Anthropogenic biomes have no potential value; their extent increases with human population growth (Ellis and Ramankutty, 2008) and the demand for new territories, which accelerated during the industrial revolution. For anthropogenic biomes of artificial origin, the first number is smaller, and the second is larger. The first value is from the 1700s and provides only approximate information.

Differences in measurements by various authors can arise from differences in measurement procedures. Sometimes, certain formations are omitted. For example, some articles mention only savanna, while others distinguish between woody and grassy savannas. A biome may also lack a clear boundary, as with natural steppes and human-created pastures, where grazing can alter and degrade vegetation.

We tried to identify areas with untouched vegetation and used data from those areas. However, this is not possible everywhere. In Europe, for example, temperate forests have long been managed for forestry, which may alter their characteristics compared to pristine forests. Foresters often plant trees in areas with suboptimal conditions or introduce non-native species (Gilliam, 2016). Many grassy areas today are used for livestock grazing, which also changes vegetation characteristics.

Antarctica (14 million km<sup>2</sup>) is excluded from this study because it is covered in ice and has minimal vegetation. Much of Greenland (2,166,086 km<sup>2</sup>) is also covered in ice, but there are areas with tundra vegetation. Since Greenland is administratively part of Denmark, we include it in Europe. The total extent of potential vegetation is estimated at  $1.36 * 10^8$  km<sup>2</sup>. According to the United Nations, the ice-free land surface (excluding Antarctica and Greenland) is  $1.34 * 10^8$  km<sup>2</sup>. The data we used are approximations and may be slightly affected by uncertainties, but this does not significantly impact the applicability of the procedure.

*Table 4* shows the carbon stock, carbon sequestration per unit area, and the number of vascular plant species per unit area. These values were used to characterise the vegetation, except for mangroves, which have very few species adapted to this specialised environment. *Table 5* summarises the sources of the data in *Table 4*.

For the number of vascular plant species per unit area (columns 3 and the last column), the data cannot simply be multiplied by the area. Instead, we used the Arrhenius equation to estimate species richness. This equation is a common tool for estimating species richness in ecology and conservation. According to Arrhenius (1921) and Kier et al. (2005), it follows the power model of the species–area relationship:

$$S_e = S_u * \left(\frac{A_e}{A_u}\right)^z \quad (\text{Eq.1})$$

$S_e$  = biome estimated number of species,

$S_u$  = number of species per unit area,

$A_e$  = area of biome,

$A_u$  = unit area (1 km<sup>2</sup>),

$z$  = parameter defining the slope.

The  $z$  values determining the slope of the curve and their literature sources are given in *Table 6*.

**Table 2.** Potential and actual area values (the first figure is potential, the second is actual)

	Africa km <sup>2</sup>	South America km <sup>2</sup>	North America km <sup>2</sup>	Asia km <sup>2</sup>	Europe km <sup>2</sup>	Pacific km <sup>2</sup>	Total km <sup>2</sup>
Lowland rainforest	4.0177*10 <sup>6</sup> – 8.7000*10 <sup>5</sup>	7.0721*10 <sup>6</sup> – 4.5400*10 <sup>6</sup>	0	3.4907*10 <sup>6</sup> – 1.7700*10 <sup>6</sup>	0	0	1.4581*10 <sup>7</sup> – 7.8000*10 <sup>6</sup>
Montane rainforest	5.4466*10 <sup>5</sup> – 1.5860*10 <sup>5</sup>	1.1506*10 <sup>6</sup> – 5.7340*10 <sup>5</sup>	0	1.5620*10 <sup>6</sup> – 4.8800*10 <sup>5</sup>	0	0	3.2573*10 <sup>6</sup> – 1.2200*10 <sup>6</sup>
Cloud forest	5.7190*10 <sup>4</sup> – 3.4328*10 <sup>4</sup>	9.6394*10 <sup>4</sup> – 8.7626*10 <sup>4</sup>	0	2.2758*10 <sup>5</sup> – 9.2676*10 <sup>4</sup>	0	0	3.8116*10 <sup>5</sup> – 2.1463*10 <sup>5</sup>
Tropical seasonal forest	3.6695*10 <sup>6</sup> – 3.4328*10 <sup>4</sup>	1.6816*10 <sup>6</sup> – 5.6840*10 <sup>5</sup>	2.2600*10 <sup>5</sup> – 1.4682*10 <sup>5</sup>	1.4266*10 <sup>6</sup> – 1.4682*10 <sup>5</sup>	0	4.6800*10 <sup>5</sup> – 3.9850*10 <sup>4</sup>	7.4717*10 <sup>6</sup> – 9.3622*10 <sup>5</sup>
Mangrove	5.3719*10 <sup>4</sup> – 3.6529*10 <sup>4</sup>	6.9614*10 <sup>4</sup> – 4.3161*10 <sup>4</sup>	0	1.2058*10 <sup>5</sup> – 7.7169*10 <sup>4</sup>	0	1.1961*10 <sup>4</sup> – 1.0287*10 <sup>4</sup>	2.5673*10 <sup>5</sup> – 1.6688*10 <sup>5</sup>
Temperate deciduous forest	1.0000*10 <sup>5</sup> – 1.0000*10 <sup>4</sup>	7.0000*10 <sup>5</sup> – 4.0000*10 <sup>5</sup>	4.2600*10 <sup>6</sup> – 2.1300*10 <sup>6</sup>	4.4200*10 <sup>6</sup> – 8.7000*10 <sup>5</sup>	4.4200*10 <sup>6</sup> – 1.1100*10 <sup>6</sup>	6.8000*10 <sup>5</sup> – 7.0000*10 <sup>4</sup>	1.4600*10 <sup>7</sup> – 4.5900*10 <sup>6</sup>
Temperate conifer forest	0	0	1.9717*10 <sup>6</sup> – 6.4619*10 <sup>5</sup>	1.4403*10 <sup>6</sup> – 2.8439*10 <sup>5</sup>	3.13*10 <sup>5</sup> – 9.4451*10 <sup>4</sup>	0	3.7250*10 <sup>6</sup> – 1.0250*10 <sup>6</sup>
Boreal forest	0	0	4.8247*10 <sup>6</sup> – 3.7923*10 <sup>6</sup>	7.1202*10 <sup>6</sup> – 2.2188*10 <sup>6</sup>	2.4992*10 <sup>6</sup> – 2.3829*10 <sup>5</sup>	0	1.4444*10 <sup>7</sup> – 6.2494*10 <sup>6</sup>
Savannas	1.40*10 <sup>7</sup> – 3.07*10 <sup>6</sup>	3.99*10 <sup>6</sup> – 1.37*10 <sup>6</sup>	7.69*10 <sup>4</sup> – 9.96*10 <sup>3</sup>	6.13*10 <sup>4</sup> – 2.05*10 <sup>4</sup>	0	2.14*10 <sup>6</sup> – 1.31*10 <sup>6</sup>	2.0268*10 <sup>7</sup> – 5.7805*10 <sup>6</sup>
Deserts	9.77*10 <sup>6</sup> – 7.30*10 <sup>6</sup>	1.14*10 <sup>6</sup> – 2.93*10 <sup>5</sup>	2.34*10 <sup>6</sup> – 4.88*10 <sup>5</sup>	1.11*10 <sup>7</sup> – 4.26*10 <sup>6</sup>	0	3.58*10 <sup>6</sup> – 2.60*10 <sup>6</sup>	2.7930*10 <sup>7</sup> – 1.4941*10 <sup>7</sup>
Temperate grasslands	3.60*10 <sup>5</sup> – 2.40*10 <sup>5</sup>	1.63*10 <sup>6</sup> – 3.46*10 <sup>5</sup>	3.10*10 <sup>6</sup> – 1.18*10 <sup>5</sup>	4.03*10 <sup>6</sup> – 2.83*10 <sup>6</sup>	7.00*10 <sup>5</sup> – 3.78*10 <sup>5</sup>	5.76*10 <sup>5</sup> – 1.73*10 <sup>3</sup>	1.0396*10 <sup>7</sup> – 3.9137*10 <sup>6</sup>
Montane grassland	3.10*10 <sup>4</sup> – 1.72*10 <sup>4</sup>	8.71*10 <sup>5</sup> – 2.24*10 <sup>5</sup>	3.19*10 <sup>5</sup> – 8.61*10 <sup>4</sup>	3.40*10 <sup>6</sup> – 8.55*10 <sup>5</sup>	1.95*10 <sup>5</sup> – 1.76*10 <sup>5</sup>	5.41*10 <sup>4</sup> – 1.62*10 <sup>4</sup>	4.8701*10 <sup>6</sup> – 1.3745*10 <sup>6</sup>
Tundra	0	0	3.94*10 <sup>6</sup> – 3.78*10 <sup>6</sup>	3.30*10 <sup>6</sup> – 3.08*10 <sup>6</sup>	1.06*10 <sup>6</sup> – 8.53*10 <sup>5</sup>	0	8.3000*10 <sup>6</sup> – 7.7130*10 <sup>6</sup>
Mediterranean vegetation	9.4695*10 <sup>5</sup> – 1.2509*10 <sup>5</sup>	1.4841*10 <sup>5</sup> – 1.1333*10 <sup>5</sup>	1.7643*10 <sup>5</sup> – 1.3649*10 <sup>5</sup>	5.6000*10 <sup>5</sup> – 1.2680*10 <sup>4</sup>	8.8900*10 <sup>5</sup> – 6.4800*10 <sup>5</sup>	8.0282*10 <sup>5</sup> – 5.1195*10 <sup>5</sup>	3.5236*10 <sup>6</sup> – 1.5475*10 <sup>6</sup>
Total	3.3551*10 <sup>7</sup> – 1.1896*10 <sup>7</sup>	1.8550*10 <sup>7</sup> – 8.5589*10 <sup>6</sup>	2.1235*10 <sup>7</sup> – 1.1334*10 <sup>7</sup>	4.2259*10 <sup>7</sup> – 1.7006*10 <sup>7</sup>	1.0076*10 <sup>7</sup> – 3.4977*10 <sup>6</sup>	8.3129*10 <sup>6</sup> – 4.5600*10 <sup>6</sup>	1.3398*10 <sup>8</sup> – 5.6853*10 <sup>7</sup>

	Africa km <sup>2</sup>	South America km <sup>2</sup>	North America km <sup>2</sup>	Asia km <sup>2</sup>	Europe km <sup>2</sup>	Pacific km <sup>2</sup>	Total km <sup>2</sup>
<b>1700-2000</b>							
Human areas	1.4140*10 <sup>3</sup> – 4.1679*10 <sup>4</sup>	7.0490*10 <sup>3</sup> – 1.4854*10 <sup>5</sup>	2.5700*10 <sup>2</sup> – 1.8185*10 <sup>5</sup>	4.3900*10 <sup>2</sup> – 3.8400*10 <sup>4</sup>	4.4390*10 <sup>3</sup> – 1.1406*10 <sup>5</sup>	4.9000*10 <sup>1</sup> – 1.3871*10 <sup>4</sup>	1.3647*10 <sup>4</sup> – 5.3840*10 <sup>5</sup>
<b>1700-2000</b>							
Cropland	5.60*10 <sup>3</sup> – 1.43*10 <sup>6</sup>	1.29*10 <sup>4</sup> – 7.54*10 <sup>5</sup>	5.60*10 <sup>3</sup> – 1.97*10 <sup>6</sup>	1.03*10 <sup>5</sup> – 3.98*10 <sup>6</sup>	1.02*10 <sup>5</sup> – 2.35*10 <sup>6</sup>	2.10*10 <sup>3</sup> – 4.28*10 <sup>5</sup>	2.6560*10 <sup>5</sup> – 1.0902*10 <sup>7</sup>
<b>1980-2000</b>							
Pasture	9.1111*10 <sup>6</sup> – 8.6988*10 <sup>6</sup>	1.0161*10 <sup>7</sup> – 1.1061*10 <sup>7</sup>	3.4300*10 <sup>6</sup> – 3.4860*10 <sup>6</sup>	4.2400*10 <sup>6</sup> – 4.5810*10 <sup>6</sup>	8.5578*10 <sup>5</sup> – 1.8234*10 <sup>6</sup>	4.5347*10 <sup>6</sup> – 4.1946*10 <sup>6</sup>	3.2333*10 <sup>7</sup> – 3.3844*10 <sup>7</sup>
<b>1990-2010</b>							
Agroforest	1.1663*10 <sup>5</sup> – 1.5409*10 <sup>5</sup>	7.8840*10 <sup>5</sup> – 1.2688*10 <sup>6</sup>	2.0481*10 <sup>5</sup> – 3.8660*10 <sup>5</sup>	8.2760*10 <sup>4</sup> – 1.3821*10 <sup>5</sup>	5.9046*10 <sup>5</sup> – 6.9318*10 <sup>5</sup>		1.7831*10 <sup>6</sup> – 2.6408*10 <sup>6</sup>

**Table 3. Sources of territorial values (FRA- Forest Resource Assessment)**

	Africa	South America	North America	Asia	Europe	Pacific
Lowland rainforest	FRA, 2000	FRA, 2000	See South America	FRA, 2000	0	See Asia
Montane rainforest	Kapos et al., 2000; Iremonger et al., 1997; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	Kapos et al., 2000; Iremonger et al., 1997; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	See South America	Kapos et al., 2000; Iremonger et al., 1997; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	0	See Asia
Cloud forest	Bubb et al., 2004; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	Bubb et al., 2004; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	See South America	Bubb et al., 2004; Bruijnzeel et al., 2011; Spracklen and Righelato, 2014	0	See Asia
Tropical seasonal forest	FRA, 2000; Miles et al., 2006	FRA, 2000; Miles et al., 2006	FRA, 2000; Miles et al., 2006	FRA, 2000; Miles et al., 2006	0	FRA, 2000; Miles et al., 2006
Mangrove	Valiela, 2006	Valiela, 2006	See at South America	Valiela, 2006	0	Valiela, 2006

	<b>Africa</b>	<b>South America</b>	<b>North America</b>	<b>Asia</b>	<b>Europe</b>	<b>Pacific</b>
Temperate deciduous forest	Silander, 2001	Silander, 2001	Silander, 2001	Silander, 2001	Silander, 2001	Silander, 2001
Temperate conifer forest		Dinerstein et al., 2019	Dinerstein et al., 2019	Dinerstein et al., 2019		
Boreal forest	Burton et al., 2003	Burton et al., 2003	Burton et al., 2003	Burton et al., 2003	Burton et al., 2003	Burton et al., 2003
Savannas	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	0	Olson et al., 2001; Williams et al., 2020
Deserts	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001-; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	0	Olson et al., 2001; Williams et al., 2020
Temperate grasslands	Carbutt et al., 2011	. Olson et al., 2001-; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	Gibson, 2009	Gibson, 2009; European Environmental Agency, 2002	Mark and McLennan, 2005
Montane grassland	Peart, 2008; Olson and Dineerstein., 2002	Dinerstein et al., 2019	Diaz and Eischeid, 2007; Testolin et al., 2020	Olson and Dineerstein, 2002; Dinerstein et al., 2019	European Environmental Agency, 2002	Dinerstein et al., 2019; Olson and Dineerstein, 2002
Tundra	0	0	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	Olson et al., 2001; Williams et al., 2020	0
Mediterranean vegetation	Cox and Underwood, 2011; European Environmental Agency, 2002 Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010	Cox and Underwood, 2011; Pinborg, 2002; Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010	Cox and Underwood, 2011; Pinborg, 2002; Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010	Cox and Underwood, 2011; European Environmental Agency, 2002; Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010	Cox and Underwood, 2011; Pinborg, 2002; Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010	Cox and Underwood, 2011; European Environmental Agency, 2002; Zahran and Gilbert, 2010; Efe et al., 2008; Blondel and Aronson, 1999; Blondel, 2010

	<b>Africa</b>	<b>South America</b>	<b>North America</b>	<b>Asia</b>	<b>Europe</b>	<b>Pacific</b>
Human areas	Goldewijk et al., 2010a					
Cropland	Goldewijk, 2001; Potapov et al., 2022					
Rangeland; Pastureland	FAO; Goldewijk et al., 2010b					
Planation	Kroeger, 2012					

**Table 4.** Carbon stock, carbon sequestration and vascular plant species number per unit area

	<b>Carbon stock C t/ha</b>	<b>Carbon sequestration C t/ha/year</b>	<b>Species number/10,000 km<sup>2</sup></b>
Lowland rainforest	210	13-17	2750
Montane rainforest	130	7	4500
Cloud forest	69	4	5000
Tropical seasonal forest	82	7.6	1862
Mangrove	79	8	70 species worldwide
Temperate deciduous broadleaf forest	109	4.6	1087
Temperate conifer forest	62.1	5.7	1111
Boreal forest	114.5	4.1	545
Savannas	23.15	6.7	1041
Deserts	3.5	1.15-2.69	457
Temperate grasslands	4	3.75	756
Montane grassland	3.35	1.96	927
Tundra	3.5	1.94	227
Mediterranean vegetation	67	5	1220
Human areas	5	2.2	1684
Cropland	1.875	1.1-5.6	1038
Pastureland	5	4.3	980
Agroforest	162.5	7.2	1671

**Table 5.** Sources of carbon stock, carbon sequestration and species number values per unit area

	Carbon stock C t/ha	Carbon sequestration C t/ha/year	Species number
Lowland rainforest	<a href="http://www.esd.ornl.gov">http://www.esd.ornl.gov</a>	Girardin et al., 2010	Barthlott et al., 2007
Montane rainforest	<a href="http://www.esd.ornl.gov">http://www.esd.ornl.gov</a>	Girardin et al., 2010	Barthlott et al., 2007
Cloud forest	Román-Cuesta et al., 2011	Girardin et al., 2010	Barthlott et al., 2007
Tropical seasonal forest	Becknell et al., 2012	Ito and Oikawa, 2004	Ellis et al., 2012
Mangrove	Yee, 2010	Yee, 2010	Polidoro et al., 2010
Temperate deciduous forest	Lal and Lorentz, 2012	Ito and Oikawa, 2004	Ellis et al., 2012
Temperate conifer forest	Thurner et al., 2014	Luysaert et al., 2007	Ellis et al., 2012
Boreal forest	Thurner et al., 2014	Ito and Oikawa, 2004	Ellis et al., 2012
Savannas	Bouvet et al., 2018	Ito and Oikawa, 2004	Ellis et al., 2012
Deserts	Houghton et al., 2009	Ito and Oikawa, 2004	Ellis et al., 2012
Temperate grasslands	Houghton et al., 2009	Saugier et al., 2001	Ellis et al., 2012
Montane grassland	Oliveras et al., 2013	Sun et al., 2021	Ellis et al., 2012
Tundra	Houghton et al., 2009	Ito and Oikawa, 2004	Ellis et al., 2012
Mediterranean vegetation	Pan et al., 2013	Flexas et al., 2012	Ellis et al., 2012
Human areas	Melillo. et al., 1990	Haberl et al., 2007	Ellis et al., 2012
Cropland	Geist, 2006	Ito and Oikawa, 2004	Ellis et al., 2012
Pastureland	Gibbs et al., 2014	Haberl et al., 2007	Ellis et al., 2012
Agroforest	Nabuurs et al., 2003	Haberl et al., 2007	Ellis et al., 2012

**Table 6.** The *z* values for the Arrhenius equation

Vegetation	<i>z</i> Value	Source
Rainforests	0.2875	Kier et al., 2005
Tropical dry forest	0.21	Kier et al., 2005
Temperate deciduous forest	0.17	Kier et al., 2005
Temperate conifer forest	0.14	Kier et al., 2005
Boreal forest	0.16	Kier et al., 2005
Savanna	0.18	Kier et al., 2005
Temperate grassland	0.12	Kier et al., 2005
Alpine	0.17	Kier et al., 2005
Tundra	0.13	Kier et al., 2005
Mediterranean	0.2	Kier et al., 2005
Deserts	0.11	Kier et al., 2005
Mangrove	-	
Cropland	0.269	Gerstner et al., 2014
Pasture	0.177	Gerstner et al., 2014
Plantation	0.13	Proenca et al., 2010
Urban	0.078	Ceschin et al., 2012

### Calculation and evaluation methods

We calculated indices (ERR - Equivalent Rainforest Rate) for tropical lowland rainforests by dividing the carbon stock (as a measure of biomass), carbon sequestration

(as a measure of production), and the number of vascular plant species per unit area (as an approximation of biodiversity) based on the values from *Table 4*. These indices express the proportion of equivalent rainforest area for each variable. If multiple literature values were available for a characteristic, we used their averages. For mangroves, we used the global species richness value instead of the per unit area value. Although this is an overestimate, its low value means it does not significantly affect the comparison.

These three characteristics are not entirely independent but provide complementary information. If they were completely independent, we would use the product of the three indices (Product ERR) to compare habitats. If they were closely related, we would consider them as different estimates of the same characteristic and use their average (Average ERR). The reality lies between these two extremes, so it is sensible to consider them together.

We multiplied the relative ratios for equivalent rainforest values by the area (in km<sup>2</sup>) of each habitat type being compared (biomes, continents). We then summed these values for each object to determine the size of the equivalent lowland rainforest (ERA - Equivalent Rainforest Area) for that object. This provides a suitable non-monetary characterisation of the natural capital (or expected ecosystem services) of the objects on a global scale. Assigning a monetary value would require pricing a unit area of rainforest, which is beyond the scope of this article.

## Results

### *Values of ERR- Equivalent Rainforest Rate*

Comparing the ERR values for biomes (*Table 7*) shows that both Average ERR and Product ERR have similar biome rankings. In both cases, the highest values are for lowland rainforest, followed by montane rainforest. Next are the cloud forest, tropical seasonal forest, and agroforest groups, though with slight differences in ranking. The lowest values are consistently found in the tundra, followed by deserts and montane grasslands, with other biomes in between.

Interestingly, the species richness of montane rainforests exceeds that of lowland rainforests in all other aspects. This can be explained by species from both lower and higher elevations thriving in these areas. These data may relate to the Intermediate Disturbance Hypothesis (IDH) in biodiversity (Wilkinson, 1999), but its analysis is beyond the scope of this article.

### *Worldwide aggregated Equivalent Rainforest Area (ERA) values for biomass and land use types*

*Table 8* shows the ERA data calculated from the Average and Product ERRs of the three attributes, by multiplying these ERRs by the area of the biomes globally. *Table 9* shows the same data aggregated by continent.

### *Application case study*

To develop the use case study, additional data on human population, environmental pressure (carbon dioxide emissions), and economy (GDP) of the continents are needed. These data are presented in *Table 10*. A comparative assessment can be made using their percentage shares, as shown in *Table 11*.

**Table 7. ERR - Equivalent Rainforest Rate data for each habitat type (biome)**

Biome	ERR Carbon stock	ERR Carbon sequestration	ERR Species number	Average ERR	Product ERR
Lowland rainforest	1.00	1.00	1.00	1.0000	1.0000
Montane rainforest	0.62	0.47	1.64	0.9074	0.4727
Cloud (moss) forest	0.33	0.27	1.82	0.8045	0.1593
Tropical seasonal (dry) forest	0.74	0.51	0.68	0.6422	0.2548
Mangrove	0.38	0.53	0.03	0.3117	0.0051
Temperate broadleaf forest	0.69	0.31	0.40	0.4641	0.0837
Temperate conifer forest	0.29	0.34	0.40	0.34	0.0407
Boreal forest	0.43	0.27	0.20	0.3000	0.0232
Tropical savanna	0.09	0.45	0.38	0.3036	0.0145
Deserts	0.01	0.13	0.17	0.1028	0.0003
Temperate grassland	0.03	0.25	0.27	0.1854	0.0022
Montane grassland	0.02	0.09	0.34	0.1488	0.0005
Tundra	0.01	0.13	0.08	0.0746	0.0001
Mediterranean vegetation	0.32	0.33	0.44	0.3653	0.0472
Human areas (urban and villages)	0.02	0.15	0.61	0.2609	0.0021
Cropland	0.01	0.22	0.38	0.2032	0.0008
Pastureland	0.02	0.29	0.36	0.2223	0.0024
Agroforest	0.77	0.48	0.61	0.6205	0.2257

**Table 8. Biomass Equivalent Rainforest Area (ERA) values on our planet**

	ERA by Average ERR [km <sup>2</sup> ]		ERA by Product ERR [km <sup>2</sup> ]	
	Potential	Actual	Potential	Actual
Lowland rainforest	1.46*10 <sup>7</sup>	7.18*10 <sup>6</sup>	1.46*10 <sup>7</sup>	7.18*10 <sup>6</sup>
Montane rainforest	2.96*10 <sup>6</sup>	1.11*10 <sup>6</sup>	1.54*10 <sup>6</sup>	5.77*10 <sup>5</sup>
Cloud (moss) forest	2.45*10 <sup>5</sup>	1.38*10 <sup>5</sup>	9.71*10 <sup>4</sup>	5.47*10 <sup>4</sup>
Tropical seasonal (dry) forest	1.59*10 <sup>5</sup>	1.04*10 <sup>5</sup>	5.77*10 <sup>4</sup>	3.77*10 <sup>4</sup>
Mangrove	6.01*10 <sup>6</sup>	1.46*10 <sup>6</sup>	1.19*10 <sup>6</sup>	2.90*10 <sup>5</sup>
Temperate broadleaf forest	7.51*10 <sup>6</sup>	4.75*10 <sup>6</sup>	1.35*10 <sup>6</sup>	8.57*10 <sup>5</sup>
Temperate conifer forest	1.29*10 <sup>6</sup>	3.56*10 <sup>5</sup>	1.52*10 <sup>5</sup>	4.18*10 <sup>4</sup>
Boreal forest	1.44*10 <sup>7</sup>	7.19*10 <sup>6</sup>	1.86*10 <sup>6</sup>	9.29*10 <sup>5</sup>
Tropical savanna	1.06*10 <sup>6</sup>	3.32*10 <sup>5</sup>	8.18*10 <sup>4</sup>	2.57*10 <sup>4</sup>
Deserts	4.43*10 <sup>6</sup>	1.37*10 <sup>6</sup>	2.11*10 <sup>5</sup>	6.55*10 <sup>4</sup>
Temperate grassland	2.96*10 <sup>6</sup>	1.11*10 <sup>6</sup>	4.85*10 <sup>4</sup>	1.82*10 <sup>4</sup>
Montane grassland	7.07*10 <sup>5</sup>	2.18*10 <sup>5</sup>	2.37*10 <sup>3</sup>	7.33*10 <sup>2</sup>
Tundra	7.51*10 <sup>5</sup>	7.51*10 <sup>5</sup>	8.75*10 <sup>3</sup>	8.75*10 <sup>3</sup>
Mediterranean vegetation	1.11*10 <sup>6</sup>	5.62*10 <sup>5</sup>	9.12*10 <sup>3</sup>	4.60*10 <sup>3</sup>
Human areas (urban and villages)		2.08*10 <sup>5</sup>		7.71*10 <sup>2</sup>
Cropland		3.21*10 <sup>6</sup>		1.08*10 <sup>4</sup>
Pastureland		3.48*10 <sup>6</sup>		1.03*10 <sup>4</sup>
Agroforest		1.97*10 <sup>5</sup>		3.36*10 <sup>2</sup>
<b>Total</b>	<b>5.94*10<sup>7</sup></b>	<b>3.45*10<sup>7</sup></b>	<b>2.11*10<sup>7</sup></b>	<b>1.01*10<sup>7</sup></b>

**Table 9.** Continents' Equivalent Rainforest Area (ERA) values as their contribution to global ecosystem services

	ERA by Average ERR [km <sup>2</sup> ]		ERA by Product ERR [km <sup>2</sup> ]	
	Potential	Actual	Potential	Actual
Europe	2.50*10 <sup>6</sup>	1.81*10 <sup>6</sup>	9.55*10 <sup>4</sup>	3.78*10 <sup>4</sup>
Asia	1.65*10 <sup>7</sup>	9.22*10 <sup>6</sup>	5.30*10 <sup>6</sup>	2.43*10 <sup>6</sup>
Africa	1.95*10 <sup>7</sup>	8.29*10 <sup>6</sup>	6.67*10 <sup>6</sup>	1.83*10 <sup>6</sup>
North America	4.87*10 <sup>6</sup>	3.35*10 <sup>6</sup>	2.38*10 <sup>5</sup>	1.03*10 <sup>5</sup>
South America	1.28*10 <sup>7</sup>	8.12*10 <sup>6</sup>	8.35*10 <sup>6</sup>	5.22*10 <sup>6</sup>
Australia and Oceania	3.21*10 <sup>6</sup>	3.76*10 <sup>6</sup>	4.26*10 <sup>5</sup>	4.70*10 <sup>5</sup>
Antarctica				
<b>Total</b>	<b>5.94*10<sup>7</sup></b>	<b>3.45*10<sup>7</sup></b>	<b>2.11*10<sup>7</sup></b>	<b>1.01*10<sup>7</sup></b>

**Table 10.** Data from continents on the characteristics of their human societies (as a measure of their use of and pressure on ecosystem services)

	Population	CO <sub>2</sub> -emission (t, 2021)	GDP (nominal, billions of USD, 2021)
Europe	7.48*10 <sup>8</sup>	5.31*10 <sup>9</sup>	2.35*10 <sup>4</sup>
Asia	4.68*10 <sup>9</sup>	2.17*10 <sup>10</sup>	3.68*10 <sup>4</sup>
Africa	1.37*10 <sup>9</sup>	1.45*10 <sup>9</sup>	2.69*10 <sup>3</sup>
North America	5.97*10 <sup>8</sup>	6.14*10 <sup>9</sup>	2.68*10 <sup>4</sup>
South America	4.34*10 <sup>8</sup>	1.07*10 <sup>9</sup>	3.25*10 <sup>3</sup>
Australia and Oceania	4.32*10 <sup>7</sup>	8.34*10 <sup>8</sup>	1.89*10 <sup>3</sup>
Antarctica	0	0	0
<b>Total</b>	<b>7.87*10<sup>9</sup></b>	<b>3.65*10<sup>10</sup></b>	<b>5.49*10<sup>4</sup></b>
<b>Source</b>	statisticstimes.com	<a href="https://ourworldindata.org">https://ourworldindata.org</a>	<a href="https://statisticstimes.com">https://statisticstimes.com</a>

**Table 11.** The number and distribution of humanity by continent according to [www.worldometer.info](http://www.worldometer.info). Data download date 01.01.2023

Continent	Population 2020	Area (km <sup>2</sup> )	Density (P/km <sup>2</sup> )	World population share
Europe	7.48*10 <sup>8</sup>	2.21*10 <sup>7</sup>	34	9.59%
Asia	4.64*10 <sup>9</sup>	3.10*10 <sup>7</sup>	150	59.54%
Africa	1.43*10 <sup>9</sup>	2.96*10 <sup>7</sup>	45	17.20%
North America	5.92*10 <sup>8</sup>	2.13*10 <sup>7</sup>	28	7.60%
South America	4.31*10 <sup>8</sup>	1.75*10 <sup>7</sup>	25	5.53%
Australia/Oceania	4.31*10 <sup>7</sup>	8.49*10 <sup>6</sup>	5	0.55%
Antarctica	0	1.37*10 <sup>7</sup>	0	0.00%

By examining the first four columns of *Table 12*, it is evident that Africa has the largest relative deficit between its potential and actual shares of providing ecosystem services. Africa, with its large equatorial regions, provides substantial ecosystem services but may have suffered the most damage, as its actual share is only one-third of its potential. In contrast, Australia and Oceania show the largest relative surplus, with an actual share

twice their potential, as they were among the last regions to be extensively exploited by human civilization. However, this is mainly of natural historical interest.

**Table 12.** Comparing continents by their contribution to and percentage of use of global ecosystem services

	Contribution to ecosystem services				Use of ecosystem services		
	Potential ERA by Avg EER [km <sup>2</sup> ]	Potential ERA by Prod EER [km <sup>2</sup> ]	Actual ERA by Avg EER [km <sup>2</sup> ]	Actual ERA by Prod EER [km <sup>2</sup> ]	Population	CO <sub>2</sub> emission (t, 2017)	GDP (nominal, billions of USD, 2021)
Europe	4.2%	0.5%	5.2%	0.4%	11.7%	17.7%	42.8%
Asia	27.8%	25.1%	26.7%	24.1%	50.8%	55.1%	67.1%
Africa	32.8%	31.6%	24.0%	18.1%	20.9%	3.8%	4.9%
North America	8.2%	1.1%	9.7%	1.0%	9.2%	18.8%	48.8%
South America	21.5%	39.6%	23.5%	51.7%	6.7%	3.2%	5.9%
Australia and Oceania	5.4%	2.0%	10.9%	4.7%	0.7%	1.4%	3.5%
Antarctica							
<b>Total</b>	100%	100%	100%	100%	100%	100%	100%

The key information comes from comparing the first four columns with the last three: Europe, Asia, and North America use global ecosystem services more than they contribute to them. In contrast, Africa, South America, Australia, and Oceania contribute significantly more than they use. Therefore, as long as this imbalance continues, it would be advisable for Europe, Asia, and North America, with larger GDPs and greater environmental impacts, to provide financial contributions. This would help Africa, South America, and Oceania continue providing ecosystem services through their conservation efforts, with lower per capita burdens. These comparisons can be broken down to countries, states, or smaller administrative units and could form the basis for detailed international agreements.

## Discussion

In this article, we reviewed habitat types from the equator to the Arctic tundra. Due to human activities, natural vegetation-covered areas on Earth have decreased almost everywhere (Hoekstra et al., 2005). In contrast, human-transformed areas, such as agricultural lands, urban areas, and infrastructure, are increasing (Steffen et al., 2007). With the rapid growth of the human population, the land needed for food production and raw materials has also increased (Bongaarts, 2009).

The tropical rainforest is one of the largest, most productive habitats with high biodiversity, which is why we chose it as a reference for comparison. The highest obtained ratios were for the rainforest, so we used it as the unit. Montane rainforests had even higher species richness per unit area. Their high biodiversity in relatively small areas makes them especially valuable for conservation (Gradstein et al., 2008; Bendix et al., 2010). This high biodiversity can be explained by the island effect (Triantis et al., 2006) and possibly by their transitional nature or the Intermediate Disturbance Hypothesis (IDH).

Generally, the calculated ERR value decreases as we move away from the equator. Temperate and boreal forests have a relatively high carbon stock per unit area but do not reach rainforest levels. Significant habitat loss has occurred in temperate forests and grasslands, leaving little intact vegetation due to dense agricultural activity and human population in these regions. The least affected habitats are tundra, montane grasslands, and deserts, which have low carbon stock and primary production due to harsh climates. These biomes also have limited agricultural significance. Agricultural areas sequester less carbon compared to intact vegetation, and their ERR values are low, approaching desert levels. Plantation forests have higher values but still do not reach the maximum, as trees are harvested before achieving the carbon stock typical of old-growth forests (Thornley and Cannell, 2000).

Natural habitat areas have not only decreased in extent but also suffer from degradation and fragmentation, reducing their ability to reach their maximum potential (Sloan et al., 2014). As a result, the ability of terrestrial vegetation to sequester atmospheric carbon dioxide and produce biomass has declined. This leads to more carbon dioxide in the atmosphere, increasing the greenhouse effect and global temperature. Vegetation plays an essential role as an ecosystem service provider, and without it, carbon dioxide accumulates in the atmosphere. Human-transformed vegetation has a low ERR value and cannot fulfil this role. Comprehensive protection of habitats is needed to preserve the largest portion of all three levels of biodiversity: genetic, taxonomic, and ecological (Olson and Dinerstein, 1998).

Asia and Africa represent the largest share of the average rainforest equivalent. These continents have the most vegetation in terms of rainforest equivalent, providing valuable ecosystem services despite human activities. For the product of rainforest equivalent, South America stands out with the highest rainforest equivalent area, indicating that its remaining natural vegetation provides the most and highest-quality ecosystem services.

Asia has the largest population and highest carbon dioxide emissions among all continents. Many developing countries in Asia have a small ecological footprint per capita. High population density is mostly in the southern part of the continent, while central and northern regions have fewer people and lower emissions. Siberia has a large area of natural vegetation, contributing significantly to Asia's ERA.

Africa has the lowest GDP among continents and ranks in the middle for carbon dioxide emissions. It has the second-largest population after Asia, but income levels are low, and the natural environment is less transformed. In terms of the product of ERA, South America has the most rainforest, which forms the basis of the new measurement method. South America has many valuable areas due to its large remaining natural vegetation, which is crucial for providing ecosystem services that support human life and quality of life.

Protecting the natural environment, optimising resource use, and improving environmental security are vital for the sustainability of developing countries. However, developing countries face ongoing environmental degradation through agriculture, water consumption, deforestation, pollution, and poor-quality food products (Glantz, 1999; Tamazian and Rao, 2010; Adebayo et al., 2021). The relationship between environmental degradation and economic growth is a key area in ecological economics (Keho, 2017). Sustainable economic development can only be achieved alongside sustainable environmental development (Tiwari et al., 2013).

The link between economic growth and CO<sub>2</sub> emissions has been extensively studied in recent decades. Many countries face the challenge of maintaining economic growth

while protecting the environment. Economic growth often leads to higher energy use and CO<sub>2</sub> emissions, contributing to climate change. On the other hand, growth can also lead to the adoption of energy-efficient and low-carbon technologies. Understanding the relationship between environmental quality and economic growth helps policymakers balance these factors. Economic growth impacts the environment, and the main goal of any economy is to maximise growth (Živanović et al., 2016).

The sources of economic growth and differences in economic performance across regions are significant and challenging aspects of modern development, making them a major focus of scientific study (Greiner et al., 2005). In industrialising countries with low per capita income, emissions of certain pollutants may increase with economic growth. However, once per capita income reaches a certain level, further growth can lead to more environmental damage. Improvements may come from lower population growth, a shift to less polluting goods, increased imports of pollution-intensive goods, environmental policies, government agencies focused on environmental control, and the development of emission-reducing technologies (Komen et al., 1997; Mushafiq and Prusak, 2023).

Developed countries usually prioritise environmental protection, but differences in protection levels exist even among countries with similar economic performance. Individual factors can also influence environmental policy decisions. Recent social science research shows a growing interest in understanding what affects public support for environmental projects. It suggests that socio-demographic factors, such as age, education, gender, race, ideology, party affiliation, and urbanisation, as well as economic variables like employment and income, shape public opinion (Marsiliani and Renström, 2000).

It is important to note some limitations (and areas for further development) in our work. Our research did not include Antarctica or the world's oceans. These areas are nominally linked to states and alliances, but they have no substantial population or independent economy, making them subjects of international agreements rather than active participants.

A more significant criticism may be that we limited the assessment of biodiversity to vascular plants for data reliability and simplicity. Therefore, we did not consider mosses, lichens, microorganisms, or animal biomass, production, or biodiversity. A discussion of these issues and relevant supplementary data can be found in the appendix of our article.

## REFERENCES

- [1] Adebayo, T. S., Akinsola, G. D., Odugbesan, J. A., Olanrewaju, V. O. (2021): Determinants of environmental degradation in Thailand: empirical evidence from ARDL and Wavelet coherence approaches. – *Pollution* 7(1): 181-196.
- [2] Alexandrov, G., Matsunaga, T. (2008): Normative productivity of the global vegetation. – *Carbon Balance and Management* 3: 8.
- [3] Armatas, C. A., Venn, T. J., Watson, A. E. (2014): Applying Q-methodology to select and define attributes for non-market valuation: a case study from northwest Wyoming, United States. – *Ecological Economics* 107: 447-456.
- [4] Arrhenius, O. (1921): Species and area. – *Journal of Ecology* 9: 95-99.
- [5] Barbier, B. E. (2007): Valuing ecosystem services as productive inputs. – *Economic Policy* 22(49): 178-229.
- [6] Barthlott, W., Hostert, A., Kier, G., Küper, W., Kreft, H., Mutke, J., Rafiqpoor, M. D., Sommer, H. (2007): Geographic patterns vascular plants diversity at continental to global scales. – *Erdkunde* 61: 305-315.

- [7] Becknell, J. M., Kucek, L. K., Powers, J. S. (2012): Aboveground biomass in mature and secondary seasonally dry tropical forests: a literature review and global synthesis. – *Forest Ecology and Management* 276: 88-95.
- [8] Bendix, J., Behling, H., Peters, T., Richter, M., Beck, E. (2010): Functional Biodiversity and Climate Change Along an Altitudinal Gradient in a Tropical Mountain Rainforest. – In: Tschardtke, T., Leuschner, C., Veldkamp, E., Faust, H., Guhardja, E., Bidin, A. (eds.) *Tropical Rainforests and Agroforests under Global Change. Environmental Science and Engineering (Environmental Engineering)*. Springer, Berlin, Heidelberg.
- [9] Bergkamp, L., Goldsmith, B. (eds.) (2013): *The EU Environmental Liability Directive: a Commentary*. – Oxford University Press, Oxford.
- [10] Blondel, J. (2010): *The Mediterranean Region: Biological Diversity in Space and Time*. – Oxford University Press, UK.
- [11] Blondel, J., Aronson, J. (1999): *Biology and Wildlife of the Mediterranean Region*. – Oxford University Press, New York.
- [12] Boberg, A. L., Morris-Khoo, S. A. (1992): The Delphi method: a review of methodology and an application in the evaluation of a higher education program. – *The Canadian Journal of Program Evaluation* 7(1): 27-39.
- [13] Bongaarts, J. (2009): Human population growth and the demographic transition. – *Philosophical Transactions of The Royal Society: Biological Sciences* 364(1532): 2985-2990.
- [14] Bouma, A. J., van Beukering, H. J. P. (2015): *Ecosystem Services: from Concept to Practice*. – Cambridge University Press, Cambridge.
- [15] Bouvet, A., Mermoz, S., Toan, T. L., Villard, L., Mathieu, R., Naidoo, L., Asner, G. P. (2018): An above-ground biomass map of African savannahs and woodlands at 25 m resolution derived from ALOS PALSAR. – *Remote Sensing of Environment* 206: 156-173.
- [16] Bruijnzeel, L. A., Scatena, F. N., Hamilton, L. S. (eds.) (2011): *Tropical Montane Cloud Forests: Science for Conservation and Management*. – Cambridge University Press, Cambridge.
- [17] Bubb, P., May, I., Miles, L., Sayer, J. (2004): *Cloud Forest Agenda*. – UNEP World Conservation Centre, Cambridge.
- [18] Burton, J. P., Messier, C., Smith, W. D., Adamowicz, L. W. (2003): *Towards Sustainable Management of the Boreal Forest*. – NRC Research Press, Ottawa.
- [19] Carbutt, C., Tau, M., Stephens, A., Escott, B. (2011): The conservation status of temperate grasslands in southern Africa. – *Grassroots* 11(1): 17-23.
- [20] Carpenter, S. R., Mooney, H. A., Agard, J., Capistrano, D., DeFries, R. S., Diaz, S., Dietz, T., Duraipah, A. K., Oteng-Yeboah, A., Pereira, H. M., Perrings, C., Reid, W. V., Sarukhan, J., Scholes, R. J., Whyte, A. (2009): Science for managing ecosystem services: beyond the Millennium Ecosystem Assessment. – *Proceedings of the National Academy of Sciences of the United States of America* 106(5): 1305-1312.
- [21] Carson, M. R., Bergstorm, C. J. (2003): *A Review of Ecosystem Valuation Techniques*. – Department of Agricultural and Applied Economics, University of Georgia, Athens.
- [22] Ceschin, S., Cancellieri, L., Caneva, G., Battisti, C. (2012): Size area, patch heterogeneity and plant species richness across archaeological sites of Rome: different patterns for different guilds. – *Vie et Milieu—Life and Environment* 62(4): 165-171.
- [23] Chan, K. M. A., Satterfield, T. (2020): The maturation of ecosystem services: social and policy research expands, but whither biophysically informed valuation? – *People and Nature* 2(4): 1021-60.
- [24] Chan, K. M. A., Shaw, M. R., Cameron, R. D., Underwood, C. E., Daily, C. G. (2006): Conservation planning for ecosystem services. – *PLoS Biology* 4(11): e379.
- [25] Chan, K. M. A., Satterfield, T., Goldstein, J. (2012): Rethinking ecosystem services to better address and navigate cultural values. – *Ecological Economics* 74: 8-18.
- [26] Chee, E. Y. (2004): An ecological perspective on the valuation of ecosystem services. – *Biological Conservation* 120: 549-565.

- [27] Costanza, R. (2000): Social goals and the valuation of ecosystem services. – *Ecosystems* 3: 4-10.
- [28] Costanza, R., Folke, C. (1997): Valuing Ecosystem Services with Efficiency, Fairness and Sustainability as Goals. – In: Daily, C. G. (ed.) *Nature's Services: Societal Dependence on Natural Ecosystems*. Island Press, Washington D. C.
- [29] Costanza, R., Kubiszewski, I. (2012): The authorship structure of “ecosystem services” as a transdisciplinary field of scholarship. – *Ecosystem Services* 1: 16-25.
- [30] Costanza, R., de Groot, R., Sutton, P., van der Ploeg, S., Anderson, J. S., Kubiszewski, I., Farber, S., Turner, R. K. (2014): Changes in the global value of ecosystem services. – *Global Environmental Change* 26: 152-158.
- [31] Costanza, R., de Groot, R., Braat, L., Kubiszewski, I., Fioramonti, L., Sutton, P., Farber, S., Grasso, M. (2017): Twenty years of ecosystem services: How far have we come and how far do we still need to go? – *Ecosystem Services* 28: 1-16.
- [32] Cox, L. R., Underwood, C. E. (2011): The importance of conserving biodiversity outside of protected areas in Mediterranean ecosystems. – *Plos ONE* 6(1): e14508.
- [33] Daily, C. G. (1997): *Nature's Services: Societal Dependence on Natural Ecosystems*. – Island Press, Washington D.C.
- [34] Daily, G. C., Alexander, S., Ehrlich, P. R., Goulder, L., Lubchenco, J., Matson, P. A., Mooney, H. A., Postel, S., Schneider, S. H., Tilman, D., Woodwell, G. M. (1997): Ecosystem services: benefits supplied to human societies by natural ecosystems. – *Issues in Ecology* 1: 1-18.
- [35] Daily, G. G., Polasky, S., Goldstein, J., Kareiva, M. P., Mooney, A. H., Pejchar, L., Ricketts, H. T., Salzman, J., Shallenberger, R. (2009): Ecosystem services in decision-making: time to deliver. – *Frontiers in Ecology and the Environment* 7(1): 21-28.
- [36] Daly, H. H. (2016): *Assessment of the Socio-Economic Value of the Goods and Services Provided by Mediterranean Forest Ecosystems: Critical and Comparative Analysis of Studies Conducted in Algeria, Lebanon, Morocco, Tunisia and Turkey*. – Plan Bleu, Valbonne.
- [37] de Groot, S. R., Wilson, A. M., Boumans, J. M. R. (2002): A typology for the classification, description and valuation of ecosystems functions, goods and services. – *Ecological Economics* 41: 393-408.
- [38] de Groot, S. R., Alkemade, R., Braat, R., Hein, L., Willemsen, L. (2010): Challenges in integrating the concept of ecosystem services and values in landscape planning, management and decision making. – *Ecological Complexity* 7: 260-272.
- [39] de Groot, R., Brander, L., van der Ploeg, S., Costanza, R., Bernard, F., Braat, L., Chitistie, M., Crossman, N., Ghermandi, A., Hein, L., Hussain, S., Kumar, P., McVittie, A., Portela, R., Rodriguez, C. L., ten Brink, P., van Beukering, P. (2012): Global estimates of the value of ecosystems and their services in monetary units. – *Ecosystem Services* 1: 50-61.
- [40] Diaz, H. F., Eischeid, J. K. (2007): Disappearing “alpine tundra” Köppen climatic type in the western United States. – *Geophysical Research Letters* 34(18): L18707.
- [41] Dinerstein, E., Vynne, C., Sala, E., Joshi, A. R., Fernando, S., Lovejoy, T. E., Mayorga, J., Olson, D., Asner, G. P., Baillie, J. E. M., Burgess, N. D., Burkart, K., Noss, R. F., Zhang, Baccini, A., Birch, T., Hahn, N., Joppa, L. N., Wikramanayake, E. (2019): A global deal for nature: guiding principles, milestones, and targets. – *Science Advances* 5(4): eaaw2869.
- [42] Efe, R., Cravins, G., Ozturk, M., Atalay, I. (2008): *Natural Environment and Culture in the Mediterranean Region*. – Cambridge Scholars Publishing, Newcastle.
- [43] Eiserhardt, W. L., Couvreur, T. L. P., Baker, W. J. (2017): Plant phylogeny as a window on the evolution of hyperdiversity in the tropical rainforest biome. – *New Phytologist* 214(4): 1408-1422.
- [44] Ellis, E. C., Ramankutty, N. (2008): Putting people in the map: anthropogenic biomes of the world. – *Frontiers in Ecology and the Environment* 6(8): 439-447.
- [45] Ellis, E. C., Antill, E. C., Kreft, H. (2012): All is not loss: plant biodiversity in the Anthropocene. – *PLoS ONE* 7(1): e30535.

- [46] Erdoğdu, M. M., Arun, T., Ahmad, I. H. (2016): Handbook of Research on Green Economic Development Initiatives and Strategies. – IGI Global, USA.
- [47] European Environmental Agency. (2002): Europe's Biodiversity–Biogeographical Regions and Seas: Biogeographical Regions in Europe. – ZooBoTech HB, Sweden.
- [48] Farber, C. S., Costanza, R., Wilson, A. M. (2002): Economic and ecological concepts for valuing ecosystem services. – *Ecological Economics* 41: 375-392.
- [49] Feld, C. K., da Silva, P. M., Sousa, J. P., De Bello, F., Bugter, R., Grandin, U., Hering, D., Lavorel, S., Mountford, O., Pardo, I., Pärtel, M., Römbke, J., Sandin, L., Jones, B., Harrison, P. (2009): Indicators of biodiversity and ecosystem services: a synthesis across ecosystems and spatial scales. – *Oikos* 118(12): 1862-1871.
- [50] Flexas, J., Loreto, F., Medrano, H. (2012): Terrestrial Photosynthesis in a Changing Environment: A Molecular, Physiological and Ecological Approach. – Cambridge University Press, New York.
- [51] FRA (Forest Resources Assessment) Programme. (2000): Global Ecological Zoning for the Global Forest Resources Assessment 2000. Final Report. – Food and Agriculture Organization of the United Nations, Rome.
- [52] Fu, B-J., Su, C-H., Wei, Y-P., Willett, R. I., Lü, Y-H., Liu, G-H. (2011): Double counting in ecosystem services valuation: causes and countermeasures. – *Ecological Research* 26: 1-14.
- [53] Geist, H. (ed.) (2006): Our Earth's Changing Land: An Encyclopedia of Land-Use and Land-Cover Change. – Greenwood Press, London.
- [54] Gerstner, K., Dormann, F. C., Václavík, T., Kerft, H., Seppelt, R. (2014): Accounting for geographical variation in species–area relationships improves the prediction of plant species richness at the global scale. – *Journal of Biogeography* 41: 261-273.
- [55] Gibbs, H., Yui, S., Plevin, R. (2014): New estimates of soil and biomass carbon stocks for global economic models. – GTAP Technical Paper No. 33.
- [56] Gibson, J. D. (2009): Grasses & Grassland Ecology. – Oxford University Press, Oxford.
- [57] Gilliam, F. S. (2016): Forest ecosystems of temperate climatic regions: from ancient use to climate change. – *New Phytologist* 212(4): 871-887.
- [58] Girardin, C. A. J., Malhi, Y., Aragão, L. E. O. C., Mamani, M., Huaraca Huaraca, W., Durand, L., Feeley, K. J., Rapp, J., Silva-Espejo, J. E., Silman, M., Salinas, N., Whittaker, R. J. (2010): Net primary productivity allocation and cycling of carbon along a tropical forest elevational transect in the Peruvian Andes. – *Global Change Biology* 16(12): 3176-3192.
- [59] Glantz, M. (1999): Creeping Environmental Problems and Sustainable Development in the Aral Sea Basin. – Cambridge University Press, Cambridge.
- [60] Goldewijk, K. K. (2001): Estimating global land use change over the past 300 years: the HYDE database. – *Global Biogeochemical Cycles* 15(2): 417-433.
- [61] Goldewijk, K. K., Beusen, A., Janssen, P. (2010a): Long-term dynamic modeling of global population and built-up area in a spatially explicit way: HYDE 3.1. – *The Holocene* 1-9.
- [62] Goldewijk, K. K., Beusen, A., van Dreht, G., de Vos, M. (2010b): The HYDE 3.1 spatially explicit database of human-induced global land-use change over the past 12,000 years. – *Global Ecology and Biogeography* 20(1): 73-86.
- [63] Gómez-Baggethun, E., Pérez, R. M. (2011): Economic valuation and the commodification of ecosystem services. – *Progress in Physical Geography* 35(5): 613-628.
- [64] Gómez-Baggethun, E., de Groot, R., Lomas, L. P., Montes, C. (2009): The History of ecosystem services in economic theory and practice: from early notions to markets and payment schemes. – *Ecological Economics* 69(6): 1209-1218.
- [65] Gradstein, S. R., Homeier, J., Gansert, D. (2008): The tropical mountain forest: patterns and processes in a biodiversity hotspot. – *Biodiversity and Ecology Series* 2.
- [66] Greiner, A. (2005): The Forces of Economic Growth: A Time Series Perspective. – Princeton University Press, Princeton, USA.

- [67] Haberl, H., Heinz Erb, K., Krausmann, F., Gaube, V., Bondeau, A., Plutzer, C., Gingrich, S., Lucht, W., Fischer-Kowalski, M. (2007): Quantifying and mapping the human appropriation of net primary production in earth's terrestrial ecosystems. – *Proceedings of the National Academy of Sciences* 104(31): 12942-12947.
- [68] Hadley, D., D'Heroncourt, J., Franzén, F., Kinell, G., Söderqvist, T., Soutukorva, Á., Brouwer, R. (2011): Monetary and non-monetary methods for ecosystem services valuation - Specification sheet and supporting material. – *Speciosa Project Report*, University of East England, Norwich.
- [69] Hancock, J. (2010): The case for an ecosystem service approach to decision-making: an overview. – *Bioscience Horizons* 3(2).
- [70] Heal, G. (1999): Valuing Ecosystem Services. – *Ecosystems* 3(1): 24-30.
- [71] Hein, L., van Koppen, K., de Groot, S. R., van Ierland, C. E. (2006): Spatial scales, stakeholders and the valuation of ecosystem services. – *Ecological Economics* 57: 209-228.
- [72] Hoekstra, J. M., Boucher, T. M., Ricketts, T. H., Roberts, C. (2005): Confronting a biome crisis: global disparities of habitat loss and protection. – *Ecology Letters* 8(1): 23-29.
- [73] Houghton, R. A., Hall, F., Goetz, S. J. (2009): Importance of biomass in the global carbon cycle. – *The Journal of Geophysical Research-Biogeosciences* 114: G00E03.
- [74] Howarth, B. R., Farber, S. (2002): Accounting for the value of ecosystem services. – *Ecological Economics* 41: 421-429.
- [75] Hufnagel, L., Mics, F., Pálkás, M., Homoródi, R. (2018): Introductory Chapter: Global Aspects and Scientific Importance of Desert Ecological Research. – In: Hufnagel, L. (ed.): *Community and Global Ecology of Deserts*. IntechOpen, London. DOI: 10.5772/intechopen.78368.
- [76] Iremonger, S., Ravilious, C., Quinlan, T. (eds.) (1997): *A Global Overview of Forest Conservation*. – World Conservation Monitoring Centre and Center for International Forestry Research, Cambridge, UK.
- [77] Ito, A., Oikawa, T. (2004): Global Mapping of Terrestrial Primary Productivity and Light-Use Efficiency with a Process-Based Model. – In: *Global Environmental Change in the Ocean and on Land*. ERRAPUB, Tokyo, pp. 343-358.
- [78] Kapos, V., Rhind, J., Edwards, M., Price, M. F., Ravilious, C., Butt, N. (2000): Developing a map of the world's mountain forests. *Forests in sustainable mountain development: a state of knowledge report for 2000*. – Task Force on Forests in Sustainable Mountain Development, pp. 4-19.
- [79] Keho, Y. (2017): Revisiting the income, energy consumption and carbon emissions nexus. – *International Journal of Energy Economics and Policy* 7(3): 356-363.
- [80] Keith, H., Mackey, B. G., Lindenmayer, D. B. (2009): Re-evaluation of forest biomass carbon stocks and lessons from the world's most carbon-dense forests. – *Proceedings of the National Academy of Sciences of the United States of America* 106(28): 11635-11640.
- [81] Kier, G., Mutke, J., Dinerstein, E., Ricketts, T. H., Küper, W., Kreft, H., Barthlott, W. (2005): Global patterns of plant diversity and floristic knowledge. – *Journal of Biogeography* 32: 1-10.
- [82] Koetse, M. J., Agarwala, M., Bullock, C., Ten Brink, P. (2015): *Monetary and Social Valuation: State-of-the-Art*. – Institute for Environmental Studies (IVM), VU University Amsterdam, the Netherlands (report prepared for EU 7th framework project OPERAs).
- [83] Komen, M. H. C., Gerking, S., Folmer, H. (1997): Income and environmental R&D: empirical evidence from OECD countries. – *Environment and Development Economics* 2: 505-515.
- [84] Kroeger, M. (2012): *Global Tree Plantation Expansion: a Review*. – ICAS Review Paper Series No. 3.
- [85] Kumar, M., Kumar, P. (2008): Valuation of the ecosystem services: a psycho-cultural perspective. – *Ecological Economics* 64: 808-819.

- [86] Lal, R., Lorentz, K. (2012): Carbon Sequestration in Temperate Forests. – In: Lal, R., Lorentz, K., Hüttl, R. F., Schneider, B. U., von Braun, J. (eds.) Recarbonization of the Biosphere: Ecosystems and the Global Carbon Cycle. Springer, Dordrecht.
- [87] Loomis, J., Kent, P., Strange, L., Fausch, K., Covich, A. (2000): Measuring the total economic value of restoring ecosystem services in an impaired river basin: results from a contingent valuation survey. – *Ecological Economics* 33: 103-117.
- [88] Loomis, J., Richardson, L., Kroeger, T., Casey, F. (2014): Valuing Ecosystem Services Using Benefit Transfer: Separating Credible and Incredible Approaches. – In: Ninan, K. N. (ed.) Valuing Ecosystem Services: Methodological Issues and Case Studies. Edward Elgar Publishing Limited, Cheltenham, UK.
- [89] Luysaert, S., Inglima, I., Jung, M., Richardson, A. D., Reichsteins, M., Papale, D., Piao, S. L., Schulzes, E. D., Wingate, L., Matteucci, G., Aragao, L., Aubinet, M., Beers, C., Bernhoffer, C., Black, K. G., Bonal, D., Bonnefond, J.-M., Chambers, J., Ciais, P., Cook, B., Davis, K. J., Dolman, A. J., Gielen, B., Goulden, M., Grace, J., Granier, A., Grelle, A., Griffis, T., Gruenwald, T., Guidolotti, G., Hanson, P. J., Harding, R., Hollinger, D. Y., Hutyrá, L. R., Kolar, P., Kruijt, B., Kutsch, W., Lagergren, F., Laurila, T., Law, B. E., Lindroth, A., Loustau, D., Malhi, Y., Mateus, J., Migliavacca, M., Misson, L., Montagnani, L., Moncrieff, J., Moors, E., Munger, J. W., Nikinmaa, E., Ollinger, S. V., Pita, G., Rebmann, C., Roupsard, O., Saigusa, N., Sanz, M. J., Seufert, G., Sierra, C., Smith, M.-L., Tang, J., Valentini, R., Vesala, T., Janssens, I. A. (2007): CO<sub>2</sub> balance of boreal, temperate, and tropical forests derived from a global database. – *Global Change Biology* 13: 2509-2537.
- [90] Mark, F. A., McLennan, B. (2005): The conservation status of New Zealand's indigenous grasslands. – *New Zealand Journal of Botany* 43: 245-270.
- [91] Marsiliani, L., Renstrom, T. I. (2000): Inequality, Environmental Protection and Growth. – CentER Discussion Paper 2000-34.
- [92] Mics, F., Rozak, A. H., Kocsis, M., Homoródi, R., Hufnagel, L. (2013): Rainforests at the beginning of the 21<sup>st</sup> Century. – *Applied Ecology and Environmental Research* 11(1): 1-20.
- [93] Miles, L., Newton, A. C., DeFries, R. S., Ravilious, C., May, I., Blyth, S., Kapos, V., Gordon, J. E. (2006): A global overview of the conservation status of tropical dry forests. – *Journal of Biogeography* 33(3): 491-505.
- [94] Mushafiq, M., Prusak, B. (2023): Nexus between stock markets, economic strength, R&D and environmental deterioration: new evidence from EU-27 using PNARDL approach. – *Environmental Science and Pollution Research* 30: 2965-32984.
- [95] Nabuurs, G. J., Ravindranath, N. H., Paustian, K., Freibauer, A., Hohenstein, W., Makundi, W., Bickel, K. (2003): Chapter 3. LUCF Sector Good Practice Guidance. – In: Penman, J., Gytarsky, M., Hiraishi, T., Krug, T., Kruger, D., Pipatti, R., Buendia, L., Miwa, K., Ngara, T., Tanabe, K., Wagner, F. (eds.) IPCC Good Practice Guidance for Land Use Change and Forestry. Kanagawa, Japan: IPCC National Greenhouse Gas Inventories Programme.
- [96] National Research Council (2005): Valuing Ecosystem Services: Toward Better Environmental Decision-Making. – The National Academic Press, Washington D.C.
- [97] Nelson, E., Mendoza, G., Regetz, J., Polasky, S., Tallis, H., Cameron, D. R., Chan, M. K., Daily, C. G., Goldstein, J., Kareiva, M. P., Lonsdorf, E., Naidoo, R., Ricketts, H. T., Shaw, M. R. (2009): Modeling multiple ecosystem services, biodiversity conservation, commodity production, and tradeoffs at landscape scales. – *Frontiers in Ecology and the Environment* 7(1): 4-11.
- [98] Newell, P., Roberts, T. J. (2017): The Globalization and Environmental Reader. – John Wiley & Sons Ltd., Chichester.
- [99] Oliveras, I., van der Eynden, M., Malhi, Y., Cahuana, N., Menor, C., Zamora, F., Haugaasen, T. (2013): Grass allometry and estimation of aboveground biomass in tropical alpine tussock grassland. – *Austral Ecology* 39(4): 408-415.

- [100] Olson, D. M., Dinerstein, E., Wikramanayake, E. D., Burgess, N. D., Powell, G. V. N., Underwood, E. C., D'Amico, J. A., Itoua, I., Strand, H. E., Morrison, J. C., Loucks, C. J., Allnutt, T. F., Ricketts, T. H., Kura, Y., Lamoreux, J. F., Wettengel, W. W., Hedao, P., Kassem, K. R. (2001): Terrestrial ecoregions of the world: a new map of life on Earth. – *Bioscience* 51(11): 933-938.
- [101] Pan, Y., Birdsey, A. R., Phillips, L. O., Jackson, B. R. (2013): The structure, distribution, and biomass of the world's forests. – *Annual Review of Ecology, Evolution and Systematics* 44: 593-622.
- [102] Park, C. C. (2003): *Tropical Rainforests*. – Routledge, London and New York.
- [103] Pascual, U., Muradian, R., Brander, L., Gómez-Baggethun, E., Martín-López, B., Verma, M., Armsworth, P., Christie, M., Cornelissen, H., Eppnik, F., Farley, J., Loomis, J., Pearson, L., Perrings, C., Polasky, S. (2009): *The Economics of Ecosystems and Biodiversity: The Ecological and Economic Foundations*. – Earthscan, London and Washington D.C.
- [104] Pascual, U., Balvanera, P., Díaz, S., Pataki, Gy., Roth, E., Stenseke, M., Watson, R. T., Dessane, E. B., Islar, M., Kelemen, E., Maris, V., Quaas, M., Subramanian, S. M., Wittmer, H., Adlan, A., Ahn, S., Al-Hafedh, Y. S., Amankwah, E., Asah, S. T., Berry, P., Bilgin, A., Breslow, S. J., Bullock, C., Cáceres, D., Daly-Hassen, H., Figueroa, E., Golden, C. D., Gómez-Baggethun, E., González-Jiménez, D., Houdet, J., Keune, H., Kumar, R., Ma, K., May, P. H., Mead, A., O'Farrell, P., Pandit, R., Pengue, W., Pichis-Madruga, R., Popa, F., Preston, S., Pacheco-Balanza, D., Saarikoski, H., Strassburg, B. B., Van den Belt, M., Verma, M., Wickson, F., Yagi, N. (2017): Valuing nature's contributions to people: the IPBES approach. – *Current Opinion in Environmental Sustainability* 26: 7-16.
- [105] Peart, B. (2008): *Compendium of Regional Templates on the Status of Temperate Grasslands Conservation and Protection*. – IUCN World Commission on Protected Areas, Vancouver, Canada.
- [106] Pinborg, U. (2002): *Europe's Biodiversity—Biogeographical Regions and Seas*. – The European Environment Agency, Copenhagen.
- [107] Polidoro, B. A., Carpenter, K. E., Collins, L., Duke, N. C., Ellison, A. M., Ellison, J. C., Farnsworth, J. E., Fernando, S. E., Kathiresan, K., Koedam, E. N., Livingstone, R. S., Miyagi, T., Moore, E. G., Nam, N. V., Ong, E. J., Primavera, H. J., Salmo, G. S., Sanciangco, C. J., Sukardjo, S., Wang, Y., Yong, H. W. J. (2010): The loss of species: mangrove extinction risk and geographic areas of global concern. – *PloS ONE* 5(4): e10095.
- [108] Potapov, P., Turubanova, S., Hansen, M. C., Tyukavina, A., Zalles, V., Khan, A., Song, X.-P., Pickens, A., Shen, Q., Cortez, J. (2022): Global maps of cropland extent and change show accelerated cropland expansion in the twenty-first century. – *Nature Food* 3: 19-28.
- [109] Proenca, M. V., Pereira, M. H., Guilherme, J., Vicente, L. (2010): Plant and bird diversity in natural forests and in native and exotic plantations in NW Portugal. – *Acta Oecologica* 36: 219-226.
- [110] Román-Cuesta, R. M., Salinas, N., Asbjornsen, H., Oliveras, I., Huaman, V., Gutiérrez, Y., Puelles, L., Kala, J., Yabar, D., Rojas, M., Astete, R., Jordán, D. Y., Silman, M., Mosondl, R., Weber, M., Stimm, B., Günter, S., Knoke, T., Malhi, Y. (2011): Implications of fires on carbon budgets in Andean cloud montane forest: the importance of peat soils and tree resprouting. – *Forest Ecology and Management* 261(11): 1987-1997.
- [111] Saugier, B., Roy, J., Mooney, H. A. (2001): Estimations of global terrestrial productivity: converging toward a single number. – *Terrestrial Global Productivity*, pp. 543-557.
- [112] Sherrouse, C. B., Clement, M. J., Semmens, J. D. (2011): A GIS application for assessing, mapping, and quantifying the social values of ecosystem services. – *Applied Geography* 31: 748-760.
- [113] Silander, A. J. (2001): *Encyclopedia of Biodiversity: Volume 5*. – Academic Press, USA.

- [114] Sloan, S., Jenkins, C. N., Joppa, L. N., Gaveau, D. L. A., Laurance, W. F. (2014): Remaining natural vegetation in the global biodiversity hotspots. – *Biological Conservation* 177: 12-24.
- [115] Spracklen, D. V., Righelato, R. (2014): Tropical montane forests are a larger than expected global carbon store. – *Biogeosciences* 11(10): 2741-2754.
- [116] Steffen, W., Crutzen, P. J., McNeill, J. R. (2007): The Anthropocene: Are humans now overwhelming the great forces of nature? – *AMBIO: A Journal of the Human Environment* 36(8): 614-621.
- [117] Sun, Y., Feng, Y., Wang, Y., Zhao, X., Yang, Y., Tang, Z., Wang, S., Su, H., Zhu, J., Chang, J., Fang, J. (2021): Field-based estimation of net primary productivity and its above- and belowground partitioning in global grasslands. – *Journal of Geophysical Research: Biogeosciences* 126: e2021JG006472.
- [118] Tallis, H., Kareiva, P. (2005): Ecosystem services. – *Current Biology* 15(18): R746.
- [119] Tamazian, A., Rao, B. B. (2010): Do economic, financial and institutional developments matter for environmental degradation? Evidence from transitional economies. – *Energy Economics* 32(1): 137-145.
- [120] Ten Brink, P. (2011): *The Economics of Ecosystems and Biodiversity in National and International Policy Making*. – Earthscan, USA.
- [121] Thornley, J. H. M., Cannell, M. G. R. (2000): Managing forests for wood yield and carbon storage: a theoretical study. – *Tree Physiology* 20(7): 477-484.
- [122] Thurner, M., Beer, C., Santoro, M., Carvalhais, N., Wutzler, T., Schepaschenko, D., Shvidenko, A., Kompter, E., Ahrens, B., Levick, S. R., Schullius, C. (2014): Carbon stock and density of northern boreal and temperate forests. – *Global Ecology and Biogeography* 23: 297-310.
- [123] Tiwari, S. K. (2013): Sustainable development and protection of environment in India: judicial perspective. – *Indian Journal of Law and Justice* 4(2): 91-110.
- [124] Triantis, K. A., Vardinoyannis, K., Tsolaki, E. P., Botsaris, I., Lika, K., Mylonas, M. (2006): Re-approaching the small island effect. – *Journal of Biogeography* 33(5): 914-923.
- [125] Valiela, I. (2009): *Global Coastal Change*. – Blackwell Publishing, Oxford.
- [126] Vo, T. Q., Kuenzer, C., Vo, M. Q., Moder, F., Oppelt, N. (2012): Review of valuation methods for mangrove ecosystem services. – *Ecological Indicators* 23: 431-446.
- [127] Wang, L., Tian, F., Wang, Y., Wu, Z., Schurgers, G., Fensholt, R. (2018): Acceleration of global vegetation greenup from combined effects of climate change and human land management. – *Global Change Biology* 24(11): 5484-5499.
- [128] Whittaker, R. H., Likens, G. E. (1973): Carbon in the biota. – *Brookhaven Symposia in Biology* 30: 281-302.
- [129] Wilkinson, D. M. (1999): The disturbing history of intermediate disturbance. – *Oikos* 84(1): 145-7.
- [130] Williams, B. A., Venter, O., Allan, J. R., Atkinson, S. C., Rehbein, J. A., Ward, M., Di Marco, M., Grantham, H. S., Ervin, J., Goetz, S. J., Hansen, A. J., Jantz, P., Pillay, R., Rodríguez-Buritica, S., Supples, C., Virnig, A. L. S., Watson, J. E. M. (2020): Change in terrestrial human footprint drives continued loss of intact ecosystems. – *One Earth* 3(3): 371-382.
- [131] Wilson, A. M., Howarth, B. R. (2002): Discourse-based valuation of ecosystem services: establishing fair outcomes through group deliberation. – *Ecological Economics* 41: 431-443.
- [132] Wittmer, H., Gundimeda, H. (2012): *The Economics of Ecosystems and Biodiversity in Local and Regional Policy and Management*. – Earthscan, USA, Canada.
- [133] Woodward, F. I., Lomas, M. R., Kelly, C. K. (2004): Global climate and the distribution of plant biomes. – *Philosophical Transactions of The Royal Society: Biological Sciences* 359(1450): 1465-1476.

- [134] Yee, M. S. (2010): REDD and BLUE Carbon: Carbon Payments for Mangrove Conservation. – Center for Marine Biodiversity and Conservation, UC San Diego, San Diego, CA.
- [135] Zahran, A. M., Gilbert, F. (2010): Climate-Vegetation: Afro-Asia Mediterranean and Red Sea Coastal Lands. – Springer, New York.
- [136] Živanović, V. (2016): The poles of growth and development as determinants of spatial and functional structure of Podrinje Region. – Zbornik radova - Geografski fakultet Univerziteta u Beogradu 64: 221-254.

## APPENDIX

Limitations, ecological aspects not considered, additional data, and suggestions for further development of the assessment system.

### *Content of appendices*

A.1. The comparative assessment of continents omitted Antarctica (Additional information and references).

A.2. The role of mosses and lichens was not taken into account in the vegetation assessment (Additional information and references).

A.3. Animal biomass, production, and biodiversity not taken into account.

A.4. We have not investigated the role of microorganisms.

A.5. We have not taken into account the seas and oceans, and to a limited extent the mangrove habitat.

### **A.1. The comparative assessment of continents omitted Antarctica**

The ice-free area of Antarctica represents only 0.5% of the continent, approximately 332,000 km<sup>2</sup> (Claridge et al., 2000). This area allows for the development of flora. Unlike other continents, vascular plants do not dominate here, making one of our indicators unsuitable. Due to the cold climate, mosses and lichens are the primary vegetation, with only two vascular plant species present (*Deschampsia antarctica*, *Colobanthus quitensis*) (Convey, 2001; Øvstedal and Lewis Smith, 2001). These species are mainly found on the peninsula, South Shetland Islands, and South Orkney Islands, where climate conditions are milder. Their biomass and production vary greatly depending on microclimatic, soil, and topographic conditions (Holdgate, 1964; Holtom and Greene, 1967; Edwards, 1972; Vera et al., 2013).

Microflora, including blue-green algae, eukaryotic algae, and fungi, are also significant. The fauna mainly consists of invertebrates. The continent can be divided into three biogeographical zones: surrounding islands, coastal areas, and the continental interior (Smith, 1984; Longton, 1988; Chown and Convey, 2007). Climatic conditions vary slightly, with the surrounding islands having the mildest climate, resulting in richer flora and fauna. As the climate becomes more extreme, species richness and biomass decrease. Overall, diversity is lower than in similar latitudes of northern tundras (Rønning, 1996; Nielsen and Wall, 2013). *Table A.1.1* shows the species richness of these three regions.

The average phytomass on continental land ranges from 5-200 g/m<sup>2</sup>, while in oceanic areas it is between 300-1000 g/m<sup>2</sup> for living, photosynthesizing tissues. When considering

the accumulation of dead tissues, it can reach up to 20-30 kg/m<sup>2</sup>. Peat formation can reach 1 mm per year, and accumulating peat can be used to study climate change over thousands of years (Longton, 1992; Royles and Griffiths, 2015).

**Table A.1.1.** Antarctic biodiversity based on Convey (2006)

OTU	Sub-Antarctic Islands	Maritime Antarctica	Continental Antarctica
Protozoa	83		33
Rotifera	59	50	13
Tardigrada	34	26	19
Nematoda	22	28	14
Platyhelminthes	4	2	0
Gastrotricha	5	2	0
Annelida	23	3	0
Mollusca	4	0	0
Crustacea	48	10	14
Insecta	210	35	49
Collembola	30	10	10
Arachnida	167	36	29
Myriopoda	3	0	0
Flowering plants/Angiosperms	60	2	0
Ferns/Pteridophyta	16	0	0
Bryophyta	250	100	25
Marchantiophyta	85	25	1
Lichenophyta	250	250	150
Fungi	70	30	0
<b>Total</b>	<b>1423</b>	<b>609</b>	<b>357</b>

For mosses, Net Primary Productivity (NPP) ranges from 5-100 g/m<sup>2</sup>/year in continental climates, and from 300-650 g/m<sup>2</sup>/year in more favourable oceanic climates (Kennedy, 1995). Lichen biomass in continental areas ranges from 46-177 g/m<sup>2</sup> (Friedmann, 1982), with an NPP of only 5 mg/m<sup>2</sup>/year (Vestal, 1988). Near Birthday Ridge (70°8' 48°0' S), combined moss and lichen biomass ranges from 50-950 g/m<sup>2</sup> (Allan Green et al., 2007). In oceanic climates, biomass values are 800-1750 g/m<sup>2</sup>, with an NPP of 250 g/m<sup>2</sup>/year (Smith, 1984). Near the Spanish base (Juan Carlos I), Beltrán-Sanz et al. (2022) found an average of 92.2 mg CO<sub>2</sub> gDW<sup>-1</sup> per year from 2009-2014. At McMurdo Dry Valleys, Geyer et al. (2017) measured 217 g C/m<sup>2</sup>/year at a site with 60 days available for growth. Growth rates for lichens vary significantly, from 0.47–0.50 mm/year on Signy and Livingston Islands to less than 0.01 mm/year in the Dry Valleys (Sancho et al., 2019).

In the more northern islands, several vascular plants occur. On South Georgia Island, vegetation includes dry grasslands (*Festuca contracta*), moist grasslands (*Deschampsia antarctica*), oligotrophic peatlands (*Rostkovia magellani*), eutrophic peatlands (*Tortula robusta* and *Juncus scheuchzerioides*), cushion plants (*Acaena magellanica*), and

tussock grasses (*Poa flabellata*) (Holdgate, 1977). In *Festuca contracta* grasslands, above-ground biomass averages 425 g/m<sup>2</sup>, and NPP can reach 340 g/m<sup>2</sup>/year. In *Poa flabellata* grasslands, values reach 7525 g/m<sup>2</sup> and 5020 g/m<sup>2</sup>/year. On Signy Island, *Deschampsia antarctica* stands reach 327 g/m<sup>2</sup> for biomass and 390 g/m<sup>2</sup>/year for NPP (Holdgate, 1977).

On Marion Island, Smith (2008) measured biomass between 173-1245 g/m<sup>2</sup>, with the lower value for mire vegetation and the higher for tussock grassland. NPP ranged from 266-949 g/m<sup>2</sup>/year, with the lower value for fellfield and the higher for tussock grassland. Climate change may lead to an expansion of ice-free areas, allowing invasive species to threaten existing ecosystems (Frenot et al., 2005).

Due to the climate, biodiversity is much lower. Convey (2006) summarised the species found on the continent, showing that there are more species on the milder northern islands. There are only 1423 species on the subantarctic islands, including both animals and plants. This is because the area is much smaller compared to the Arctic tundras. On the continent, the limited extent of ice-free areas restricts the biodiversity of flora and fauna.

Mosses, lichens, and areas with vascular plants provide habitat for invertebrates. On Signy Island, invertebrate biomass can reach 8.5 g/m<sup>2</sup>, and on South Georgia Island, it can reach 17 g/m<sup>2</sup>, with production of 10 g/m<sup>2</sup>/year (Heal and MacLean, 1975). Bacterial biomass in soil ranges from 0.04-0.63 µg/g on South Georgia Island to 2-12 µg/g on Signy Island (Rosswall and Heal, 1975). Díaz-Puente et al. (2021) measured soil microbial carbon on Deception Island, King George Island, and Cierva Point, finding values of 33.55 mg/kg, 75.69 mg/kg, and 134.96 mg/kg, respectively. Areas covered with moss and lichen had higher microbial biomass and carbon content.

The metabolic quotient [q(CO<sub>2</sub>)] was 7.16, 3.91, and 12.64, highest in vegetated areas. For comparison, in tropical rainforests in the Amazon, the microbial carbon content was 1287 µg/g in the upper 5 cm of soil, which decreased after deforestation (Luizao et al., 1992). In Venezuela, López-Hernández (2017) found 98-218 mg/g of microbial carbon in primary forest soils. In desert soil, 6.3 mmol C per kg was measured, equivalent to 0.0756 g/kg of soil (Xu et al., 2013).

Studying microbial diversity is challenging because many microorganisms in extreme environments are not cultivable in labs (Merino et al., 2019). Molecular technologies like pyrosequencing are helpful in such studies. In McMurdo Valley, Thompson et al. (2020) found 71 families and 90 genera of phagotrophic protists, and 32 genera and 23 families of ciliates using molecular techniques. Archer et al. (2017) found 501 Operational Taxonomic Units (OTUs) in sandstone and granite using the 16S rRNA method. Sandstone showed higher diversity due to its porous structure, which favours microbial communities. The orientation of the rock also affects which groups dominate, as it influences radiation exposure.

In the Mitchell Peninsula, Ji et al. (2016) collected 93 soil samples and identified 6928 OTUs across 40 phyla, with higher diversity due to better climate and nutrient content. In contrast, Lopatina et al. (2013) found fewer OTUs in snow samples near the Russian bases Druzhnaya and Leningradskaya. Surprisingly, a *Pseudomonas* species grew at 37°C, indicating it is not a psychrophile. It likely arrived in Antarctica in an inactive form.

Kim et al. (2007) compared Antarctic soils to tropical soils and found 396 OTUs in terra preta soil in the Jamari National Forest, Amazon, using the oligonucleotide fingerprint grouping (OFRG) method.

## REFERENCES OF ANTARCTICA

- [1] Allan Green, T. G., Schroeter, B., Sancho, L. G. (2007): Plant Life in Antarctica. – In: Pugnaire, F., Valladares, F. (eds.) Functional Plant Ecology. CRC Press, Boca Raton.
- [2] Archer, S. D. J., de los Ríos, A., Lee, K. C., Niederberger, T. S., Craig Cary, S., Coyne, K. J., Douglas, S., Lacap-Bugler, D. C., Pointing, S. B. (2017): Endolithic microbial diversity in sandstone and granite from the McMurdo Dry Valleys, Antarctica. – Polar Biology 40: 997-1006.
- [3] Beltrán-Sanz, N., Raggio, J., Gonzalez, S., Dal Grande, F., Prost, S., Green, A., Pintado, A., Sancho, L. G. (2022): Climate change leads to higher NPP at the end of the century in the Antarctic Tundra: response patterns through the lens of lichens. – Science of the Total Environment 835: 155495.
- [4] Chown, S. L., Convey, P. (2007): Spatial and temporal variability across life's hierarchies in the terrestrial Antarctic. – Philosophical Transactions of the Royal Society B 362: 2307-2331.
- [5] Claridge, G. G. C., Campbell, I. B., Sheppard, D. S. (2000): Carbon pools in Antarctica and their significance for global climate change. – In: Lal, R., Kimble, J. M., Stewart, B. A. (eds.) Global climate change and cold regions ecosystems. Boca Raton, FL.
- [6] Convey, P. (2001): Antarctic ecosystems. – In: Levin, S. (ed.) Encyclopedia of Biodiversity. Academic Press, San Diego.
- [7] Convey, P. (2006): Antarctic terrestrial ecosystems: responses to environmental change. – Polarforschung 75(2-3): 101-111.
- [8] Díaz-Puente, F. J., Schmid, T., Pelayo, M., Rodríguez-Rastrero, M., Sierra Herraiz, M. J., O'Neill, T., López-Martínez, J. (2021): Abiotic factors influencing soil microbial activity in the northern Antarctic Peninsula region. – Science of the Total Environment 750: 141602.
- [9] Edwards, J. A. (1972): Studies in *Colobanthus quitensis* (Kunth) Bartl. and *Deschampia antarctica* Desv.: V. Distribution, ecology and vegetative performance on Signy Island. – British Antarctic Survey Bulletin 28: 11-28.
- [10] Frenot, Y., Chown, S. L., Whinam, J., Selkirk, P. M., Convey, P., Skotnicki, M., Bergstrom, D. M. (2005): Biological invasions in the Antarctic: extent, impacts and implications. – Biological Reviews 80(1): 45-72.
- [11] Friedmann, E. I. (1982): Endolithic microorganisms in the Antarctic cold desert. – Science 215: 1045-1053.
- [12] Geyer, K. M., Takacs-Vesbach, C. D., Gooseff, M. N., Barrett, J. E. (2017): Primary productivity as a control over soil microbial diversity along environmental gradients in a polar desert ecosystem. – PeerJ 25(5): e3377.
- [13] Heal, O. W., MacLean, S. F. (1975): Comparative Productivity In Ecosystems—Secondary Productivity. – In: van Dobben, W. H., Lowe-McConnell, R. H. (eds.) Unifying Concepts in Ecology. Junk, The Hague.
- [14] Holdgate, M. W. (1964): Terrestrial Ecology in the Maritime Antarctic. – In: Carrick, R., Holdgate, M. W., Prevost, J. (eds.) Biologie Antarctique. Hermann, Paris.
- [15] Holdgate, M. W. (1977): Terrestrial Ecosystems in the Antarctic. – Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences 279: 5-25.
- [16] Holtom, A., Greene, S. W. (1967): The growth and reproduction of Antarctic flowering plants. – Philosophical Transactions of the Royal Society B 252: 323-328.
- [17] Ji, M., van Dorst, J., Bissett, A., Brown, M. V., Palmer, A. S., Snape, I., Siciliano, D., Ferrari, B. C. (2016): Microbial diversity at Mitchell Peninsula, Eastern Antarctica: a potential biodiversity “hotspot”. – Polar Biology 39: 237-249.
- [18] Kennedy, A. D. (1995): Antarctic terrestrial ecosystem response to global environmental change. – Annual Review of Ecology and Systematics 26: 683-704.

- [19] Kim, J.-S., Sparovek, G., Longo, R. M., Jose De Melo, W., Crowley, D. (2007): Bacterial diversity of terra preta and pristine forest soil from the Western Amazon. – *Soil Biology & Biochemistry* 39: 684-690.
- [20] Longton, R. E. (1988): *Biology of Polar Bryophytes and Lichens*. – Cambridge University Press, Cambridge.
- [21] Longton, R. E. (1992): *The Role of Bryophytes and Lichens in Terrestrial Ecosystems*. – In: Bates, J. W., Farmer, A. M. (eds.) *Bryophytes and Lichens in a Changing Environment*. Clarendon Press, Oxford.
- [22] Lopatina, A., Krylenkov, V., Severinov, K. (2013): Activity and bacterial diversity of snow around Russian Antarctic stations. – *Research in Microbiology* 164: 949-958.
- [23] López-Hernández, D. (2017): Soil quality indexes response to land use change in Puerto Ayacucho, Venezuelan Amazonia. – *Annals of Advanced Agricultural Sciences* 1(2).
- [24] Luizao, R. C. C., Bonde, T. A., Rosswall, T. (1992): Seasonal variation of soil microbial biomass—the effects of clearfelling a tropical rainforest and establishment of pasture in the central Amazon. – *Soil Biology and Biochemistry* 24(8): 805-813.
- [25] Merino, N., Aronson, H. S., Bojanova, D. P., Feyhl-Buska, J., Wong, M. L., Zhang, S., Giovannelli, D. (2019): Living at the extremes: extremophiles and the limits of life in a planetary context. – *Frontiers in Microbiology* 10: 780.
- [26] Nielsen, U. N., Wall, D. H. (2013): The future of soil invertebrate communities in polar regions: different climate change responses in the Arctic and Antarctic? – *Ecology Letters* 16(3): 409-419.
- [27] Øvstedal, D. O., Lewis Smith, R. I. (2001): *Lichens of Antarctica and South Georgia: A Guide to Their Identification and Ecology*. – Cambridge University Press, Cambridge.
- [28] Rønning, O. I. (1996): *The Flora of Svalbard*. – Norsk Polarinstitut, Oslo.
- [29] Rosswall, T., Heal, O. W. (1975): The IBP tundra biome - an introduction. – *Ecological Bulletins* 20: 7-16.
- [30] Royles, J., Griffiths, H. (2015): Invited review: climate change impacts in polar-regions: lessons from Antarctic moss bank archives. – *Global Change Biology* 21(3): 1041-1057.
- [31] Sancho, L. G., Pintado, A., Allan Green, T. G. (2019): Antarctic studies show lichens to be excellent biomonitors of climate change. – *Diversity* 11: 42.
- [32] Smith, R. I. L. (1984): *Terrestrial Biology of the Antarctic and sub-Antarctic*. – In: Laws, R. M. (ed.) *Antarctic Ecology*. Academic Press, London.
- [33] Smith, V. R. (2008): *Terrestrial and Freshwater Primary Production and Nutrient Cycling*. – In: Chown, S. L., Froneman, P. W. (eds.) *The Prince Edward Islands Land-Sea Interactions in a Changing Ecosystem*. SUN PRESS, Baskerville.
- [34] Thompson, A. R., Geisen, S., Adams, B. J. (2020): Shotgun metagenomics reveal a diverse assemblage of protists in a model Antarctic soil ecosystem. – *Environmental Microbiology* 22(11): 4620-4632.
- [35] Vera, M. L., Fernández-Teruel, T., Quesada, A. (2013): Distribution and reproductive capacity of *Deschampsia antarctica* and *Colobanthus quitensis* on Byers Peninsula, Livingston Island, South Shetland Islands, Antarctica. – *Antarctic Science* 25(2): 292-302.
- [36] Vestal, J. R. (1988): Carbon metabolism of the cryptoendolithic microbiota from the Antarctic desert. – *Applied and Environmental Microbiology* 54: 960-965.
- [37] Xu, X., Thornton, P. E., Post, W. M. (2013): A global analysis of soil microbial biomass carbon, nitrogen and phosphorus in terrestrial ecosystems. – *Global Ecology and Biogeography* 22: 737-749.

## A.2. The roles of mosses and lichens was not taken into account in the vegetation assessment

Vascular plants have the highest biomass, but non-vascular plants also contribute significantly to biomass and carbon storage. They provide essential ecosystem services

and play a crucial role in mitigating climate change (Blok et al., 2011). These groups include Bryophyta, Marchantiophyta, Anthocerotophyta, and Lichenophyta. They occur in most terrestrial ecosystems and less often in aquatic environments. The more extreme the habitat, the more important their role and contribution to vegetation biomass. In extreme areas, higher-order plants cannot survive due to harsh conditions.

Lichens can photosynthesize and assimilate carbon, playing a role similar to plants. They are symbiotic organisms composed of algae and fungi. Lichens are highly tolerant of changing environmental conditions. The fungal partner is 98% Ascomycota and 2% Basidiomycota. In 10% of lichens, cyanobacteria are the partner, and in 90%, eukaryotic algae are the partner (Honegger, 2009). Lichens can survive in extremely low temperatures in Antarctica and tolerate desiccation for long periods.

Lichens lack specialized organs and absorb water and nutrients over their entire surface, but they also lose water through the surface. This limits their growth conditions. Without protection, they dry out quickly, and their physiological activity stops, leading to slow growth. Lichens also absorb toxic substances from the environment if unprotected. Due to their stress tolerance, they can survive on rock surfaces with few competitors. Their sensitivity to pollution makes them good ecological indicators.

Mosses have leaf- and root-like structures but lack vascular tissues. They do have cells for transport, though these are not like the tracheae of higher plants. Like lichens, mosses are highly resistant to desiccation and are poikilohydric. They have a haploid-diploid life cycle with a dominant gametophyte phase. Mosses are the closest living relatives of the first land plants. They bridge modern vascular plants and their algal ancestors, representing a transition to land plants (Kenrick and Crane, 1997).

In dry and cold areas, carbon uptake is about 16 g/m<sup>2</sup>/year, equivalent to 16 t/km<sup>2</sup>/year. Using the value of 170 t/km<sup>2</sup>/year from Girardin et al. (2010), this results in a rainforest equivalent of 0.0941. In areas covered with herbaceous plants and grasses, non-vascular plants have a carbon uptake of 23 g/m<sup>2</sup>/year, which equals 0.1352 rainforest equivalent. In areas with no soil and only rock surfaces, carbon uptake is 8 g/m<sup>2</sup>/year (Elbert et al., 2009), corresponding to 0.0471 rainforest equivalent. The epiphytic layer provides 28 g/m<sup>2</sup>/year of carbon uptake, resulting in a rainforest equivalent of 0.1647.

Biomass ranges from 1 to 1200 g in dry weight. The median value is 260 g/m<sup>2</sup> for soil and 130 g/m<sup>2</sup> for epiphytes. Converted to per square kilometre, these values are 260 and 130 tons, respectively. A conversion factor of 2 was used based on the methodology by Petersson et al. (2012). Therefore, carbon content ranges from 130 to 75 t/km<sup>2</sup>. The rainforest value of 2100 t/ha corresponds to rainforest equivalents of 0.0619 and 0.0357.

In tropical rainforests, moss biomass on the forest floor varies from 10-12 g/m<sup>2</sup> in lowlands and increases with altitude. In mountainous areas, it can reach 200-500 kg/ha, and in cloud forests, it can reach up to 10,300 kg/ha. In cloud forests, mosses can retain up to 30,000 L of water per hectare when it rains, playing an important role in water management and nutrient supply (Pócs, 1982; Frahm et al., 2003). The lower biomass in lowland areas may be due to high temperatures causing respiration that cannot be balanced by photosynthesis under low light. In cloud forests, lower temperatures favour mosses and lichens (Richards, 1984; Wagner et al., 2013).

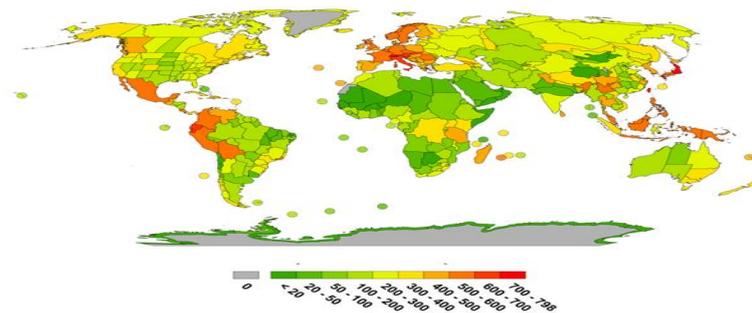
There are about 13,000 species of mosses (Goffinet et al., 2008) and 13,500 species of lichens (Hawksworth et al., 1995). The Phylum Bryophyta consists of approximately 13,000 moss species, divided into eight classes. These include pleurocarpous mosses (4–7 classes, or 42% of species) and acrocarpous mosses (23 orders), which are not linked to a single evolutionary group. Ninety percent of existing moss species belong

to two subclasses: Dicranidae (acrocarpous haplolepidae) and Bryidae (diplolepideous-alternate mosses with both acrocarpous and pleurocarpous members) (Newton et al., 2009).

Schmit and Mueller (2007) estimate that there are at least 20,000 species of lichens, which is more than the previous estimate by Hawksworth et al. (1995). Most lichen species are found in temperate zones rather than tropical regions. An exception is Ecuador, which has a very high species count. The Andes Mountain range runs through Ecuador, and despite being near the equator, the climate is relatively moderate, which may explain the high diversity.

Many known species come from the Alps, the British Isles, and the Scandinavian Peninsula, but there may still be many undescribed species in tropical countries (Feurerer and Hawksworth, 2007). In Austria, Köckinger et al. (2011) report 1138 moss species, while Switzerland has 1100 species (Hofmann and Schnyder, 2010). The Alps, covering an area of 170,000 km<sup>2</sup>, are one of the most thoroughly studied regions. Nimis et al. (2018) list 3009 species, excluding some uncertain findings. The Alps have 4450 vascular plant species, equivalent to 2200 species per 10,000 km<sup>2</sup> (Aeschimann et al., 2011).

Using the Arrhenius equation, the estimated number of rainforest species for 170,000 km<sup>2</sup> is 2977, resulting in a rainforest equivalent of 1.04. The z-value is set at 0.15, as recommended by Geffert et al. (2013). *Figure A.2.1* shows the geographic distribution of species richness.



**Figure A.2.1.** Number of moss species per 10,000 km<sup>2</sup> based on Geffert et al. (2013)

In general, species richness increases closer to the equator. This pattern is often seen across different scales, habitats, and taxa (Mittelbach et al., 2007; Cox et al., 2016). However, there are exceptions. Taxa that evolved during warmer periods have a steeper gradient, while those from colder periods show a less pronounced gradient due to their lower affinity for tropical climates (Romdal et al., 2013). They have lower temperature optima than higher-order plants (Glime, 2007). Additionally, there are fewer habitats with poikilohydric conditions in tropical climates (Mateo et al., 2016). Lichens do not show a linear gradient in species richness. Instead, they show a U-shaped pattern in the relationship between species richness and latitude (Holt et al., 2015).

Mosses and lichens have traits that suit cold climates. Many species have a net assimilation rate at 10-15°C, and even below freezing, assimilation and respiration still occur (Longton, 1992).

Due to global climate change, rising temperatures and decreased precipitation may reduce moss-covered areas and their biomass. Mosses will also spend more time in an inactive state, without absorbing carbon dioxide (Li et al., 2021).

An important ecosystem service provided by mosses and lichens is soil formation and protection against degradation. Due to human activities, their role in this service may decrease, which will negatively affect other organisms (Allen and Lendemer, 2016).

#### REFERENCES OF MOSSES, LICHENS

- [1] Aeschimann, D., Rasolofso, N., Theurillat, J.-P. (2011): Analyse de la flore des Alpes. 1: historique et biodiversité. – *Candollea* 66: 27-55.
- [2] Allen, J. L., Lendemer, J. C. (2016): Climate change impacts on endemic, high-elevation lichens in a biodiversity hotspot. – *Biodiversity and Conservation* 25: 555-568.
- [3] Blok, D., Heijmans, M. M. P. D., Schaepman-Strub, G., van Ruijven, J., Parmentier, F. J. W., Maximov, T. C., Berendse, F. (2011): The Cooling Capacity of Mosses: Controls on Water and Energy Fluxes in a Siberian Tundra Site. – *Ecosystems* 14: 1055-1065.
- [4] Cox, C. B., Ladle, R., Moore, P. D. (2016): *Biogeography: An Ecological and Evolutionary Approach*. – Wiley Blackwell, Chichester, UK.
- [5] Elbert, W., Weber, B., Büdel, B., Andreae, M. O., Pöschl, U. (2009): Microbiotic crusts on soil, rock and plants: neglected major players in the global cycles of carbon and nitrogen? – *Biogeosciences* 6: 6983-7015.
- [6] Feuerer, T., Hawksworth, D. L. (2007): Biodiversity of lichens, including a world-wide analysis of checklist data based on Takhtajan's Xoristic regions. – *Biodiversity and Conservation* 16(1): 85-98.
- [7] Frahm, J.-P., Pócs, T., Oshea, B., Koponen, T., Piippo, S., Enroth, J., Rao, P., Fang, Y.-M. (2003): *Manual of Tropical Bryology*. – *Bryophyte Diversity and Evolution* 23: 1.
- [8] Geffert, J. L., Frahm, J.-P., Barthlott, W., Mutke, J. (2013): Global moss diversity: spatial and taxonomic patterns of species richness. – *Journal of Bryology* 35(1): 1-11.
- [9] Glime, J. M. (2007): *Bryophyte ecology*. – Michigan Technological University, Botanical Society of America & International Association of Bryologists.
- [10] Goffinet, B., Buck, W. R., Shaw, A. J. (2008): Morphology, Anatomy, and Classification of the Bryophyta. – In: Goffinet, B., Shaw, A. J. (eds.) *Bryophyte Biology*. Cambridge University Press, New York.
- [11] Hawksworth, D. L., Kirk, P. M., Sutton, B. C., Pegler, D. N. (1995): *Ainsworth and Bisby's Dictionary of the Fungi*. – CAB International, Wallingford.
- [12] Hofmann, H., Schnyder, N. (2010): Switzerland: where the Mediterranean meets the Arctic. – *Field Bryology* No. 102.
- [13] Holt, E. A., Bradford, R., Garcia, I. (2015): Do lichens show latitudinal patterns of diversity? – *Fungal Ecology* 15: 63-72.
- [14] Honegger, R. (2009): Lichen-Forming Fungi and Their Photobionts. – In: Deising, H.-B. (eds.) *Plant Relationships*. Springer, Berlin, Heidelberg.
- [15] Kenrick, P., Crane, P. R. (1997): *The Origin and Early Diversification of Land Plants: A Cladistic Study*. – Smithsonian Institution Press, Washington, DC.
- [16] Köckinger, H., Schröck, C., Krisai, R., Zechmeister, H. G. (2011): Checklist of Austrian Bryophytes. – Division of Conservation Biology, Vegetation Ecology and Landscape Ecology, University of Vienna.
- [17] Li, X., Hui, R., Zhang, P., Song, N. (2021): Divergent responses of moss- and lichen-dominated biocrusts to warming and increased drought in arid desert regions. – *Agricultural and Forest Meteorology* 303: 108387.
- [18] Mateo, R. G., Broennimann, O., Normand, S., Petitpierre, B., Araújo, M. B., Svenning, J.-C., Baselga, A., Fernández-González, F., Gómez-Rubio, V., Muñoz, J., Suarez, G. M., Luoto, M., Guisan, A., Vanderpoorten, A. (2016): The mossy north: an inverse latitudinal diversity gradient in European bryophytes. – *Scientific Report* 6: 25546.
- [19] Mittelbach, G. G., Schemske, D. W., Cornell, H. V., Allen, A. P., Brown, J. M., Bush, M. B., Harrison, S. P., Hurlbert, A. H., Knowlton, N., Lessios, H. A., McCain, C. M.,

- McCune, A. R., McDade, L. A., McPeck, M. A., Near, T. J., Price, T. D., Ricklefs, R. E., Roy, K., Sax, D. F., Schluter, D., Sobel, J. M., Turelli, M. (2007): Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. – *Ecology Letters* 10: 315-331.
- [20] Newton, A. E., Wikstrom, N., Shaw, A. J. (2009): Mosses (Bryophyta). – In: Hedges, S. B., Kumar, S. (eds.) *The Timetree of Life*. Oxford University Press, Oxford.
- [21] Nimis, P. L., Hafellner, J., Roux, C., Clerc, P., Mayrhofer, H., Martellos, S., Bilovitz, P. O. (2018): The lichens of the Alps—an annotated checklist. – *MycKeys* 31: 1-634.
- [22] Petersson, H., Holm, S., Ståhl, G., Alger, D., Fridman, J., Lehtonen, A., Lundström, A., Mäkipää, R. (2012): Individual tree biomass equations or biomass expansion factors for assessment of carbon stock changes in living biomass—a comparative study. – *Forest Ecology and Management* 270: 78-84.
- [23] Pócs, T. (1982): Tropical Forest Bryophytes. – In: Smith, A. J. E. (ed.) *Bryophyte Ecology*. Chapman & Hall, London.
- [24] Richards, P. W. (1984): The Ecology of Tropical Forest Bryophytes. – In: Schuster, R. M. (ed.) *New Manual of Bryology*. Hattori Botanical Laboratory, Miyazaki.
- [25] Romdal, T. S., Araújo, M. B., Rahbek, C. (2013): Life on a tropical planet: niche conservatism and the global diversity gradient. – *Global Ecology and Biogeography* 22: 344-350.
- [26] Schmit, J. P., Mueller, G. M. (2007): An estimate of the lower limit of global fungal diversity. – *Biodiversity and Conservation* 16: 99-111.
- [27] Wagner, S., Zotz, G., Allen, N. S., Bader, M. Y. (2013): Altitudinal changes in temperature responses of net photosynthesis and dark respiration in tropical bryophytes. – *Annals of Botany* 111: 455-465.

### A.3. Animal biomass, production and biodiversity were not taken into account

Heterotrophic organisms occupy the next trophic levels, and their biomass and production depend on how much primary production they consume and transfer to the next level. Whittaker and Likens (1972) estimated consumption rates as follows: 1% for cultivated lands, 2-3% for deserts and tundras, 4-7% for forests, and 10-15% for steppes and savannas. In aquatic ecosystems, estimates are 40% in the open ocean, 40% in upwelling zones, 30% on continental shelves, and 15% in kelp forests and river mouths. On land, the average is around 7%, while in the ocean, it is 37%. These values are estimates based on limited data and species studied, and we lack complete data for all species. *Table A.3.1* shows animal biomass values for different habitat types.

In rainforests, the carbon content of plant biomass is 210 tons per hectare, while average animal biomass carbon is 9 g/m<sup>2</sup>, which equals 0.09 tons per hectare. The ratio of 0.09/210 equals 0.00043, representing the rainforest equivalent. Production is 0.064 tons of carbon per square kilometre. Using a value of 17 t/ha for rainforests, the ratio of 0.064/17 equals 0.0038, representing the rainforest equivalent. This shows that only a small amount of carbon reaches heterotrophic animals at higher trophic levels.

The increase in species richness towards the equator, known as the latitudinal gradient, has long been recognised in ecology (Pianka, 1966; Mittelbach et al., 2007). The tropics have higher species richness, particularly for vertebrates. In contrast, the Nearctic and Palearctic regions have temperate or cold climates with fewer plant species, lower biomass, and lower production. This is also reflected in animal diversity, as adapting to cold climates is stressful, and plant food sources are less diverse. *Table A.3.2* shows vertebrate species richness in different zoogeographical realms, reflecting climate conditions and vegetation diversity, which forms the basis of the food chain.

**Table A.3.1.** Average biomass and production of animals by ecosystem based on Whittaker and Likens (1972)

Ecosystem	Animal biomass t C/ha	production t C/ha
Tropical rainforest	0.09	0.064705882
Tropical seasonal forest	0.054	0.04
Temperate evergreen forest	0.045	0.024
Temperate deciduous forest	0.07	0.027142857
Boreal forest	0.022	0.014166667
Woodland and shrubland	0.022	0.01375
Savanna	0.068	0.07
Temperate grassland	0.031	0.033333333
Tundra and alpine	0.002	0.001875
Desert	0.002	0.0015
Rock, ice and sand	0.000004	4.16667*10 <sup>-6</sup>
Cultivated land	0.002	0.002857143
Swamp and marsh	0.045	0.09
Lake and stream	0.022	0.048
Total terrestrial	0.031	0.024966443
Open ocean	0.011	0.034337349
Upwelling zone	0.045	0.125
Continental shelf	0.027	0.073308271
Algal bed and reef	0.09	0.183333333
Estuaries	0.068	0.178571429
Total marine	0.0124	0.038116343

**Table A.3.2.** Species richness of terrestrial vertebrate species by biogeographic realm based on Millennium Ecosystem Assessment (2005)

Biogeographic realm	Amphibians	Birds	Mammals	Reptiles	Total
Australasian	545	1669	688	1305	4207
Antarctic	0	36	0	0	36
Afrotropical	930	2228	1161	1703	6022
Indo-Malayan	882	2000	940	1396	5218
Nearctic	298	696	481	470	1945
Neotropical	2732	3808	1282	2561	10,383
Oceanic	3	272	15	50	340
Palaearctic	395	1528	903	774	3600
Total	5785	12,237	5470	8259	31,751

Due to human activities, all three characteristics have significantly decreased. The current extinction rate is about 48 species per million per year, up to 40 times the average Cenozoic extinction rate (Barnosky et al., 2011). Most endangered species are in the tropics, following the trend of species richness (Pereira et al., 2012). While animal biomass and species richness decline, human and domestic animal biomass, as well as human-made structures, are increasing. In 2020, the mass of buildings and infrastructure was 1100 Gt, and non-degradable plastic was 8 Gt (Elhacham et al., 2020). In 1900, humans carried 13 Mt of carbon, and domestic animals accounted for 35 Mt. Over the past century, these values have increased to 55 and 120 Mt, and they continue to grow rapidly (Smil, 2011).

## REFERENCES OF ANIMAL BIOMASS, PRODUCTIVITY AND DIVERSITY

- [1] Barnosky, A. D., Matzke, N., Tomiya, S., Wogan, G. O. U., Swartz, B., Quental, T., Marshall, C., McGuire, J. L., Lindsey, E. L., Maguire, K. C., Mersey, B., Ferrer, E. A. (2011): Has the Earth's sixth mass extinction already arrived? – *Nature* 471(7336): 51-57.
- [2] Elhacham, E., Ben-Uri, L., Grozovski, J., Bar-On, Y. M., Milo, R. (2020): Global human-made mass exceeds all living biomass. – *Nature* 588: 442-444.
- [3] Millennium Ecosystem Assessment. (2005): *Ecosystems and Human Well-being: Current State and Trends: Findings of the Condition and Trends Working Group.* – Island Press, Washington.
- [4] Mittelbach, G. G., Schemske, D. W., Cornell, H. V., Allen, A. P., Brown, J. M., Bush, M. B., Harrison, S. P., Hurlbert, A. H., Knowlton, N., Lessios, H. A., McCain, C. M., McCune, A. R., McDade, L. A., McPeck, M. A., Near, T. J., Price, T. D., Ricklefs, R. E., Roy, K., Sax, D. F., Schluter, D., Sobel, J. M., Turelli, M. (2007): Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. – *Ecology Letters* 10: 315-331.
- [5] Pereira, H. M., Navarro, L. M., Martins, I. S. (2012): Global biodiversity change: the bad, the good, and the unknown. – *Annual Review of Environment and Resources* 37: 25-50.
- [6] Pianka, E. R. (1966): latitudinal gradients in species diversity: a review of concepts. – *The American Naturalist* 100(910).
- [7] Smil, V. (2011): *Harvesting the biosphere: the human impact.* – *Population and Development Review* 37(4): 613-636.
- [8] Whittaker, R. H., Likens, G. E. (1972): *Carbon in the Biota.* – In: Woodwell, G. M., Pecan, E. V. (eds.) *Carbon and the Biosphere.* Brookhaven National Laboratory, Brookhaven Symposia in Biology, 24. Upton, New York.

### A.4. We have not investigated the role of microorganisms

Ecosystem processes largely depend on microorganisms and their metabolic activities (Schlesinger, 1997; Madsen, 2011). For example, microorganisms play a key role in the global carbon cycle by decomposing organic matter and producing carbon dioxide and methane, which are returned to the atmosphere (Conrad, 1996; Singh et al., 2010; Bridgham et al., 2012). Earth is a closed system where matter cycles between the lithosphere, atmosphere, hydrosphere, and biosphere. Microbial metabolism drives these biogeochemical cycles, ensuring a continuous replenishment of carbon dioxide through processes like photosynthesis.

Microorganisms are crucial for maintaining ecosystem health, climate stability, agriculture, and human well-being. Their biomass per unit area and carbon content vary across ecosystems. He et al. (2020) provided biomass estimates using methods like phospholipid fatty acids, microscopic data, CFU, respiration data, and glucosamine and muramic acid concentrations, presented in *Tables A.4.1.* and *A.4.2.* These values are for the upper 30 cm of soil. Interestingly, the highest values are found in cold climates like taiga and tundra, rather than in tropical rainforests. This is likely due to the high organic matter content in taiga and tundra soils (He et al., 2019).

For the rainforest, the biomass values are 64.42 and 51.58 g per hectare, which equal 0.6442 and 0.5158 tons. Dividing these by the rainforest equivalent value of 210 tons gives 0.0031 and 0.0025 equivalents.

The microbial metabolic quotient is the ratio of microbial respiration to biomass, indicating the activity of microbial communities in the soil. Its average value varies by biome and changes with microbial biomass levels. Xu et al. (2017) conducted a global

synthesis of published data. In a tropical forest, the microbial metabolic quotient is 1.35 mmol C mol MBC<sup>-1</sup> h<sup>-1</sup> (Microbial Biomass Carbon). Tropical forest microbial biomass carbon is 51.58 g per m<sup>2</sup>, or 4.3 mol. 1.35 mmol equals 0.00135 mol. The ratio of 4.3 mol of biomass carbon per m<sup>2</sup> becomes 58 mol per hectare, or 696 g. Given the rainforest's biomass carbon content of 210 tons, the microbial metabolic quotient is quite low. *Table A.4.3.* shows microbial quotients across different habitat types.

Delgado-Baquerizo and Eldridge (2019) estimated global microbial diversity using 237 soil samples from all biomes and continents. They identified 25,424 OTUs using 16S rRNA analysis. The chart shows OTU distribution per biome. The highest species richness, or OTU count, is found in arid and temperate regions with lower soil carbon content and limited precipitation. Diversity negatively correlates with soil carbon content. *Figure A.4.1.* shows the geographic distribution of OTUs.

**Table A.4.1.** Fungal and bacterial biomass carbon content per unit soil per biomolecule based on He et al. (2020)

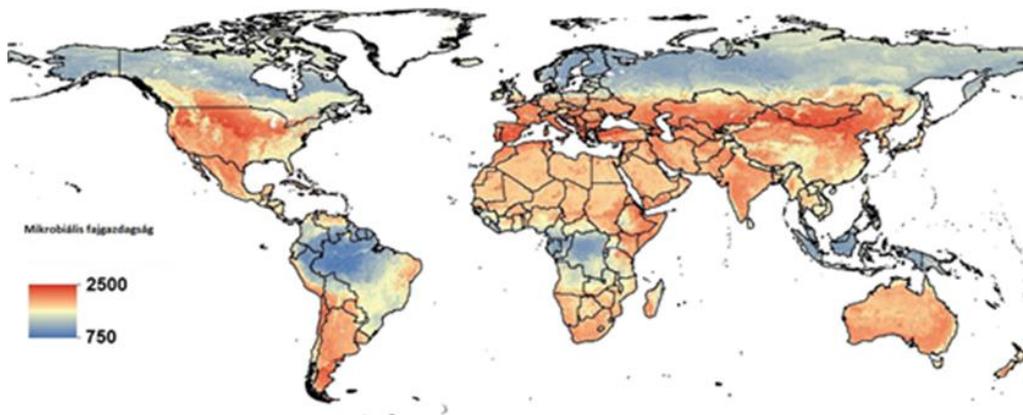
Ecosystems	Fungal biomass carbon (mg kg <sup>-1</sup> soil)	Bacterial biomass carbon (mg kg <sup>-1</sup> soil)
Unvegetated ground	192.74	24.6
Desert	16.92	6.83
Grassland	215.19	62.69
Pasture	632.15	270.65
Cropland	212.69	65.77
Shrub	218.14	45.42
Savanna	103.36	44.37
Tropical forest	451.4	209.96
Temperate forest	258.39	53.05
Boreal forest	1234.08	226.37
Tundra	3683.59	428.37
Wetlands	329.81	92.58

**Table A.4.2.** Fungal and bacterial biomass carbon per unit area based on He et al. (2020)

Ecosystem	Fungi biomass C density (g C m <sup>-2</sup> )	Bacteria biomass C density (g C m <sup>-2</sup> )
Boreal forest	304.44	58.66
Temperate forest	88.89	29.88
Tropical forest	64.42	51.58
Grassland	88.69	46.14
Shrub	48.06	17.31
Tundra	226.96	32.65
Desert	59.04	15.28
Wetlands	70.44	32.96
Croplands	67.61	30.09
Pasture	62.34	23.68
Globe	96.92	33.5

**Table A.4.3.** Microbial metabolic ratio values per biome according to Xu et al. (2017)

Ecosystems	Soil (mmol C mol MBC <sup>-1</sup> h <sup>-1</sup> )
Bare soils/desert	3.21
Boreal forest	1.28
Cropland	3.11
Grassland	1.11
Natural wetlands	3.75
Pasture	1.12
Shrubland	1.96
Temperate broadleaf	1.19
Temperate conifer	0.99
Tropical/subtropical	1.35
Global average	2.01
Area-weighted global average	1.81



**Figure A.4.1.** Geographical distribution of microbial OTUs (phylotype) according to Delgado-Baquerizo and Eldridge (2019)

## REFERENCES OF MICROORGANISMS

- [1] Bridgham, S. D., Cadillo-Quiroz, H., Keller, J. K., Zhuang, Q. (2012): Methane emissions from wetlands: biogeochemical, microbial, and modeling perspectives from local to global scales. – *Global Change Biology* 19(5): 1325-1346.
- [2] Conrad, R. (1996): Soil microorganisms as controllers of atmospheric trace gases (H<sub>2</sub>, CO, CH<sub>4</sub>, OCS, N<sub>2</sub>O, and NO). – *Microbiology Review* 60: 609.
- [3] Delgado-Baquerizo, M., Eldridge, D. J. (2019): Cross-biome drivers of soil bacterial alpha diversity on a worldwide scale. – *Ecosystems* 22: 1220-1231.
- [4] He, L., Hou, E., Veen, G. F., Farnon Ellwood, M. D., Dijkstra, P., Sui, X., Zhang, S., Wen, D., Chu, C. (2019): Soil microbial biomass increases along elevational gradients in the tropics and subtropics but not elsewhere. – *Global Ecology and Biogeography* 29(2): 1-10.
- [5] He, L., Mazza Rodrigues, J. L., Soudzilovskaia, N. A., Barceló, M., Olsson, P. A., Song, C., Tedersoo, L., Yuan, F., Yuan, F., Lipson, D. A., Xu, X. (2020): Global biogeography of fungal and bacterial biomass carbon in topsoil. – *Soil Biology and Biochemistry* 151: 108024.
- [6] Madsen, E. L. (2011): Microorganisms and their roles in fundamental biogeochemical cycles. – *Current Opinion in Biotechnology* 22: 456-464.

- [7] Schlesinger, W. H. (1997): Biogeochemistry: An Analysis of Global Change. – Academic Press, San Diego.
- [8] Singh, B. K., Bardgett, R. D., Smith, P., Reay, D. S. (2010): Microorganisms and climate change: terrestrial feedbacks and mitigation options. – Nature Reviews Microbiology 8: 779-790.
- [9] Xu, X., Schimel, J. P., Janssens, I., Song, X., Song, C., Yu, G., Sinsabaugh, R. L., Tang, D., Zhang, X., Thornton, P. (2017): Global pattern and controls of soil microbial metabolic quotient. – Ecological Monographs 87(3): 429-441.

### **A.5 We have not taken into account the seas and oceans, and to a limited extent, the mangrove habitat**

In the sea, 207,821 animal species have been described (Mugnai et al., 2021). Marine habitats are less well-known than terrestrial ones, so many new species are expected to be discovered (Danovaro et al., 2017). Most known species are found in shallow waters, and species richness decreases with depth, although the proportion of unknown species is higher in deeper waters. Toward the poles, the decline in species richness is less pronounced due to the lack of extreme temperature changes in the water. Species richness is also higher on continental shelves, near continents, where there is more nutrient availability and higher production. Habitat complexity also affects species richness, which is not necessarily limited to the tropics (Costello and Chaudhary, 2017).

Terrestrial plants are excellent climate indicators because climate regulates their annual growth, reproduction, and senescence cycles. Seasonal phases in plant activity provide opportunities to observe climate-related changes. Terrestrial plants follow a 12-month cycle, while phytoplankton have much greater temporal variability, up to 100 times higher, due to their rapid growth and consumption rates (Calbet and Landry, 2004; Behrenfeld et al., 2006). Given these scale differences, differences in periodicity are expected.

While plankton phenology is not as distinct as that of plants, seasonal changes are still observed in temperate regions. Phytoplankton blooms occur in spring due to rising temperatures and increased radiation (Cushing, 1959; Sommer et al., 1986; Smayda, 1997; Friedland et al., 2017). These blooms last for a few weeks due to limited nutrients, cell sinking, and consumption. Another peak in nutrient abundance occurs in late summer and autumn (Longhurst, 1995). These blooms are sensitive to climate change, and their timing changes similarly to those on land (Edwards and Richardson, 2004; Winder and Schindler, 2004). *Table A.5.1* shows changes in photosynthetic pigment concentration, which approximates biomass changes.

In the Atlantic Ocean, chlorophyll-a concentration is higher in the temperate zone (>40°) and the tropical region (10° S – 20° N), but lower in the subtropical region between them. Despite the lower concentration, the subtropical region contributes significantly to global oceanic primary and export production due to its vast area (Karl et al., 1996; Karl, 1999; Lin et al., 2011). Picoplankton dominates phytoplankton in terms of NPP, chlorophyll-a, and cell density, though nano- and microplankton are also present in significant amounts (Zubkov et al., 1998; Marañón et al., 2000; Fernández et al., 2003).

In the equatorial region, phytoplankton biomass and NPP are high year-round (Pérez et al., 2005a, b). Picoplankton, mainly *Prochlorococcus* and *Synechococcus*, dominates, but *Synechococcus* spp., picoeukaryotes, and nanoflagellates also become more common (Zubkov et al., 1998; Vaultot et al., 2008). Pigments from dinoflagellates and diatoms are found in higher concentrations (Gibb et al., 2000; Barlow et al., 2002).

**Table A.5.1.** *Phytoplankton biomass and production in the seas, based on data from the International Ocean-Colour Coordinating Group (www.ioccg.org). Biomass and production are estimated on the basis of carbon or pigment per unit area or volume*

Biomass	Typical oceanic range (per unit volume)	Typical oceanic range (per unit area)
Mass of carbon per unit volume and area	10-60 µg C/L	1-2 g C/m <sup>2</sup>
Mass of photosynthetic pigment per unit volume and area	0.01-2 µg Chl a/L	10-70 mg Chl a/m <sup>2</sup>
Primary production	10-100 mg C/m <sup>3</sup> /day	75-1000 mg C/m <sup>2</sup> /day

Like in the subtropical zone, picoplankton remains dominant here (Marañón et al., 2000; Pérez et al., 2005b). Near coastal upwelling areas, NPP and chlorophyll-a concentration stay high, with diatoms and dinoflagellates becoming more prevalent (Gibb et al., 2000; Barlow et al., 2002). In these areas, biomass and NPP are dominated by nano- and microplankton (Marañón et al., 2000; Tarran et al., 2006).

The polar regions are the least studied in this regard. Research is challenging due to harsh conditions, continuous ice cover, long winter darkness, rapid seasonal changes, and complex physical-biological interactions in the marine ecosystem (Lee et al., 2015). Since the 1990s, phytoplankton NPP has increased by 30% due to longer seasons and reduced ice cover (Arrigo and van Dijken, 2015). Ice and snow limit light penetration, but thinning ice allows more light to reach the ocean (Arrigo and van Dijken, 2011).

For example, the Chukchi Sea's shallow waters have become more productive due to a longer warm period, leading to summer algal blooms (Arrigo et al., 2008; Yun et al., 2019). However, seawater salinity has decreased, and the Ekman transport effect has strengthened (McPhee et al., 2009; Mauritzen, 2012). This causes the deepening of the nitrocline and chlorophyll maximum (McLaughlin and Carmack, 2010). The stronger Ekman effect and stratification reduce nutrient supply to the euphotic layer, decreasing NPP in the summer around Canada. Reduced salinity appears to negatively affect marine phytoplankton production (Yun et al., 2014).

In the Arctic region, NPP on continental shelves reaches about 225 mg C/m<sup>2</sup>/day or 27 g C/m<sup>2</sup>/year, while in the open ocean it is around 75 mg C/m<sup>2</sup>/day or 9 g C/m<sup>2</sup>/year (Subba Rao and Platt, 1984). Pabi et al. (2008) reported NPP in the Arctic from March to September as 420 ± 26 mg C/m<sup>2</sup>/day, with an annual average of 419 ± 33 Tg from 1998-2006 and a 26% variation per year.

Joo et al. (2012) identified 71 taxa in the Chukchi and Beaufort Seas, including Dinophyceae, Cryptophyceae, Bacillariophyceae, Chrysophyceae, Dictyochophyceae, Prasinophyceae, and Prymnesiophyceae. In the Bering Sea, *Fragilariopsis* was most common at the surface, while *Cryptomonas* sp. dominated at the Subsurface Chlorophyll Maximum (SCM). In the Bering Strait, *Phaeocystis* sp. was most common at both surface and SCM depth. *Chaetoceros* sp. dominated at the surface in the Chukchi Sea, while *Halosphaera* sp. was dominant at the surface in the Canadian Basin, with *Navicula* sp. dominant at the SCM depth. *Table A.5.1* summarises the phytoplankton data.

In the ocean, the highest NPP values do not necessarily occur at the equator, unlike on land. Nutrient availability limits NPP, and various factors affect nutrient concentrations. Rare events can cause sudden increases in nutrients. For example, Hamme et al. (2010) reported increased iron levels after a volcanic eruption, which boosted NPP. Light is also

crucial for photosynthesis, so its intensity, duration, and depth of penetration are important for algae. In polar regions, long winter darkness limits NPP due to a lack of light (Sigman and Hain, 2012). Some ocean areas experience upwelling, where currents bring nutrient-rich water to the surface, increasing productivity (Gómez-Letona et al., 2017). Conversely, low nutrient levels can result in low algal productivity (Sigman and Hain, 2012).

Seagrasses belong to two families, Potamogetonaceae and Hydrocharitaceae (Alismatales), which include 12 genera and about 50 species (Hemminga and Duarte, 2000). Other sources divide them into three families: Hydrocharitaceae, Cymodoceaceae, and Zosteraceae (Les et al., 1997; Ross et al., 2016). Seagrasses are found worldwide, except in polar regions (Orth et al., 2006). They regulate the physical, chemical, and biological characteristics of their surroundings and provide many ecosystem services (Constanza et al., 1997; Hemminga and Duarte, 2000; Wright and Jones, 2006).

Seagrasses are important food sources for marine turtles (*Chelonia mydas*), dugongs (*Dugong dugon*), and manatees (*Trichechus* spp.). They also provide habitat for many fish species (Beck et al., 2001). By stabilising sediments, seagrasses can store large amounts of carbon (Orth et al., 2006). They have developed unique adaptations to underwater life, including internal gas transport, epidermal chloroplasts, underwater pollination, and propagation (den Hartog, 1970; Papenbrock, 2012). Like other photosynthetic marine organisms, the abundance and distribution of seagrasses along vertical gradients largely depend on light availability.

As ocean depth increases, light decreases rapidly, challenging plant photosynthetic machinery, which very few species can handle. Underwater radiation is determined by the light attenuation coefficient ( $k$ ), which depends on factors like eutrophication, turbidity, sedimentation, latitude, and topography (Minguito-Frutos et al., 2023). This makes seagrasses highly sensitive to changes in water quality. Due to the declining water quality from human activities, fishing practices, and invasive species, seagrass meadows are continually shrinking (Orth et al., 2006).

Kelp forests grow in cold, shallow, rocky coastal waters and are primarily composed of brown algae from the Laminariales order (Dayton, 1985; Bolton, 2010). The largest species, *Macrocystis*, can reach up to 45 m and are found on the western coasts of North and South America. *Nereocystis leutkeana* dominates from central California to Alaska, *Ecklonia maxima* in South Africa, and *Alaria fistulosa* in East Asia. Globally, only 20 species form these forests, distributed across 16 mostly monotypic genera (Abbott and Hollenberg, 1976; Reed and Brzezinski, 2009; Smale, 2020).

Kelp forests are morphologically diverse, with different layers similar to terrestrial rainforests (prostrate, stipitate, and canopy forms) (Dayton, 1985; Smale and Moore, 2017). This structural diversity supports various marine mammals, fish, crustaceans, sea urchins, molluscs, and other algae, making it one of the world's most species-rich ecosystems (Mann, 1973; Port et al., 2016). Despite high productivity, individual kelp organisms have short lifespans, typically living only 2-5 years and reaching maximum length in 1-3 years (Steneck and Dethier, 1994).

In temperate zones, mangrove forests are replaced by salt marshes, which are formed by halophytic herbaceous plants in the intertidal zone. These marshes develop in areas with low wave action, allowing sediment to settle and accumulate, creating a suitable habitat for salt-tolerant plants. Near river deltas, salt marshes often gradually transition into freshwater marshes (Allen and Pye, 1992; Wang et al., 2023).

Open water areas often interrupt the vegetation, creating complex habitats shaped by tidal movements (tidal creeks) (Allen, 2000). Zonation occurs based on distance from the sea and elevation (Chapman, 1974; Moffett et al., 2010). There is an inverse relationship between a plant's competitive ability and its stress tolerance. Species with stronger competitive abilities occupy less stressful areas, while those with weaker abilities are confined to more stressful zones, such as areas with high salt concentration and oxygen depletion (Bertness, 1992; Pennings and Bertness, 2001).

In North America, *Spartina alterniflora* dominates the US coast, while *Puccinellia phryganodes* is common on the Canadian coast up to the Arctic (Poaceae). In Western Europe, *Spartina maritima* and *Spartina anglica* are most common. Other characteristic plants include *Limonium nashii*, *Salicornia* spp., *Aster tenuifolius*, *Distichlis spicata*, *Gerardia maritima*, *Spartina patens*, *Spergularia marina*, and *Ruppia maritima*. Typical large algae species are *Ascophyllum nodosum*, *Fucus vesiculosus*, *Enteromorpha*, *Ulva* spp., and *Codium fragile*. The biomass of *A. nodosum* is often greater than *S. alterniflora*. Diatoms are the most characteristic microscopic algae (Teal, 1986; Rinke et al., 2021).

These marshes provide habitat for many arthropods, birds, and mammals (Vince, 1979; Wong et al., 2011). They serve as breeding and nursery grounds for fish and play a key role in erosion control (Levin et al., 2001). Environmental pollution has caused heavy metal accumulation in sediments and plant tissues (Giblin, 1985; Tupan and Azrianingsih, 2016). Overgrazing threatens marshes by altering plant communities and animal populations (Knottnerus, 2005). Some marshes are diked off for agriculture (Reise, 2005), and salt production also poses a threat by creating ponds and removing vegetation (Laszlo, 2001). *Table A.5.2* summarises data on seagrasses, macroalgae, and salt marshes, while *Tables A.5.3* and *A.5.4* provide data on net primary productivity (NPP) in marine realms.

**Table A.5.2.** *Multicellular algae (kelp), seagrass and salt marsh biomass and production data. Production was measured as assimilated carbon. For kelp and seagrass, I found carbon stock data in the form of Mg CO<sub>2</sub> e/ha*

<b>Kelp forest</b>		
Carbon stock	120-720 g C/m <sup>2</sup>	Weston et al., 2012
Assimilated carbon	400 g/m <sup>2</sup> /	Yee, 2010
<b>Sea grass</b>		
Carbon stock	7.29 ± 1.52 Mg C/ha	Fourqurean et al., 2012
Assimilated carbon	Between -77 and + 85 Mg CO <sub>2</sub> e/ha/year	Sifleet et al., 2011
<b>Salt marsh</b>		
Biomass	5.1-18.3 Mg CO <sub>2</sub> e/ha	Sifleet et al., 2011
Assimilated carbon	0.01-62.81 Mg CO <sub>2</sub> e ha/year	Sifleet et al., 2011

By slightly modifying the concept, it can also be used to describe aquatic habitats. The net primary productivity (NPP) of phytoplankton depends on the amount of dissolved nutrients in the water. According to Taelman et al. (2014), unlike on land, the highest ocean NPP value (exergy-based potential NPP) is found in the temperate Pacific. The calculation method is detailed in their paper (Eppley-VGPM model). *Tables A.5.5* and *A.5.6* summarise the exergy-based potential NPP values and their area-weighted products.

**Table A.5.3.** NPP values for biogeographical realms. Spatial data are from Spalding et al. (2007)

Realm	Area (km <sup>2</sup> )	Mean potential NPP (MJexm <sup>-2</sup> yr <sup>-1</sup> )	NPP per realm (MJexrealm <sup>-1</sup> yr <sup>-1</sup> )
Central Indo-Pacific	1.20*10 <sup>7</sup>	10.12	1.22*10 <sup>8</sup>
Eastern Indo-Pacific	2.78*10 <sup>7</sup>	1.92	5.34*10 <sup>7</sup>
Open Ocean	1.89*10 <sup>7</sup>	1.67	3.16*10 <sup>7</sup>
Southern Ocean	2.24*10 <sup>8</sup>	3.72	8.32*10 <sup>8</sup>
Temperate Australasia	1.03*10 <sup>7</sup>	1.51	1.56*10 <sup>7</sup>
Temperate Northern Atlantic	5.65*10 <sup>6</sup>	5.48	3.10*10 <sup>7</sup>
Temperate Northern Pacific	9.85*10 <sup>6</sup>	15.96	1.57*10 <sup>8</sup>
Temperate South America	9.65*10 <sup>6</sup>	16.18	1.56*10 <sup>8</sup>
Temperate Southern Africa	6.04*10 <sup>6</sup>	10.6	6.40*10 <sup>7</sup>
Tropical Atlantic	1.99*10 <sup>6</sup>	14.48	2.88*10 <sup>7</sup>
Tropical Eastern Pacific	1.37*10 <sup>7</sup>	4.87	6.79*10 <sup>7</sup>
Western Indo-Pacific	4.19*10 <sup>6</sup>	7.04	2.95*10 <sup>7</sup>
Marine systems	1.53*10 <sup>7</sup>		

**Table A.5.4.** NPP values for biogeographic realms in descending order. Since the Temperate Northern Pacific has the highest value, this provides the basis for the calculation of the equivalent value

Realm	Area (km <sup>2</sup> )	Mean potential NPP (MJexm <sup>-2</sup> yr <sup>-1</sup> )	NPP per realm (MJexrealm <sup>-1</sup> yr <sup>-1</sup> )
Temperate Northern Pacific	1.20*10 <sup>7</sup>	10.12	1.22*10 <sup>8</sup>
Temperate Northern Atlantic	2.78*10 <sup>7</sup>	1.92	5.34*10 <sup>8</sup>
Temperate Southern Africa	1.89*10 <sup>7</sup>	1.67	3.16*10 <sup>7</sup>
Temperate South America	2.24*10 <sup>8</sup>	3.72	8.32*10 <sup>8</sup>
Arctic	1.03*10 <sup>7</sup>	1.51	1.56*10 <sup>7</sup>
Tropical Eastern Pacific	5.65*10 <sup>6</sup>	5.48	3.10*10 <sup>7</sup>
Temperate Australasia	9.85*10 <sup>6</sup>	15.96	1.57*10 <sup>8</sup>
Tropical Atlantic	9.65*10 <sup>6</sup>	16.18	1.56*10 <sup>8</sup>
Western Indo-Pacific	6.04*10 <sup>6</sup>	10.6	6.40*10 <sup>7</sup>
Open ocean	1.99*10 <sup>6</sup>	14.48	2.88*10 <sup>7</sup>
Central Indo-Pacific	1.39*10 <sup>7</sup>	4.87	6.79*10 <sup>7</sup>
Eastern Indo-Pacific	4.19*10 <sup>6</sup>	7.04	2.95*10 <sup>7</sup>
Southern Ocean	1.53*10 <sup>7</sup>	10.12	

**Table A.5.5.** The equivalents for marine biogeographic realms. Calculated according to the method already presented for terrestrial plants

Realm	Equivalence
Temperate Northern Pacific	1
Temperate Northern Atlantic	0.986402967
Temperate Southern Africa	0.894932015
Temperate South America	0.65512979

Arctic	0.625463535
Tropical Eastern Pacific	0.435105068
Temperate Australasia	0.33868974
Tropical Atlantic	0.300988875
Western Indo-Pacific	0.235475896
Open ocean	0.229913473
Central Indo-Pacific	0.118665019
Eastern Indo-Pacific	0.103213844
Southern Ocean	0.093325093

**Table A.5.6.** *The product of the equivalent and the area data.*

Realm	Equals and area multiplied
Temperate Northern Pacific	9.65*10 <sup>6</sup>
Temperate Northern Atlantic	9.72*10 <sup>6</sup>
Temperate Southern Africa	1.78*10 <sup>6</sup>
Temperate South America	3.95*10 <sup>6</sup>
Arctic	7.51*10 <sup>6</sup>
Tropical Eastern Pacific	1.82*10 <sup>6</sup>
Temperate Australasia	1.92*10 <sup>6</sup>
Tropical Atlantic	4.19*10 <sup>6</sup>
Western Indo-Pacific	3.61*10 <sup>6</sup>
Open ocean	5.14*10 <sup>7</sup>
Central Indo-Pacific	3.30*10 <sup>6</sup>
Eastern Indo-Pacific	1.96*10 <sup>6</sup>
Southern Ocean	9.63*10 <sup>5</sup>

## REFERENCES OF THE SEAS

- [1] Abbott, I. A., Hollenberg, G. J. (1976): Marine Algae of California. – Stanford University Press, Stanford, CA, USA.
- [2] Allen, J. R. L. (2000): Morphodynamics of Holocene salt marshes: a review sketch from the Atlantic and Southern North Sea coasts of Europe. – Quaternary Science Reviews 19: 1155-1231.
- [3] Allen, J. R. L., Pye, K. (1992): Saltmarshes: Morphodynamics, Conservation and Engineering Significance. – Cambridge University Press, Cambridge, UK.
- [4] Arrigo, K. R., van Dijken, G., Pabi, S. (2008): Impact of a shrinking Arctic ice cover on marine primary production. – Geophysical Research Letters 35: L19603.
- [5] Arrigo, K. R., van Dijken, G. L. (2011): Secular trends in Arctic Ocean net primary production. – Geophysical Research Letters 116(C9).
- [6] Arrigo, K. R., van Dijken, G. L. (2015): Continued increases in Arctic Ocean primary production. – Progress in Oceanography 136: 60-70.
- [7] Barlow, R. G., Aiken, J., Holligan, P. M., Cummings, D. G., Maritorena, S., Hooker, S. (2002): Phytoplankton pigment and absorption characteristics along meridional transects in the Atlantic Ocean. – Deep Sea Research Part I: Oceanographic Research Papers 49(4): 637-660.

- [8] Barlow, R. G., Aiken, J., Moore, G. F., Holligan, P. M., Lavender, S. (2004): Pigment adaptations in surface phytoplankton along the eastern boundary of the Atlantic Ocean. – *Marine Ecology Progress Series* 281: 13-26.
- [9] Beck, M. W., Heck, K. L., Able, K. W., Childers, D. L., Eggleston, D. B., Gillanders, B. M., Halpern, B., Hays, C. G., Hoshino, K., Minello, T. J., Orth, R. J., Sheridan, P. F., Weinstein, M. P. (2001): The identification, conservation, and management of estuarine and marine nurseries for fish and invertebrates: a better understanding of the habitats that serve as nurseries for marine species and the factors that create site-specific variability in nursery quality will improve conservation and management of these areas. – *BioScience* 51(8): 633-641.
- [10] Behrenfeld, M. J., O'Malley, R. T., Siegel, D. A., McCain, C. R., Sarmiento, J. L., Feldman, G. C., Milligan, A. J., Falkowski, P. G., Letelier, R. M., Boss, E. S. (2006): Climate-driven trends in contemporary ocean productivity. – *Nature* 444: 752-755.
- [11] Bertness, M. D. (1992): The ecology of a New England salt marsh. – *American Scientist* 80: 260-268.
- [12] Bolton, J. J. (2010): The biogeography of kelps (Laminariales, Phaeophyceae): a global analysis with new insights from recent advances in molecular phylogenetics. – *Helgoland Marine Research* 64: 263-279.
- [13] Calbet, A., Landry, M. (2004): Phytoplankton growth, microzooplankton grazing, and carbon cycling in marine systems. – *Limnology and Oceanography* 49(1): 51-57.
- [14] Chapman, V. J. (1974): Salt Marshes and Salt Deserts of the World. – In: Queen, W. H. (ed.) *Ecology of Halophytes*. Academic Press, New York.
- [15] Constanza, R., d'Arge, R., de Groot, R., Farber, S., Grasso, M., Hannon, B., Limburg, K., Naeem, S., O'Neill, R. V., Paruelo, J., Raskin, R. G., Sutton, P., van den Belt, M. (1997): The value of the world's ecosystem services and natural capital. – *Nature* 387: 253-260.
- [16] Costello, M. J., Chaudhary, C. (2017): Marine biodiversity, biogeography, deep-sea gradients, and conservation. – *Current Biology* 27: 511-527.
- [17] Cushing, D. H. (1959): The seasonal variation in oceanic production as a problem in population dynamics. – *Journal du Conseil/Conseil Permanent International pour l'Exploration de la Mer* 24(3): 455-464.
- [18] Danovaro, R., Corinaldesi, C., Dell'Anno, A., Snelgrove, P. V. R. (2017): The deep-sea under global change. – *Current Biology* 27: 461-465.
- [19] Dayton, P. K. (1985): Ecology of kelp communities. – *Annual Review Ecology Systems* 16: 215-245.
- [20] den Hartog, C. (1970): *The Seagrasses of the World*. – North-Holland Publishing Co., Amsterdam.
- [21] Edwards, M., Richardson, A. J. (2004): Impact of climate change on marine pelagic phenology and trophic mismatch. – *Nature* 430: 881-884.
- [22] Fernández, E., Marañón, E., Morán, X. A. G., Serret, P. (2003): Potential causes for the unequal contribution of picophytoplankton to total biomass and productivity in oligotrophic waters. – *Marine Ecology Progress Series* 254: 101-109.
- [23] Fourqurean, W. J., Duarte, M. C., Kennedy, H., Marbá, N., Holmer, M., Mateo, A. M., Apostolaki, T. E., Kendrick, A. G., Krause-Jensen, D., McGlathery, J. K., Serrano, O. (2012): Seagrass ecosystems as a globally significant carbon stock. – *Nature Geoscience* 5(7): 505-509.
- [24] Friedland, K. D., Mouw, C. B., Asch, R. G., Ferreira, A. S. A., Henson, S., Hyde, K. J. W., Morse, R. E., Thomas, A. C., Brady, D. C. (2017): Phenology and time series trends of the dominant seasonal phytoplankton bloom across global scales. – *Global Ecology and Biogeography* 27(5): 551-569.
- [25] Gibb, S. W., Barlow, R. G., Cummings, D. G., Rees, N. W., Trees, C. C., Holligan, P., Suggett, D. (2000): Surface phytoplankton pigment distributions in the Atlantic Ocean: an assessment of basin scale variability between 50° N and 50° S. – *Progress in Oceanography* 45(3-4): 339-368.

- [26] Giblin, A. E. (1985): Comparisons of the Processing of Elements by Ecosystems. II. Metals. – In: Godfrey, P. J., Benforado, J. (eds.) *Ecological Considerations in Wetland Treatment of Wastewater*. Van Nostrand Reinhold Company, New York.
- [27] Gómez-Letona, M., Ramos, A. G., Coca, J., Arístegui, J. (2017): Trends in primary production in the canary current upwelling system—a regional perspective comparing remote sensing models. – *Frontiers in Marine Science* 4: 370.
- [28] Hamme, R. C., Webley, P. W., Crawford, W. R., Whitney, F. A., DeGrandpre, M. D., Emerson, S. R., Eriksen, C. C., Giesbrecht, K. E., Gower, J. F. R., Kavanaugh, M. T., Peña, M. A., Sabine, C. L., Batten, S. D., Coogan, L. A., Grundle, D. S., Lockwood, D. (2010): Volcanic ash fuels anomalous plankton bloom in subarctic northeast Pacific. – *Geophysical Research Letters* 37(19): L19604.
- [29] Hemminga, M. A., Duarte, C. M. (2000): *Seagrass Ecology*. – Cambridge University Press, Cambridge, UK.
- [30] Herbland, A., Le Bouteiller, A., Raimbault, P. (1987): Does the nutrient enrichment of the equatorial upwelling influence the size structure of phytoplankton in the Atlantic Ocean? – *Oceanologica Acta* 6: 115-120.
- [31] International Ocean-Colour Coordinating Group. – Available at: [ioccg.org](http://ioccg.org)
- [32] Joo, H. M., Lee, S. H., Jung, S. W., Dahms, H. U., Lee, J. H. (2012): Latitudinal variation of phytoplankton communities in the western Arctic Ocean. – *Deep Sea Research Part II Topical Studies in Oceanography* 81-84: 3-17.
- [33] Karl, D. M. (1999): A sea of change: biogeochemical variability in the North Pacific Subtropical Gyre. – *Ecosystems* 2: 181-214.
- [34] Karl, D. M., Christian, J. R., Dore, S. E., Hebel, D. V., Letelier, R. M., Tupas, L. M., Winn, C. D. (1996): Seasonal and interannual variability in primary production and particle flux at Station ALOHA. – *Deep Sea Research Part II: Topical Studies in Oceanography* 43(2-3): 539-568.
- [35] Knottnerus, O. S. (2005): History of human settlement, cultural change and interference with the marine environment. – *Helgoland Marine Research* 59: 2-8.
- [36] Laszlo, P. (2001): *Salt: Grain of Life*. – Columbia University Press, New York.
- [37] Lee, Y. J., Matrai, P. A., Friedrichs, M. A., Saba, V. S., Antoine, D., Ardyna, M., Asanuma, I., Babin, M., Bélanger, S., Benoît-Gagné, M., Devred, E., Fernández-Méndez, M., Gentili, B., Hirawake, T., Kang, S. H., Kameda, T., Katlein, C., Lee, S. H., Lee, Z., Mélin, F., Scardi, M., Smyth, T. J., Tang, S., Turpie, K. R., Waters, K. J., Westberry, T. K. (2015): An assessment of phytoplankton primary productivity in the Arctic Ocean from satellite ocean color/in situ chlorophyll-a based models. – *Journal of Geophysical Research: Oceans* 120: 6508-6541.
- [38] Les, D. H., Cleland, M. A., Waycott, M. (1997): Phylogenetic studies in the Alismatidae, II: Evolution of the marine angiosperms (seagrasses) and hydrophily. – *Systematic Botany* 22: 443-463.
- [39] Levin, L. A., Boesch, D. F., Covich, A., Dahm, C., Erséus, C., Ewel, K. C., Kneib, R. T., Moldenke, A., Palmer, M. A., Snelgrove, P., Strayer, D., Weslawski, J. M. (2001): The function of marine critical transition zones and the importance of sediment biodiversity. – *Ecosystems* 4: 430-451.
- [40] Lin, P., Liu, H., Yu, Y., Zhang, X. (2011): Response of sea surface temperature to chlorophyll-a concentration in the Tropical Pacific: annual mean, seasonal cycle, and interannual variability. – *Advances in Atmospheric Sciences* 28(3): 492-510.
- [41] Longhurst, A. (1995): Seasonal cycles of pelagic production and consumption. – *Progress in Oceanography* 36(2): 77-167.
- [42] Mann, K. H. (1973): Seaweeds: their productivity and strategy for growth. – *Science* 182: 975-981.
- [43] Marañón, E., Holligan, P. M., Varela, M., Mouriño, B., Bale, A. J. (2000): Basinscale variability of phytoplankton biomass, production and growth in the Atlantic Ocean. – *Deep Sea Research Part I: Oceanographic Research Papers* 47(5): 825-857.

- [44] Marañón, E., Holligan, P. M., Barciela, R., Gonzalez, N., Mourino, B., Pazo, M. J., Varela, M. (2001): Patterns of phytoplankton size-structure and productivity in contrasting open ocean environments. – *Marine Ecology Progress Series* 216: 43-56.
- [45] Masotti, I., Moulin, C., Alvain, S., Bopp, L., Tagliabue, A., Antoine, D. (2011): Large-scale shifts in phytoplankton groups in the Equatorial Pacific during ENSO cycles. – *Biogeosciences* 8: 539-550.
- [46] Mauritzen, C. (2012): Oceanography arctic freshwater. – *Nature Geoscience* 5: 162-164.
- [47] McLaughlin, F. A., Carmack, E. C. (2010): Deepening of the nutricline and chlorophyll maximum in the Canada Basin interior. – *Geophysical Research Letters* 37: L24602.
- [48] McPhee, M. G., Proshutinsky, A., Morison, J. H., Steele, M., Alkire, M. B. (2009): Rapid change in freshwater content of the Arctic Ocean. – *Geophysical Research Letters* 36: L10602.
- [49] Minguito-Frutos, M., Boada, J., Pagès, J., Marco-Méndez, C., Arthur, R., Adams, M., Alcoverro, T. (2023): Species-specific acclimatization capacity of key traits explains global vertical distribution of seagrass species. – *Global Ecology and Biogeography* 32(6): 976-986.
- [50] Moffett, K. B., Robinson, D. A., Gorelick, S. M. (2010): Relationship of salt marsh vegetation zonation to spatial patterns in soil moisture, salinity, and topography. – *Ecosystems* 13: 1287-1302.
- [51] Mugnai, F., Megléc, E., Costantini, F., Abbiati, M., Bavestrello, G., Bertasi, F., Bo, M., Capa, M., Chenuil, A., Colangelo, M. A., De Clerck, O., Gutiérrez, J. M., Lattanzi, L., Leduc, M., Marin, D., Matterson, K. O., Mikac, B., Plaisance, L., Ponti, M., Riesgo, A., Rossi, V., Turicchia, E., Waeschenbach, A., Wangenstein, O. (2021): Are well-studied marine biodiversity hotspots still blackspots for animal barcoding? – *Global Ecology and Conservation* 32: e01909.
- [52] Orth, R. J., Carruthers, T. J. B., Dennison, W. C., Duarte, C. M., Fourqurean, J. W., Heck, K. L., Hughes, A. R., Kendrick, G. A., Kenworthy, W. J., Olyarnik, S., Short, F. T., Waycott, M., Williams, S. L. (2006): A global crisis for seagrass ecosystems. – *BioScience* 56(12): 987-996.
- [53] Pabi, S., van Dijken, G. L., Arrigo, K. R. (2008): Primary production in the Arctic Ocean, 1998–2006. – *Journal of Geophysical Research* 113: C8.
- [54] Papenbrock, J. (2012): Highlights in seagrasses' phylogeny, physiology, and metabolism: what makes them special? – *ISRN Botany* 7: 103892.
- [55] Pennings, S. C., Bertness, M. D. (2001): Salt Marsh Communities. – In: Hay, M. E. (ed.) *Marine Community Ecology*. Sinauer Associates, Sunderland.
- [56] Pérez, V., Fernández, E., Marañón, E., Serret, P., García-Soto, C. (2005a): Seasonal and interannual variability of chlorophyll a and primary production in the Equatorial Atlantic: in situ and remote sensing observations. – *Journal of Plankton Research* 27(2): 189-197.
- [57] Pérez, V., Fernández, E., Marañón, E., Serret, P., Varela, R., Bode, A., Varela, M., Varela, M. M., Morán, X. A. G., Woodward, E. M. S., Kitidis, V., García-Soto, C. (2005b): Latitudinal distribution of microbial plankton abundance, production, and respiration in the Equatorial Atlantic in autumn 2000. – *Deep Sea Research Part I: Oceanographic Research Papers* 52(5): 861-880.
- [58] Port, J. A., O'Donnell, J. L., Romero-Maraccini, O. C., Leary, P. R., Litvin, S. Y., Nickols, K. J., Yamahara, K. M., Kelly, R. P. (2016): Assessing vertebrate biodiversity in a kelp forest ecosystem using environmental DNA. – *Molecular Ecology* 25: 527-541.
- [59] Rabe, B., Karcher, M., Schauer, U., Toole, J. M., Krishfield, R. A., Pisarev, S., Kauker, F., Gerdes, R., Kikuchi, T. (2011): An assessment of Arctic Ocean freshwater content changes from the 1990s to the 2006–2008 period. – *Deep Sea Research Part I: Oceanographic Research Papers* 58(2): 173-185.
- [60] Reed, D. C., Brzezinski, M. A. (2009): Kelp Forests. – In: Laffoley, D., Grimsditch, G. (eds.) *The Management of Natural Coastal Carbon Sinks*. International Union for Conservation of Nature.

- [61] Reise, K. (2005): Coast of change: habitat loss and transformations in the Wadden Sea. – *Helgoland Marine Research* 59: 9-12.
- [62] Rinke, M., Maraun, M., Scheu, S. (2021): Spatial and temporal variations in salt marsh microorganisms of the Wadden Sea. – *Ecology and Evolution* 12(3): e8767.
- [63] Ross, T. G., Barrett, C. F., Gomez, M. S., Lam, V. K. Y., Henriquez, C. L., Les, D. H., Davis, J. I., Cuenca, A., Petersen, G., Seberg, O., Thadeo, M., Givnish, T. J., Conran, J., Stevenson, D. W., Graham, S. W. (2016): Plastid phylogenomics and molecular evolution of Alismatales. – *Cladistics* 32: 160-178.
- [64] Sifleet, S., Pendleton, L., Murray, C. B. (2011): State of the science on coastal blue carbon: a summary for policy makers. – Nicholas Institute for Environmental Policy Solution, Duke University.
- [65] Sigman, D. M., Hain, M. P. (2012): The biological productivity of the ocean. – *Nature Education Knowledge* 3(6): 1-16.
- [66] Smale, D. A. (2020): Impacts of ocean warming on kelp forest ecosystems. – *New Phytologist* 225: 1447-1454.
- [67] Smale, D. A., Moore, P. J. (2017): Variability in kelp forest structure along a latitudinal gradient in ocean temperature. – *Journal of Experimental Marine Biology and Ecology* 486: 255-264.
- [68] Smayda, T. J. (1997): What is a bloom? A commentary. – *Limnology and Oceanography* 42(5): 1132-1136.
- [69] Sommer, U., Gliwicz, Z. M., Lampert, W., Duncan, A. (1986): The PEG-model of seasonal succession of planktonic events in fresh waters. – *Archiv für Hydrobiologie* 106(4): 433-471.
- [70] Spalding, M. D., Fox, H. E., Allen, G. R., Davidson, N., Ferdaña, Z. A., Finlayson, M., Halpern, B. S., Jorge, M. A., Lombana, A., Lourie, S. A., Martin, K. D., McManus, E., Molnar, J., Recchia, C. A., Robertson, J. (2007): Marine ecoregions of the world: a bioregionalization of coastal and shelf areas. – *BioScience* 57(7): 573-583.
- [71] Steneck, R., Dethier, M. N. (1994): A functional group approach to the structure of algal-dominated communities. – *Oikos* 69: 476-498.
- [72] Subba Rao, D. V., Platt, T. (1984): Primary production of Arctic waters. – *Polar Biology* 3: 191-201.
- [73] Taelman, S. E., De Meester, S., Schaubroeck, T., Sakshaug, E., Alvarenga, R. A. F., Dewulf, J. (2014): Accounting for the occupation of the marine environment as a natural resource in life cycle assessment: an exergy based approach. – *Resources, Conservation and Recycling* 91: 1-10.
- [74] Tarran, G., Zubkov, M., Fuchs, B., Heywood, J. (2006): Latitudinal changes in the standing stocks of nano- and picoplankton in the Atlantic Ocean. – *Deep Sea Research Part II: Topical Studies in Oceanography* 53(14-16): 1516-1529.
- [75] Teal, J. M. (1986): *The Ecology of Regularly Flooded Salt Marshes of New England: A Community Profile*. – Woods Hole Oceanographic Institution, MA.
- [76] Tupan, C. I., Azrianingsih, R. (2016): Accumulation and deposition of lead heavy metal in the tissues of roots, rhizomes and leaves of seagrass *Thalassia hemprichii* (Monocotyledoneae, Hydrocharitaceae). – *Napoca* 9(3): 580-589.
- [77] Vaulot, D., Eikrem, W., Viprey, M., Moreau, H. (2008): The diversity of small eukaryotic phytoplankton ( $\leq 3 \mu\text{m}$ ) in marine ecosystems. – *FEMS Microbiology Reviews* 32: 795-820.
- [78] Vince, S. W. (1979): Response of herbivores to salt marsh fertilization. – PhD thesis, Boston University, Boston, Mass.
- [79] Wang, X., Xin, P., Zhou, Z., Zhang, F. (2023): A systematic review of morphological models of salt marshes. – *Water Science and Engineering* 16(4): 313-323.
- [80] Weston, K. A., Gregg, R., Morecroft, M. (2012): Carbon storage by habitat: review of the evidence of the impacts of management decisions and condition of carbon stores and sources. – Natural England Research Reports, Number NERR043.

- [81] Winder, M., Schindler, D. E. (2004): Climatic effects on the phenology of lake processes. – *Global Change Biology* 10(11): 1844-1856.
- [82] Wong, M. C., Peterson, C. H., Piehler, M. F. (2011): Evaluating estuarine habitats using secondary production as a proxy for food web support. – *Marine Ecology Progress Series* 440: 11-25.
- [83] Wright, J. P., Jones, C. G. (2006): The concept of organisms as ecosystem engineers ten years on: progress, limitations, and challenges. – *BioScience* 56: 203-209.
- [84] Yee, M. S. (2010): REDD and BLUE Carbon: Carbon Payments for Mangrove Conservation. – Capstone Advisory Committee Final Capstone Project Signature Form.
- [85] Yun, M. S., Whitledge, T. E., Kong, M., Lee, S. H. (2014): Low primary production in the Chukchi Sea shelf 2009. – *Continental Shelf Research* 76: 1-11.
- [86] Yun, M. S., Joo, H. M., Kang, J. J., Park, J. W., Lee, J. H., Kang, S. H., Sun, J., Lee, S. H. (2019): potential implications of changing photosynthetic end-products of phytoplankton caused by sea ice conditions in the northern Chukchi Sea. – *Frontiers in Microbiology* 10: 2274.
- [87] Zubkov, M. V., Sleight, M. A., Tarran, G. A., Burkill, P. H., Leakey, R. J. G. (1998): Picoplanktonic community structure on an Atlantic transect from 50° N to 50° S. – *Deep Sea Research Part I: Oceanographic Research Papers* 45(8): 1339-1355.
- [88] Zubkov, M. V., Sleight, M. A., Burkill, P. H., Leakey, R. J. G. (2000): Picoplankton community structure on the Atlantic Meridional Transect: a comparison between seasons. – *Progress in Oceanography* 45(3-4): 369-386.