INFLUENCE OF ARBUSCULAR MYCORRHIZAL FUNGI AND PLANT GROWTH PROMOTING YEAST ON GROWTH AND YIELD OF CROPS FOR SUSTAINABLE AGRICULTURE

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Abstract. This review focused on useful effects of arbuscular mycorrhizal fungi and plant growthpromoting yeast on agricultural plants as possible biological agents in increasing nutrient absorption, crop yield and tolerance to biotic and abiotic stress conditions. Chemical pesticides and fertilizers are becoming prevalent in causing environmental harm. To solve this problem, arbuscular mycorrhizal fungus and plant growth-promoting yeast are utilized. According to recent research, arbuscular mycorrhizal fungi and plant growth promoting yeast are more beneficial for improving soil health, nutrient absorption, nutrient mobilization, bioremediation, pest control, and disease management in an ecofriendly, efficient and cost effective manner than the inorganic chemicals. Arbuscular mycorrhizal fungi and plant growth promoting yeast both have distinct roles in plant growth and combining their inoculations can produce the best results. Utilizing plant growth-promoting yeast and arbuscular mycorrhizal fungi inoculation will decrease the need for chemical fertilizers and pesticides, which will result in attaining sustainable agriculture.

Keywords: biological agent, uptake of nutrients, reduce the excess use of chemical fertilizers, yield enhancement, acts as biofertlizer

Abbreviations. AMF - Arbuscular Mycorrhizal Fungi; PGPY - Plant Growth Promoting Yeast

Introduction

Continuous and excessive use of chemical fertilizers is harmful to the soil, the environment, and to human health and also spoil soil fertility (Naik et al., 2019). The new way of crop production causes the demand in organic fertilizers as the farmers are aiming for organic agriculture. AM fungi is the soil borne symbiotic fungi which is helpful for plants by symbiotic relations, it makes symbiosis with 80-90% of plant species and 90% of crop plants (Smith and Read, 2010) like cereals, pulses, vegetables, and horticultural plants (Kivlin et al., 2011). AM fungi increases the supply of water and nutrients such as P and N to the crop plants using extraradicle, intraradicle hyphae and arbuscules (Parniske, 2008) it can confer resistance to biotic and abiotic stress (Sun

et al., 2018). AM fungi can be used in several crops to enhance nutrient intake, improve physiological characteristics and improve plant growth and yield. In early days yeast is only used for preparing bread, wine and beer. But role of yeast has expanded from food industry to various industries especially agriculture (Barnett, 2011). Yeast involved in several processes like breaking down of complex molecules, nutrient recycling, biodegradation of hydrocarbons. The yeast which are used in the field of agriculture which promotes the plant growth is called plant growth promoting yeast (PGPY) PGPY regulates the synthesis of phytohormones, boosting the availability of nutrients, strengthening resistance to biotic and abiotic stress, suppressing phytopathogens etc. (Nimsi et al., 2023). This review aims to give insight into combined inoculation of AM fungi and plant growth promoting yeast to attain maximum benefit for sustainable agriculture.

Arbuscular mycorrhizal fungi

The term mycorrhiza is derived from the Greek word "mukés", meaning fungus, and "rhiza," meaning roots, Mycorrhizal fungi grow in the roots of plants and form a symbiotic relationship known as "mycorrhiza". They come together to form a network of tiny filaments that attach themselves to plant roots in order to extract water and nutrients from the soil. Arbuscular Mycorrhizal Fungi (AMF) is one of the best options for sustainable agriculture and food security. AMF symbiosis is said to be a very old symbiosis. According to the archeological records, it is originated millions of years ago (Humphreys et al., 2010). AM Fungi are widely found in terrestrial ecosystems (Verbruggen et al., 2012). Around 80 to 90 % of vascular plant species and 90% of crops form symbiotic association with AM fungi (Smith and Read, 2010). Some families like Amaranthaceae, Brassicaceae, Cruciferae, Chenopodiaceae, Caryophyllaceae, Juncaceae, Cyperaceae, and Polygonaceae do not form association with AM fungi due to the deterrent and inhibitory compounds produced by the host crops (Brundrett, 2009). Most of the AM fungal species belonging to the phylum Mucoromycota and sub phylum Glomeromycota (Spatafora et al., 2016). Glomerales, Archaeosperales, Paraglomerales and Diversisporales are four orders of AM fungi. 11 families, 25 genera and nearly 250 species are belonging to AMF (Schüßler et al., 2001; Spatafora et al., 2016). Although many AMF species have been studied globally, the following species are most frequently used as models viz., Rhizophagus irregularis, Gigaspora rosea, Gigaspora margarita, Gigaspora gigantea, and Funneliformis mosseae (Schüssler and Walker, 2010). AMF is an obligate symbiont and to complete their lifecycle they need the host plant. They increase crop yield by increasing uptake of nutrients such as nitrogen phosphorus and potassium (Anderson et al., 2018). Role of AMF is not only for promoting the plant growth and protect the plants from fungal infections (Smith et al., 2010; Jung et al., 2012). Through extra and intraradical hyphae fungi can supply the nutrients like Phosphate and Nitrogen. Nutrients are absorbed by mycelium osmotrophically and penetrate more area when compared to non-mycorrhizal roots (Duponnois et al., 2011). AMF has been reported to enhance soil properties like soil aggregation, available nutrients in soil, water retention, microbial activity, N cycle, C cycle, and P cycle, and soil acidity correction, in addition to its effect on growth and promotion (Jamiołkowska et al., 2018; Parihar et al., 2020).

Plant growth promoting yeast

Yeast is unicellular microorganisms that are inhabitants of soil and plants but exists in a lower number compared to bacteria and filamentous fungi (Yurkov et al., 2018). Yeasts are under the kingdom fungi, phylum Ascomycota and Basidiomycota. Yeast is a heterotrophic microorganism, which need carbon and nitrogen for their energy. Utilize carbohydrates to produce CO₂ and H₂O in the presence of oxygen. In absence of oxygen, they convert the carbohydrates to alcohol by the process called fermentation. Their shape and size varies depending on the species, some of the yeast species are round, ellipsoidal, oval, or cylindrical in shape. The size of the cells ranging from 3-10 µm. Adhesion and biofilm formation improves their ability to survive in competitive environment. Sexual and asexual reproduction is the two modes of reproduction. Budding is the first type of asexual reproduction; it needs suitable environmental conditions. which include temperature and nutrient availability. Candida, Saccharomyces, Pichia and Rhodotorula are the microorganisms that can reproduce through budding. Schizosaccharomyces are the microorganisms reproduce through fission. In extreme conditions like deficiency of nutrients, Yeast can produce spores. The shape and size of the spore is based on the yeast species (Kowalska et al., 2022). Yeast's role in agricultural ecosystems is not yet completely analysed (Fu et al., 2016) and its growing area needs to be tapped (Jeyashri et al., 2019). Application of yeast in agricultural and horticultural crops elaborated in Table 1.

Yeast	Usage in Agriculture	References	
Candida tropicales	Used as biofertilizer	Mekki and Ahmed (2005)	
Candida oleophila	Control grey mould in kivi fruit	Sui et al. (2020)	
Candida famata	Acts as a parasite forcontrol of Colletotrichum gloeosporioides by releasing enzymes	Magallon-Andalon et al. (2012	
Candida tropicalis HY	Acts as a plant growth promoter by ACC deaminase activity, IAA production and solubilization of P	Amprayn et al. (2012)	
Candida tropicalis	Prevent Banana from Antracnose	Zhimo et al. (2016)	
R. mucilaginosa	Degradation of insecticides like acetamiprid (AAP) and thiacloprid (THI)	Dai et al. (2010)	
Pichiaanomala	Prevent fruit rot in guava	Mohamed et al. (2009)	
Rhodotorula graminis (WP1) andRhodotorula mucilaginosa (PTD 2 and PTD3)	It acts as a Plant growth regulator by the production of Indole-3-acetic acid	Xin et al. (2009)	
Trichosporo nasahii	Reduce antracnose infestation in papaya	Hassan et al. (2021)	
Kluyveromyces walti, Pachytrichospora transvaalensis and S. cataegensis	It acts as a Biofertilizer	Agamy et al. (2013)	
P. kudriavzevii	Helpful for degrading atrazine	Abigail et al. (2013)	
Lipomyces starkeyi	Biodegradation of triazine herbicides	Nishimura et al. (2002)	
Pichia membranaefaciens	It acts as a Biocontrol agent and Inhibit growth of Colletotrichum gloeosporioides	Zhou et al. (2016)	

Table 1. Application of yeast in sustainable agriculture

AM fungi and PGPY as potential biological agents

AM fungi are said to be the natural plant growth enhancers and potential biological agents (Begum et al., 2019) and establish reciprocal symbionts with the roots of crop plants (Bennett and Groten, 2022) AMF inoculation provides more water to the crops, enhances moisture uptake and improves accumulation of dry (Chandrasekaran et al., 2019). AM fungi offered various mechanisms to the host plants to get rid of various abiotic stresses (Yadav et al., 2023). By serving an excellent bioagent AM fungi combat abiotic stress combined together with reduced inorganic fertilizers (Wang et al., 2023). In maize AM fungi resulted in higher leaf gas exchange and photosynthate under heat stress compared to uninoculated control (Mathur et al., 2018). Glomalin a super glue produced by AM fungi helps in sequestration of more carbon from the soil, aggregates more soil particles (Gomathy et al., 2018) and safeguard the soil from dessication through enhancing the water holding capacity of soil (Syamsiyah et al., 2018). Food crops can be cultivated even in the heavy metal contaminated soil due to the inoculation of AM fungi as it accumulates the heavy metals through various mechanisms and keep crops free of metal accumulation (Gomathy et al., 2022). Exploiting AM fungi as potential biological agent for plant growth in several ecosystems pave the way for organic farming lead to sustainable agriculture. AM fungi colonized plants exhibited higher antioxidant enzyme activity (Gomathy et al., 2021), increased level of secondary metabolites (Castellanos-Morales et al., 2010) profuse production of root exudates (Gomathy et al., 2018), more quantity of carotenoids and volatile compounds (Bona et al., 2017), enhanced accumulation of anthocyanins, chlorophyll (Balsam et al., 2011) and increased level of sugars, organic acids, vitamin C, flavonoids, and minerals (Zeng et al., 2014). AM fungi inoculated cucumber plants showed resistant to soil borne pathogens alleviate salt stress and overcome chill temperature (Aljawasim et al., 2020).

Plant growth promoting yeast can be act as a potential biological agents in different sectors of sustainable farming (Mukherjee et al., 2020) The potential use of yeast starts from baking, fermentation etc. and its vital role in agriculture (Barnett and Barnett, 2011) is gaining momentum nowadays and the interest of yeast in agriculture is increasing spontaneously. PGPY is widely used as biocontrol agents towards control of many diseases of crop plants (Pimenta et al., 2010). They are very well known to have the ability to solubilize phosphorus (Amprayn et al., 2012), offered resistance to stress (El-Zohri et al., 2017) and promote the growth and yield of plant as a potential biological agent (Hesham and Mohamed, 2011). Yeasts produces copious amount of plant growth promoting enzymes, IAA, GA and cell wall degrading enzymes that retard the growth of plant pathogens (Fu et al., 2016). Yeast has also been employed for the degradation of pesticides from the different sources (Han et al., 2019) reported that Clavispora lusitaniae showed pendimethalin degradation in CMB liquid culture (74% of 200 mg L^{-1}). Even the renowned baking yeast Saccharomyces cerevisiae has been employed for plant growth promotion and biocontrol agent in adverse conditions (Amprayn et al., 2012; Ibrahim and El-Fikil, 2019; Ferraz et al., 2019) reported that cacao witches broom disease was efficiently controlled by yeast.

Williopsis saturninus is endophytic yeast obtained from the maize roots and proven to produce indole-3-acetic acid (IAA) and indole-3- pyruvic acid (IPYA) (Nassar et al., 2005). Cytokinins are vital hormones that promotes plant growth and one among them is zeatin, reported to be synthesized from yeast *Sporobolomyces roseus* and *Aureobasidium pullulans* isolated from the rhizosphere soil of plants (Streletskii et al., 2019; Hesham et al., 2017, 2018) reported that *R. mucilaginosa, C. albidus*, *P. membranifaciens, H. uvarum* and *C. alifornica* had the ability to kill pathogens and the *Rhodotorula* species such as *R. lactose, R. nymphaeae, R. graminis, R. slooffiae* degrade the petroleum compounds from the soil through direct and indirect mechanisms. Even the crop residues can easily be degraded to enrich the soil *via* cellulose degrading enzymes of *S. cerevisiae* (Bae et al., 2015). Different genus and species of yeast can be explored further for various beneficial activities to serve as potential biological agents.

Influence of AM fungi and PGPY in nutrient uptake

AM Fungi colonization promotes nutrient uptake in plants. It is proved that AMF can improve the amount of macro and micronutrients significantly, which leads to increase in biomass production (Chen et al., 2017; Mitra et al., 2020). AM fungi can maintain approximately 90% of the plant's phosphorus, 60% of nitrogen and 20% of carbon (Smith et al., 2008). The level and the quantity of nitrogen, phosphorus, calcium and other nutrients like sulphur, potassium, iron, copper, manganese and zinc in plants can be increased by using AM fungi (Cao et al., 2015; Mustafa et al., 2016; Mei et al., 2019; Khan et al., 2021). The ability of *Rhizobium* to fix Nitrogen may be enhanced if the host plant is also in symbiosis with AM fungi. In this case, Rhizobium and AM fungi synergistically work together to increase the rate of colonization, inorganic nutrient uptake and plant development (Harrison, 1999). Legume crops are preferred to make symbiosis with AM Fungi as these symbionts are more helpful to supply P. This can be very helpful to the legume plants under the nutrient deficit conditions. Each and every AM fungal species are unique in formation of nodules that ultimately helps and improves Nitrogen fixation (Ibijbijen et al., 1996; Gomathy et al., 2021). AMF ensures the plant to get essential nutrients, provide mineral nutrients like nitrogen, phosphorus, potassium, calcium, zinc and sulphur. AMF give nutritional support to the plants even in unfavourable conditions. AMF produce arbuscules which is helpful in exchange of inorganic minerals, carbon and phosporus (Li et al., 2016; Prasad et al., 2017). Absorption of minerals and water from the faraway places are possible for AM fungi through the hyphae that are very thin when compared to root hair. By using the fungal hyphae they will penetrate even very small tiny pores of the soil (Allen et al., 2011). Micronutrients like zinc, copper, iron and manganese can also be absorbed by the AMF under nutrient deficient areas (Canton et al., 2016; Liu et al., 2018). By using the fungal hyphae AMF can expand the fungal network to the nutrient deficient areas, which helps the plant to penetrate larger volume of soil.

Plant growth can also be increased by inoculating the crops with yeast that solubilize the nutrients, produce different phytohormones, enzymes that can be helpful for plant growth (Amprayn et al., 2012; Silambarasan et al., 2019; Kumla et al., 2020). Phosphorus is a mineral that is tightly bound with other chemical cations and the availability of P is very less. The application of gibbrellic acid with yeast will improve the nutrient uptake of crop plants (Youssef et al., 2022). *Lachancea thermotolerans* is the yeast species which has more P solubilizing capacity, *Saccharomyces spp.* and *Hanseniaspora uvarum* strains contain more phosphate solubilizing capacity (Fernandez et al., 2020). Yeast can provide nutrients and helpful for nitrogen fixation, phosphorus and potassium solubilization and release trace elements like iron and zinc. Yeast produces phytohormones like auxins, cytokinins and gibbrelins which can promote the growth of plants (Hernández et al., 2021). *Candida tropicalis* is the yeast which is isolated from soil which fixes nitrogen and also solubilizes phosphorus (Mukherjee et al., 2015). *Torulaspora globosais* can solubilize K from the ultramafic alkaline rock (Rosa et al., 2012).

AM Fungi and PGPY in biotic and abiotic stress management

Biotic stress

Plant pathogen

AM fungi improve the plant defence mechanism and provide more ability to get resistant against pests and pathogens (Begum et al., 2019). Plant pathogens are more prevalent in the root area of the crop plants which can trigger the plants against the pathogens by releasing some phenolic compounds which can reduce the pathogen growth. AM fungi confer the plants with more resistance that develop physical barrier and reduce the spread of pathogens (Nanjundappa et al., 2019; George et al., 2023). Plants obtain their nutrients mostly via the joint activity of their roots and the microorganisms that are associated with them. These microbes are important for both suppressing soil pathogens, mobilizing and absorbing nutrients (Ismail et al., 2013). *Rhizophagus irregularisis* reduces the disease in tomato caused by *Candidatus Liberibacter solanacearum*. AM fungi not only reduce the infestation caused by fungi but also protect the plants from nematode infestation caused by *Meloidogyne incognita* in coffee. *Fusarium oxygensporum* f. sp. *Lycopersici* is the soil borne fungus which causes wilt disease in tomato which can reduces the yield (Akköprü and Demir, 2005; FAOSTAT, 2019).

Yeast has an ability to control the pathogenic microorganisms and increases the tolerance of the plant against pathogens (Kamel et al., 2016; Ibrahim and El-Fikil, 2019). Yeast has various antagonistic properties, that is helpful to know that their mode of action against the pathogen and increase their viability (Parafati et al., 2016). Usually, yeast has the ability to grow faster and form biofilms on the outer surface of the spoiled fruit. Due to formation of biofilms competition for nutrients occur between the pathogen and yeast. Yeast uses all the nutrients due to deficiency of nutrients the pathogenic spore germination is reduced. Aureobasidium pullulans increases the competition of nutrients to the pathogen Penicillium expansum (Bencheqroun et al., 2007), Rhodotorula glutinis also control the plant pathogen Penicillium expansum by developing siderospores (Calvente et al., 1999). Yeast not only prevent plants from pathogen attack by creating a competition for space and nutrients but also by producing volatile organic compounds (VOCs) that has parasitic effect on plant pathogens. The parasitic effect of yeast produced VOCs on plant pathogens is a multifaceted mechanism involving direct inhibition, priming of plant defenses, and disruption of pathogen behavior. Yeast produces volatile compounds during their primary and secondary metabolism (Korpi et al., 2009). The mixtures of volatiles called volatilome. VOC include alcohols, aldehydes, cyclohexanes, benzene derivatives, heterocyclic compounds, hydrocarbons, ketones, phenols, thioalcohols and thioesters. Physical contact is not needed between pathogen and VOC (Huang et al., 2012; Parafati et al., 2017). Genus Hanseniaspora prevents the plants from Botrytis cinerea by releasing VOCs (Ruiz-Moyano et al., 2020). Volatile organic compounds and glycoproteins produced by yeast disrupt the cell wall of pathogens and acts as a biocontrol agent (Freimoser et al., 2019). Rhizoctonia solani is a soil borne fungal pathogen which can

cause severe problem in root structure it can be prevented by yeast *Rhodotorula glutinis, Candida valida and Trichosporon asahii* (El-Tarabily, 2004). Another pathogen control mechanism is parasitism, which is directly intract between yeast and pathogen. Yeast secrete lytic enzyme and develop upon the pathogenic fungi which causes death of pathogens. To control *Colletotrichum gloeosporioides*, the yeast *Candida famata, Rhodotorula mucilaginosa* use β -1,3-glucanase enzyme it causes parasitic effect on pathogens (Magallon-Andalon et al., 2012; Lima et al., 2013). Different genus and species of yeasts which suppress the plant pathogens are given in *Table 2*.

Yeast	Plant pathogen	References
A. pullulans	F. culmorum	Wachowska and Głowacka. (2014)
Saccharomyces cerevisiae	Penicillium expansum	Scherm et al. (2003)
Candida famata	Colletotrichum gloeosporioides	Magallon et al. (2012)
C. oleophila	P. expansum, B. cinerea	Jolanta Kowalska et al. (2022)
Cystofilobasidium infirmominiatum	Penicillium italicum	Vero et al. (2013)
M. pulcherrima	B. cinerea	Mondino et al. (2012)
M. fructicola	B. cinerea, Monilinia spp.	Jolanta Kowalska et al. (2022)
Aureobasidium pullulans	Penicillium expansum	Vero et al. (2013)
Candida oleophila	Botrytis and Penicillium	Ballet et al. (2015); Sebastien and Jijakli (2014); Mondino et al. (2012)

Table 2. Application of yeast against Plant pathogen

Insects

There are number of studies stating role of yeast in insecticidal activity. Combined inoculation of Cydia pomonella granulo virus and yeasts like *M. pulcherrima*, Cryptococcus tephrensis or Aerobasidium pullulans increase the death rate of larva of apple moth larval infestation (Knight and Witzgall, 2013). Sex pheromone for Helicoverpa armigera was prepared by the fermentation of Yarrowia lipolytica yeast that can be used as pheromone trap (Holkenbrink et al., 2020).

Abiotic stress

Drought

Drought reduces the plant yield and causes a severe stress among the crop plants (Posta et al., 2020). It causes harmful effects on growth of the plant by disturbing the activity of enzymes, ion uptake and assimilation of nutrients (Ahanger et al., 2017). Due to reduction of water the stomatal function will affect photosynthesis which decreases the yield of the crops (Osakabe et al., 2014). AM fungi can tolerate the drought and enhance water uptake (Posta et al., 2020). AMF produces larger volume of extraradicle hyphae which can helpful to tolerate the drought stress as water requirement of crop plants can be met out ever from the distant places (Gianinazzi et al., 2010). Another response of plant to mitigate stress is production of phytohormones. The balance of hormones controls plant's ability to withstand abiotic stress. The most basic signaling hormone during stress is abscisic acid (ABA), which affects root hydraulic conductivity,

transpiration rate and aquaporin expression (Ouledali et al., 2019). ABA reduces the water loss by closure of stomata. AM fungi inoculation will efficiently control the stomata by regulating the level of ABA (De Ollas and Dodd 2016; Ouledali et al., 2019). Some other phytohormones also play an important role in water stress are strigolactone and auxin (Mostofa et al., 2018). AM fungi enhance these phytohormones in response to drought stress (Ruiz-Lozano et al., 2016). AM Fungal species like *Rhizophagus irregularis, F. mosseae,* and *R. fasciculatus* give major response against drought stress in crops like maize, tomato and wheat (Allen et al., 1983; Bárzana et al., 2012; Chitarra et al., 2016). Influence of AMF against drought stress in various agricultural and horticultural crops such as sorghum, wheat, chilli, tomato, onion etc. along with the mechanism are given in *Table 3*.

Salinity

Salinity of the soil increases major problem in food security as salinity stress will reduce the plant development in the vegetative stage and also reduce yield. AM fungi are helpful for plants to tolerate salinity and increase the productivity (Talaat and Shawky, 2014; Abdel Latef and Chaoxing, 2014). AM fungi can also help to produce jasmonic acid, salicylic acid and some other inorganic acids (Hashem et al., 2018) in some crops like maize, wheat and tomato will tolerate against salinity stress with the inoculation of *R. irregularis, F. mosseae,* and *R. fasciculatus* (Daei et al., 2009; Hajiboland et al., 2009; Estrada et al., 2013). In *Antirrhinum majus* plants AM fungi increase the water potential and water use efficiency in salt stress conditions (El-Nashar, 2017).

Heavy metals

Mining sites and polluted sites with heavy metals contain AM fungi that are specifically adapted to soil pollution by heavy metals (Hildebrandt et al., 2007; Wang, 2017). Ecotypes are the AM fungi isolates which is mostly live in mine areas that highly polluted with heavy metals which has ability to withstand heavy metals depending on internal and external factors (Gildon and Tinker, 1981; Leyval et al., 1997; Joner and Leyval, 1997; Smith and Read, 2008). AM fungi retain the metal in the mycorrhizal plant roots and improve shoot biomass by restricting translocation to the aerial parts (Janeeshma and Puthur, 2020). Inoculation with AM fungi showed the best results in terms of percentage of seed germination, sustainability of seedlings, fresh weight and dry weight of plants. In two different heavy metal polluted soils, root colonization of maize plants with *Glomus* isolates reduced heavy metal solates like K, P, and Mg in roots (Kaldorf et al., 1999). Bioremediation of heavy metal is done by hyphal "metal binding" which decreases the availability of elements like Cu, Pb, Co, Cd, and Zn (Audet and Charest, 2007). Response of AM fungi on Plant stress is given in *Figure 1*.

Role of AM fungi and yeast in crop productivity

The effect of climate change on agricultural yield has posed a danger to global food security. Climate change such as increase in temperature, altered rainfall pattern, extreme weather that exaberate soil erosion, runoff cause adverse effect in crop production (Ahmed et al., 2015; Van der Linden and Goldberg, 2020).

Сгор	AMF	Response of AMF to drought stress condition	Reference
Sorghum	R. irregularis	Improved uptake of N ¹⁵	Symanczik et al. (2018)
Pistachio	G. etunicatum	Enhanced nutrient concentrations (P, N, K, Ca, Fe, Zn, and Cu), total chlorophyll, leaf area, proline content, flavonoids contents, CAT and POD activities soluble sugar, soluble proteins contents, shoot and root wt.	Abbaspour et al. (2012)
Wheat	R. intraradices, F. mosseae, F. geosporum	Increase in Relative Water Content	Mathur et al. (2018)
Lavender	R. irregularis F. mossea	Drecrease in antioxidant compounds like (glutathione, ascorbate and H2O2) Increase in N, K content, water content, biomass	Marulanda et al. (2007)
Wheat	F. geosporum	Stimulation of nutrient and water uptake	Ibrahim et al. (2011)
Chilli, Tomato	Rhizophagusfasciculatus Rhizophagusirregularis	Increased chlorophyll contents, , root length biomass, shoot length, and reduced proline content	Beltrano et al. (2013); Hajiboland et al. (2009)
Wheat	G. claroideum	Increased chlorophyll content and total dry weight	Beltrano et al. (2008)
Tomato, Corn	Rhizophagusirregularis	Increased in rate of apoplastic water flow	Bárzana et al. (2012)
Onion	Glomus etunicatus	Increased phosphorus content, fresh and dry wt.	Nelsen and Safir (1982)
Tomato	Rhizophagusirregularis	Increase in primary branches, plant height, fruit yields, shoot and root dry matter, , number of flowers and fruits, leaf relative water content (RWC), quality of fruits (less acidity and quantities of ascorbic acid and total soluble solids),N and P contents water use efficiency (WUE)	Subramanian et al. (2006)
White clover	R. irregularis	Increased nutrients content (P, K, Ca, Mg, Zn and B),glutathione reductase activity, proline concentrations, dry weight, relative water content	Ortiz et al. (2015)
Wheat	R. fasciculatus F. mosseae	Increased leaf osmotic adjustment and stomatal conductance	Allen et al. (1983)
Tomato	F. mosseae R. irregularis	Increased proline concentrations, stomatal density, plant height and biomass, , capacity to absorb CO2, intrinsic water use efficiency (iWUE) index,and reduction in leaf and root ABA contents, hydrogen peroxide	Chitarra et al. (2016)
Barley	Glomus intraradices	Increased activity of phosphatase enzyme, , Phosphorus content root volume and phosphatase enzyme activity	Bayani et al. (2015)
Soyabean	AMF	Incerased leaf area index, leaf proline, relative growth rate, photosynthesis rate, fresh weight and dry weight of seeds	Pavithra and Yapa (2018)
Sweet potato	Glomus spp	Enhanced soluble sugars, Proline adjustment in osmotic potential	Yooyongwech et al. (2016)

Table 3. Influence of AMF against drought stress

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Crop	AMF	Response of AMF to drought stress condition	Reference
Trifoliate orange	Funneliformismosseae, Paraglomusoccultum	Increased hyphal water absorption rate, leaf water potential and hyphal length	Zhang et al. (2018)
	Rhizophagusintraradices	Increase in grain biomass and copper, iron, manganese, zinc and gliadins contents in grains	Goicoechea et al. (2016)
Geranium	Rhizophagusintraradices, Funneliformismosseae	Enhanced plant biomass, essential oil content, nutrient content, glomalin related soil proteins , plant biomass, and essential oil content	Amiri et al. (2015)
Bambara groundnut	Glomus intraradices, Gigasporagregaria, Scutellosporagregaria	Enhanced soluble sugars, mineral content, acid phosphatase, reduction proline content	Tsoata et al. (2015)
Snapdragon	Glomus deserticola	Increased leaf area, shoot length, shoot and root diameter, water content, leaf number per plant, Proline and chlorophyll	Asrar et al. (2012)
Black locust (pea)	Funneliformismosseae and Rhizophagusintraradices	Enhanced net photosynthetic rate and dry biomass, water use efficiency.	Yang et al. (2014)
Lettuce and tomato	Rhizophagusirregularis, Glomus intraradices	Enhanced biomass production, ABA accumulation and synthesis, strigolactone production efficiency of photosystem II	Ruiz-Lozano et al. (2016)
Hardy sugarcane	Glomus spp.	Enhanced antioxidant enzymes, levels of metabolites, phenolics, glutathione, ascorbic acid, chlorophyll fluorescence, leaf water uptake, levels of metabolites and plant biomass	Mirshad and Puthur (2016)
Wheat	Glomus mosseae	Enhanced osmotic potential, chlorophyll content and fluorescence, activities of antioxidant enzymes, ascorbic acid, enzymes of N and P metabolism, and contents of N, P, and K	Rani et al. (2018)
Pangola grass	Rhizophagusirregularis	Enhanced stomatal conductance, peroxidation, shoot dry matter, lipid, H ₂ O ₂ in shoot and root	Pedranzani et al. (2016)
Strawberry	F. mosseae BEG25, F. geosporus BEG11	Increase in WUE, plant survival, shoot and root fresh weights	Boyer et al. (2016)
Wheat	Glomus mosseae, Glomus fasciculatum, Gigasporadecipiens	Increase in total chlorophyll pigments and other plant growth parameters	Pal and Pandey. (2016)

To mitigate this problem Arbuscular Mycorrhizal Fungi is helpful in agriculture as it plays a crucial role in mitigating the impacts of climate change on agriculture, particularly through their influence on phosphorus cycling and availability. Arbuscular Mycorrhizal fungi enhances phosphorus uptake, reduces the need for chemical fertilizers, improves soil carbon sequestration and increases plant resilience to climate stress. In tropical soils, Arbuscular Mycorrhizal fungi play an important role as phosphorus content is very less in tropical soils (Adisa et al., 2019). AMF consume only 25% of phosphorus another 75% of phosphorus will be utilized by the plants. AMF and plant symbiosis play an important role in crop productivity and ecosystem functioning. They are the key importance for sustainable crop development (Gianinazzi et al., 2010).

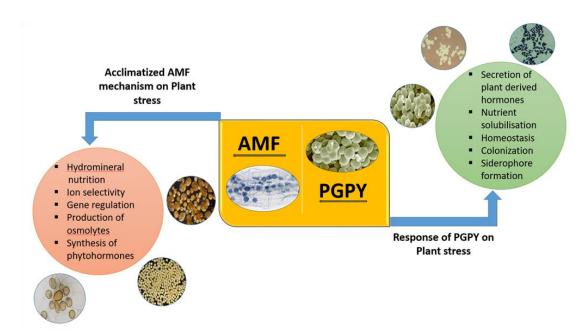


Figure 1. Response of AM fungi on plant stress

Arbuscular mycorrhizal fungi is helpful in vast production of maize (Sabia et al., 2015), elephant foot yam (Lu et al., 2015) and potato (Hijri, 2016). It has the greater potential for improving the crop production and also improves the synthesis of phytochemicals in food crops and prepares them good for consumption (Sbrana et al., 2014; Rouphael et al., 2015). In tomato Fusarium oxysporum f. sp. Lycopersici is the disease causing pathogen which affects the plants, by using AMF it can produce antimicrobial properties and restrict the mycelial development of the pathogen. Reduced disease incidence may increase the growth of the crops, biomass, nitrogen, phosphorus, and potassium content, and productivity of the crop (Kumari et al., 2019). AMF can improve soil physical characteristics such as available nutrients in soil, water retention capacity, activity of microorganisms in soil, C, N, P cycle by the improvement of soil characteristics it is helpful for enhancing the production and productivity of the crop plants (Sadhana, 2014; Jamiołkowska et al., 2018; Parihar et al., 2020). Combined inoculation of AMF and nitrogen fixing bacteria will increase the nodulation potential, fixation of Nitrogen and plant productivity (Herrera et al., 1993). In agroforestry crops, Schizolobium parahyba is inoculated with AMF and PGPR will improve the quality of wood and increase the weight of wood by 20% when compare to the inorganic fertilizers (Blake, 1919). AMF also give major impact on the crops under salinity stress conditions, inoculation of AMF will improve the growth rate and productivity of crops under saline conditions (Talaat and Shawky, 2014; Abdel Latef and Chaoxing, 2014; El-Nashar, 2017).

Reduction in Nitrogen level will cause severe decrease in crop productivity to mitigate this problem yeast can be helpful for increasing the nutrient availability to the crop plants (Leghari et al., 2016). Yeast mixed with biofertilizers will improve the plant weight, dry matter, grain and straw yield of the rice crop (El-Sirafy et al., 2011). Which increases the productivity in marigold crops the foliar application of yeast will enhance the plant growth and productivity (Taha et al., 2020). In various crops *Saccharomyces cerevisiae* increases the plant growth and productivity characteristics (Al-Maeini and

Al-Isawi, 2017) spraying of yeast extract hives the high weight of seeds. Foliar application of yeast may increases the plant growth stimulants, like gibberellins, auxins, and cytokines, which promote cell division and growth. The primary effect of foliar yeast spray may be that yeast increases nutrient mineral absorption through overall improvement (Ahmed et al., 2023). Efficiency of Arbuscular mycorrhyzal fungi and Plant growth promoting yeast on crop productivity is given in *Figure 2*.

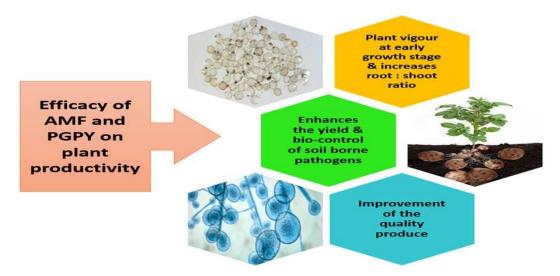


Figure 2. Efficiency of Arbuscular mycorrhyzal fungi and Plant growth promoting yeast on crop productivity

Interaction with other biofertilizer

Most of the biofertilizers will interact with AM fungi. Some of the strains of *Rhizobium* will combinely inoculated with AM fungi which will improves the Nitrogen content, increases the number of root nodules and rate of mycorrhizal infection. This type of symbiosis will give positive effect in Legume crops (Razakatiana et al., 2020). AM fungi also favours the legume crops for their nodulation under some extreme conditions like low fertility (Razakatiana et al., 2020), flood and drought conditions (Hao et al., 2019). The fungal biocontrol agent like *Trichoderma viride* and AM fungi is combinely helps in improving the growth parameters, chlorophyll and carotenoid contents (Metwally et al., 2020). Phosphate solubilizing bacteria like *P. fluorescence* and *B. megaterium* are inoculated with AM fungi which will improve the solubilization of phosphate which helps to increase productivity. Agroforestry crops like Eucalyptus will be inoculated by AM Fungi and beneficial microbes will improve the quality of seedlings by improving the biomass and in matured plants it will increases the wood quality (Karthikeyan and Prakash, 2008).

Endophytic yeast increases the nod factor when inoculate with *Rhizobium sp* in legume crops (Geetha et al., 2020). Yeast like *Saccharomyces cerevisiae* will increases the formation of nodules, length and dryweight of the crop plants when inoculated with *Rhizobium trifolii* (Tuladhar and Rao, 1985). Interaction of *Bacillus megaterium* with yeast will increase the concentration of available phosphorus in soil (Abdul-Hassein and Hassan, 2021), these types of interaction is cost effective and ecofriendly (Karthikeyan and Prakash, 2008).

Conclusion

This review focused on favorable effect of AM fungi and plant growth promoting yeast on crop plants as potential biological agents, role in uptake of nutrients, increasing crop productivity and tolerates biotic and abiotic stress conditions. AMF treated plants have double the amount of biomass compared to the control plants. Compared to control plants, plant growth promoting yeast produces two-fold times of antioxidant enzymes that help the plants to overcome biotic and abiotic stress. Nowadays there is an increase in chemical fertilizers and pesticides which increases hazardous damage to the environment. To mitigate this problem AM fungi and PGPY can be effectively and efficiently used. Recent studies concluded that AM fungi and PGPY is more helpful in nutrient uptake, mobilization of nutrients, soil health, bioremediation, pests and disease management. Co inoculation of yeast and AM fungi play some specific role in plant development, reduction in usage of chemical fertilizers and pesticides, which ultimately results in sustainable agriculture.

REFERENCES

- [1] Abbaspour, H., Saeidi-Sar, S., Afshari, H., Abdel-Wahhab, M. (2012): Tolerance of mycorrhiza infected pistachio (*Pistacia vera* L.) seedling to drought stress under glasshouse conditions. Journal of plant physiology 169(7): 704-709.
- [2] Abdel Latef, A. A. H., Chaoxing, H. (2014): Does inoculation with *Glomus mosseae* improve salt tolerance in pepper plants? Journal of Plant Growth Regulation 33: 644-653.
- [3] Abdul-Hassein, H. M., Hassan, K. U. (2021): Effect of the Biofertilizer (*Bacillus Megaterium*) and the Addition of Yeast Spraying on the Vegetative growths in Phosphorous Availability, Growth and Yield of Onions *Allium cepa* L. Bionatura: Latin American Journal of Biotecnology and Life Sciences 13(6).
- [4] Abigail, E. A., Salam, J. A., Das, N. (2013): Atrazine degradation in liquid culture and soil by a novel yeast *Pichia kudriavzevii* strain Atz-EN-01 and its potential application for bioremediation. Journal of Applied Pharmaceutical Science 3(6): 35-43.
- [5] Adisa, I. O., Pullagurala, V. L. R., Peralta-Videa, J. R., Dimkpa, C. O., Elmer, W. H., Gardea-Torresdey, J. L., White, J. C. (2019): Recent advances in nano-enabled fertilizers and pesticides: a critical review of mechanisms of action. – Environmental Science: Nano 6(7): 2002-2030.
- [6] Agamy, R., Hashem, M., Alamri, S. (2013): Effect of soil amendment with yeasts as biofertilizers on the growth and productivity of sugar beet. – African Journal of Agricultural Research 8(1): 46-56.
- [7] Ahanger, M. A., Tittal, M., Mir, R. A., Agarwal, R. (2017): Alleviation of water and osmotic stress-induced changes in nitrogen metabolizing enzymes in *Triticum aestivum* L. cultivars by potassium. Protoplasma 254: 1953-1963.
- [8] Ahmed, K. F., Wang, G., Yu, M., Koo, J., You, L. (2015): Potential impact of climate change on cereal crop yield in West Africa. Climatic Change 133: 321-334.
- [9] Ahmed, A. A. O., Salah, A., Sedik, F., Ghanim, A. (2023): Effect of foliar treatment with yeast and nitrogen fertilization on the productivity of sesame. Aswan University Journal of Environmental Studies 4(2): 15-24.
- [10] Akköprü, A., Demir, S. (2005): Biological control of *Fusarium* wilt in tomato caused by *Fusarium oxysporum f. sp. lycopersici* by AMF *Glomus intraradices* and some *Rhizobacteria.* Journal of Phytopathology 153(9): 544-550.
- [11] Allen, M. F., Boosalis, M. G. (1983): Effects of two species of VA mycorrhizal fungi on drought tolerance of winter wheat. New Phytologist 93(1): 67-76.

http://www.aloki.hu • ISSN 1589 1623 (Print) • ISSN 1785 0037 (Online)

- [12] Allen, M. F. (2011): Linking water and nutrients through the vadose zone: a fungal interface between the soil and plant systems. Journal of Arid Land 3(3): 155-163.
- [13] Al-Maeini, W., Al-Isawi, Y. (2017): Effects of foliar application of yeast extract on seed yield, and seed yield components of five sorghum cultivars. ANBAR Journal of Agricultural sciences 15(1):152-161.
- [14] Amiri, R., Nikbakht, A., Etemadi, N. (2015): Alleviation of drought stress on rose geranium [*Pelargonium graveolens* (L.) Herit.] in terms of antioxidant activity and secondary metabolites by mycorrhizal inoculation. – Scientia Horticulturae 197: 373-380.
- [15] Amprayn, K.-O., Rose, M. T., Kecskés, M., Pereg, L., Nguyen, H. T., Kennedy, I. R. (2012): Plant growth promoting characteristics of soil yeast (*Candida tropicalis HY*) and its effectiveness for promoting rice growth. – Applied Soil Ecology 61: 295-299.
- [16] Anderson, R., Keshwani, D., Guru, A., Yang, H., Irmak, S., Subbiah, J. (2018): An integrated modeling framework for crop and biofuel systems using the DSSAT and GREET models. – Environmental Modelling & Software 108: 40-50.
- [17] Asrar, A., Abdel-Fattah, G., Elhindi, K. (2012): Improving growth, flower yield, and water relations of snapdragon (*Antirhinum majus* L.) plants grown under well-watered and water-stress conditions using arbuscular mycorrhizal fungi. Photosynthetica 50: 305-316.
- [18] Audet, P., Charest, C. (2007): Dynamics of arbuscular mycorrhizal symbiosis in heavy metal phytoremediation: meta-analytical and conceptual perspectives. Environmental Pollution 147(3): 609-614.
- [19] Ballet, N., Souche, J., Vandekerckove, P. (2015): Efficacy of *Candida oleophila*, strain O, in preventing postharvest diseases of fruits. – Paper presented at the III International Symposium on Postharvest Pathology: Using Science to Increase Food Availability 1144.
- [20] Barnett, J. A., Barnett, L. (2011): Yeast research: a historical overview. American Society for Microbiology Press.
- [21] Bárzana, G., Aroca, R., Paz, J. A., Chaumont, F., Martinez-Ballesta, M. C., Carvajal, M., Ruiz-Lozano, J. M. (2012): Arbuscular mycorrhizal symbiosis increases relative apoplastic water flow in roots of the host plant under both well-watered and drought stress conditions. – Annals of Botany 109(5): 1009-1017.
- [22] Baslam, M., Garmendia, I., Goicoechea, N. (2011): Arbuscular mycorrhizal fungi (AMF) improved growth and nutritional quality of greenhouse-grown lettuce. Journal of Agricultural and Food Chemistry 59(10): 5504-5515.
- [23] Bayani, R., Saateyi, A., Faghani, E. (2015): Influence of arbuscular mycorrhiza in phosphorus acquisition efficiency and drought-tolerance mechanisms in barley. – International Journal of Biosciences 7(1): 86-94.
- [24] Begum, N., Qin, C., Ahanger, M. A., Raza, S., Khan, M. I., Ashraf, M., Nadeem, A., Zhang, L. (2019): Role of arbuscular mycorrhizal fungi in plant growth regulation: implications in abiotic stress tolerance. – Frontiers in Plant Science 10: 1068.
- [25] Beltrano, J., Ronco, M. G. (2008): Improved tolerance of wheat plants (*Triticum aestivum* L.) to drought stress and rewatering by the arbuscular mycorrhizal fungus *Glomus claroideum*: Effect on growth and cell membrane stability. Brazilian Journal of Plant Physiology 20: 29-37.
- [26] Beltrano, J., Ruscitti, M., Arango, C., Ronco, M. (2013): Changes in the accumulation of shikimic acid in mycorrhized *Capsicum annuum* L. grown with application of glyphosate and phosphorus. – Theoretical and Experimental Plant Physiology 25: 125-136.
- [27] Bencheqroun, S. K., Bajji, M., Massart, S., Labhilili, M., El Jaafari, S., Jijakli, M. H. (2007): In vitro and in situ study of postharvest apple blue mold biocontrol by *Aureobasidium pullulans*: evidence for the involvement of competition for nutrients. – Postharvest Biology and Technology 46(2): 128-135.
- [28] Bennett, A. E., Groten, K. (2022): The costs and benefits of plant–arbuscular mycorrhizal fungal interactions. Annual Review of Plant Biology 73: 649-672.

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- [29] Blake, S. F. (1919): New South American Spermatophytes collected by HM Curran. Contributions from the United States National Herbarium 20(7): 237-245.
- [30] Bona, E., Cantamessa, S., Massa, N., Manassero, P., Marsano, F., Copetta, A., Guido, L., Giovanni, A., Elisa, G., Berta, G. (2017): Arbuscular mycorrhizal fungi and plant growthpromoting *pseudomonads* improve yield, quality and nutritional value of tomato: a field study. – Mycorrhiza 27: 1-11.
- [31] Brundrett, M. C. (2009): Mycorrhizal associations and other means of nutrition of vascular plants: understanding the global diversity of host plants by resolving conflicting information and developing reliable means of diagnosis. Plant and Soil 320: 37-77.
- [32] Calvente, V., Benuzzi, D., de Tosetti, M. S. (1999): Antagonistic action of siderophores from *Rhodotorula glutinis* upon the postharvest pathogen *Penicillium expansum.* International Biodeterioration & Biodegradation 43(4): 167-172.
- [33] Canton, G. C., Bertolazi, A. A., Cogo, A. J., Eutrópio, F. J., Melo, J., de Souza, S. B., Krohling, C., Campostrini, E., da Silva, A. G., Façanha, A. R. (2016): Biochemical and ecophysiological responses to manganese stress by ectomycorrhizal fungus *Pisolithus tinctorius* and in association with *Eucalyptus grandis*. – Mycorrhiza 26: 475-487.
- [34] Cao, J., Wang, C., Huang, Y. (2015): Interactive impacts of earthworms (*Eisenia fetida*) and arbuscular mycorrhizal fungi (*Funneliformis mosseae*) on the bioavailability of calcium phosphates. Plant and Soil 396: 45-57.
- [35] Castellanos-Morales, V., Villegas, J., Wendelin, S., Vierheilig, H., Eder, R., Cárdenas-Navarro, R. (2010): Root colonisation by the arbuscular mycorrhizal fungus *Glomus intraradices* alters the quality of strawberry fruits (Fragaria× ananassa Duch.) at different nitrogen levels. – Journal of the Science of Food and Agriculture 90(11): 1774-1782.
- [36] Chen, S., Zhao, H., Zou, C., Li, Y., Chen, Y., Wang, Z., Jiang, Y., Liu, A., Zhao, P., Wang, M., Ahammed, G. J. (2017): Combined inoculation with multiple arbuscular mycorrhizal fungi improves growth, nutrient uptake and photosynthesis in cucumber seedlings. – Frontiers in Microbiology 8: 2516.
- [37] Chitarra, W., Pagliarani, C., Maserti, B., Lumini, E., Siciliano, I., Cascone, P., Schubert, A., Gambino, G., Balestrini, R., Guerrieri, E. (2016): Insights on the impact of arbuscular mycorrhizal symbiosis on tomato tolerance to water stress. – Plant Physiology 171(2): 1009-1023.
- [38] Aljawasim, B. D., Khaeim, H. M., Manshood, M. (2020): Assessment of arbuscular mycorrhizal fungi (*Glomus spp.*) as potential biocontrol agents against damping-off disease *Rhizoctonia solani* on cucumber. Journal of Crop Protection 9(1): 141-147.
- [39] Daei, G., Ardekani, M., Rejali, F., Teimuri, S., Miransari, M. (2009): Alleviation of salinity stress on wheat yield, yield components, and nutrient uptake using arbuscular mycorrhizal fungi under field conditions. – Journal of Plant Physiology 166(6): 617-625.
- [40] Dai, Y.-J., Ji, W.-W., Chen, T., Zhang, W.-J., Liu, Z.-H., Ge, F., Yuan, S. (2010): Metabolism of the neonicotinoid insecticides acetamiprid and thiacloprid by the yeast *Rhodotorula mucilaginosa* strain IM-2. – Journal of agricultural and food chemistry 58(4): 2419-2425.
- [41] De Ollas, C., Dodd, I. C. (2016): Physiological impacts of ABA–JA interactions under water-limitation. Plant Molecular Biology 91: 641-650.
- [42] Duponnois, R., Ouahmane, L., Kane, A., Thioulouse, J., Hafidi, M., Boumezzough, A., Prin, Y., Baudoin, E., Galiana, A., Dreyfus, B. (2011): Nurse shrubs increased the early growth of Cupressus seedlings by enhancing belowground mutualism and soil microbial activity. – Soil Biology and Biochemistry 43(10): 2160-2168.
- [43] El-Nashar, Y. (2017): Response of snapdragon (*Antirrhinum majus* L.) to blended water irrigation and arbuscular mycorrhizal fungi inoculation: uptake of minerals and leaf water relations. Photosynthetica 55: 201-209.
- [44] El-Sirafy, Z., Abd-El-Hameed, A., El-Mahdy, R. E. (2011): Role of biofertilization and different rates of nitrogen with foliar spraying of nitrogen and yeast in rie productivity

and nutrient uptakes. – Journal of Soil Sciences and Agricultural Engineering 2(7): 717-731.

- [45] El-Zohri, M., Medhat, N., Saleh, F.-E. M., El-Maraghy, S. S. (2017): Some biofertilizers relieved the stressful drawbacks of calcareous soil upon black seed (*Nigella sativa* L.) through inhibiting stress markers and antioxidant enzymes with enhancing plant growth. Egyptian Journal of Botany 57(1): 75-92.
- [46] Estrada, B., Aroca, R., Maathuis, F. J., Barea, J. M., Ruiz-Lozano, J. M. (2013): Arbuscular mycorrhizal fungi native from a Mediterranean saline area enhance maize tolerance to salinity through improved ion homeostasis. – Plant, Cell & Environment 36(10): 1771-1782.
- [47] FAOSTAT (2019): Statistical Database. Food and Agriculture Organization of the United Nations [FAO]. Retrieved from http://www.fao.org/faostat/en/.
- [48] Fernandez-San Millan, A., Farran, I., Larraya, L., Ancin, M., Arregui, L., Veramendi, J. (2020): Plant growth-promoting traits of yeasts isolated from Spanish vineyards: Benefits for seedling development. – Microbiological Research 237: 126480.
- [49] Ferraz, P., Cássio, F., Lucas, C. (2019): Potential of yeasts as biocontrol agents of the phytopathogen causing cacao witches' broom disease: is microbial warfare a solution? – Frontiers in Microbiology 10: 1766.
- [50] Freimoser, F. M., Rueda-Mejia, M. P., Tilocca, B., Migheli, Q. (2019): Biocontrol yeasts: Mechanisms and applications. – World Journal of Microbiology and Biotechnology 35: 1-19.
- [51] Fu, S.-F., Sun, P.-F., Lu, H.-Y., Wei, J.-Y., Xiao, H.-S., Fang, W.-T., Cheng, B. Y., Chou, J.-Y. (2016): Plant growth-promoting traits of yeasts isolated from the phyllosphere and rhizosphere of *Drosera spatulata Lab.* Fungal Biology 120(3): 433-448.
- [52] Geetha Thanuja, K., Annadurai, B., Thankappan, S., Uthandi, S. (2020): Non-rhizobial endophytic (NRE) yeasts assist nodulation of Rhizobium in root nodules of blackgram (*Vigna mungo L.*). Archives of Microbiology 202: 2739-2749.
- [53] George, N. P., Ray, J. G. (2023): The inevitability of arbuscular mycorrhiza for sustainability in organic agriculture-a critical review. Frontiers in Sustainable Food Systems 7: 1124688.
- [54] Gianinazzi, S., Gollotte, A., Binet, M.-N., van Tuinen, D., Redecker, D., Wipf, D. (2010): Agroecology: the key role of arbuscular mycorrhizas in ecosystem services. – Mycorrhiza 20(8): 519-530.
- [55] Gildon, A., Tinker, P. (1981): A heavy metal-tolerant strain of a mycorrhizal fungus. Transactions of the British Mycological society 77(DEC): 648-649.
- [56] Goicoechea, N., Bettoni, M. M., Fuertes-Mendizabal, T., González-Murua, C., Aranjuelo, I. (2016): Durum wheat quality traits affected by mycorrhizal inoculation, water availability and atmospheric CO₂ concentration. – Crop and Pasture Science 67(2): 147-155.
- [57] Gomathy, M., Sabarinathan, K., Sivasankari Devi, T., Pandiyarajan, P. (2018): Arbuscular mycorrhizal fungi and glomalin-super glue. – Intrenational Journal of Current Microbiology and Applied Sciences 7(7): 2853-2857.
- [58] Gomathy, M., Sabarinathan, K., Subramanian, K., Ananthi, K., Kalaiyarasi, V., Jeyshri, M., Dutta, P. (2021): Niche for microbial rejuvenation and biodegradation of pollutants. Microbial Rejuvenation of Polluted Environment. – Rhizosphere 1: 1-22.
- [59] Gomathy, M., Sabarinathan, K., Subramaian, K., Sivashankari Devi, T., Ananthi, K., Kalaiselvi, P., Jeyshree, M. (2022): Microbial remediation of chromium. – In: Microbial metabolism of metals and metalloids, Springer, pp. 255-278.
- [60] Hajiboland, R., Aliasgharzad, N., Barzeghar, R. (2009): Influence of arbuscular mycorrhizal fungi on uptake of Zn and P by two contrasting rice genotypes. – Plant Soil Environment 55(3): 93-100.

- [61] Han, S., Kim, G.-Y., Han, J.-I. (2019): Biodiesel production from oleaginous yeast, *Cryptococcus sp.* by using banana peel as carbon source. Energy Reports 5: 1077-1081.
- [62] Hao, Z., Xie, W., Jiang, X., Wu, Z., Zhang, X., Chen, B. (2019): Arbuscular mycorrhizal fungus improves rhizobium–glycyrrhiza seedling symbiosis under drought stress. Agronomy 9(10): 572.
- [63] Harrison, M. J. (1999): Molecular and cellular aspects of the arbuscular mycorrhizal symbiosis. Annual review of plant biology 50(1): 361-389.
- [64] Hashem, A., Abd Allah, E. F., Alqarawi, A. A., Egamberdieva, D. (2018): Arbuscular mycorrhizal fungi and plant stress tolerance. – Plant microbiome: stress response, pp. 81-103.
- [65] Hassan, H., Mohamed, M. T. M., Yusoff, S. F., Hata, E. M., Tajidin, N. E. (2021): Selecting antagonistic yeast for postharvest biocontrol of *Colletotrichum gloeosporioides* in papaya fruit and possible mechanisms involved. – Agronomy 11(4): 760.
- [66] Hernández-Fernández, M., Cordero-Bueso, G., Ruiz-Muñoz, M., Cantoral, J. M. (2021): Culturable yeasts as biofertilizers and biopesticides for a sustainable agriculture: A comprehensive review. – Plants 10(5): 822.
- [67] Herrera, M., Salamanca, C., Barea, J. (1993): Inoculation of woody legumes with selected arbuscular mycorrhizal fungi and rhizobia to recover desertified Mediterranean ecosystems. Applied and Environmental Microbiology 59(1): 129-133.
- [68] Hesham, A. E.-L., Mohamed, H. M. (2011): Molecular genetic identification of yeast strains isolated from Egyptian soils for solubilization of inorganic phosphates and growth promotion of corn plants. Journal of microbiology and biotechnology 21(1): 55-61.
- [69] Hesham, A., Alrumman, S., Al-Dayel, M., Salah, H. (2017): Screening and genetic identification of acidic and neutral protease-producing yeasts strains by 26S rRNA gene sequencing. Cytology and Genetics 51: 221-229.
- [70] Hesham, A. E.-L., Alrumman, S. A., Al-Qahtani, A. D. S. (2018): Degradation of toluene hydrocarbon by isolated yeast strains: molecular genetic approaches for identification and characterization. Russian Journal of Genetics 54: 933-943.
- [71] Hijri, M. (2016): Analysis of a large dataset of mycorrhiza inoculation field trials on potato shows highly significant increases in yield. Mycorrhiza 26(3): 209-214.
- [72] Hildebrandt, U., Regvar, M., Bothe, H. (2007): Arbuscular mycorrhiza and heavy metal tolerance. Phytochemistry 68(1): 139-146.
- [73] Holkenbrink, C., Ding, B.-J., Wang, H.-L., Dam, M. I., Petkevicius, K., Kildegaard, K. R., Wenning, L., Sinkwitz, C., Lorántfy, B., Koutsoumpeli, E., Koutsoumpeli, E. (2020): Production of moth sex pheromones for pest control by yeast fermentation. Metabolic Engineering 62: 312-321.
- [74] Huang, R., Che, H., Zhang, J., Yang, L., Jiang, D., Li, G. (2012): Evaluation of *Sporidiobolus pararoseus* strain YCXT3 as biocontrol agent of *Botrytis cinerea* on postharvest strawberry fruits. – Biological Control 62(1): 53-63.
- [75] Humphreys, C. P., Franks, P. J., Rees, M., Bidartondo, M. I., Leake, J. R., Beerling, D. J. (2010): Mutualistic mycorrhiza-like symbiosis in the most ancient group of land plants. – Nature Communications 1(1): 103.
- [76] Ibijbijen, J., Urquiaga, S., Ismaili, M., Alves, B., Boddey, R. (1996): Effect of arbuscular mycorrhizal fungi on growth, mineral nutrition and nitrogen fixation of three varieties of common beans (*Phaseolus vulgaris*). – New Phytologist 134(2): 353-360.
- [77] Ibrahim, A., Abdel-Fattah, G., Eman, F., Abd El-Aziz, M., Shohr, A. (2011): Arbuscular mycorrhizal fungi and spermine alleviate the adverse effects of salinity stress on electrolyte leakage and productivity of wheat plants. Phyton 51(2): 261-276.
- [78] Ibrahim, H. A., El-Fiki, I. (2019): Study on the effect of yeast in compost tea efficiency in controlling chocolate leaf spot disease in broad bean (*Vicia faba*). – Organic Agriculture 9: 175-188.

- [79] Ismail, Y., McCormick, S., Hijri, M. (2013): The arbuscular mycorrhizal fungus, *Glomus irregulare*, controls the mycotoxin production of *Fusarium sambucinum* in the pathogenesis of potato. FEMS microbiology letters 348(1): 46-51.
- [80] Jamiołkowska, A., Księżniak, A., Gałązka, A., Hetman, B., Kopacki, M., Skwaryło-Bednarz, B. (2018): Impact of abiotic factors on development of the community of arbuscular mycorrhizal fungi in the soil: a Review. International Agrophysics 32(1): 133-140.
- [81] Janeeshma, E., Puthur, J. T. (2020): Direct and indirect influence of arbuscular mycorrhizae on enhancing metal tolerance of plants. – Archives of Microbiology 202: 1-16.
- [82] Jeyashri, M., Gomathy, M., Sabarinathan, K., Subhashini, R., Suresh, S. (2019): Screening of Phyllosphere Yeast of Rice for the Production Enzymes and Solubilisation of Minerals. – International Journal of Current Microbiology and Applied Sciences 8(8): 465-472.
- [83] Joner, E., Leyval, C. (1997): Uptake of 109Cd by roots and hyphae of a *Glomus mosseae Trifolium subterraneum* mycorrhiza from soil amended with high and low concentrations of cadmium. The New Phytologist 135(2): 353-360.
- [84] Jung, S. C., Martinez-Medina, A., Lopez-Raez, J. A., Pozo, M. J. (2012): Mycorrhizainduced resistance and priming of plant defenses. – Journal of Chemical Ecology 38: 651-664.
- [85] Kaldorf, M., Kuhn, A., Schröder, W., Hildebrandt, U., Bothe, H. (1999): Selective element deposits in maize colonized by a heavy metal tolerance conferring arbuscular mycorrhizal fungus. – Journal of Plant Physiology 154(5-6): 718-728.
- [86] Kamel, S., Ebtsam, M. M., Massoud, O. (2016): Potentiality of some yeast species as biocontrol agents against *Fusarium oxysporum f. sp. cucumerinum* the causal agent of cucumber wilt. – Egyptian Journal of Biological Pest Control 26(2): 185-193.
- [87] Karthikeyan, A., Prakash, M. S. (2008): Effects of arbuscular mycorrhizal fungi, Phosphobacterium and Azospirillum sp. on the successful establishment of *Eucalyptus camaldulensis* Dehn. in bauxite mine spoils. – Forests, Trees and Livelihoods 18(2): 183-191.
- [88] Khan, Y., Yang, X., Zhang, X., Yaseen, T., Shi, L., Zhang, T. (2021): Arbuscular mycorrhizal fungi promote plant growth of *Leymus chinensis* (Trin.) Tzvelev by increasing the metabolomics activity under nitrogen addition. Grassland Science 67(2): 128-138.
- [89] Kivlin, S. N., Hawkes, C. V., Treseder, K. K. (2011): Global diversity and distribution of arbuscular mycorrhizal fungi. Soil Biology and Biochemistry 43(11): 2294-2303.
- [90] Knight, A. L., Witzgall, P. (2013): Combining mutualistic yeast and pathogenic virus-a novel method for codling moth control. Journal of Chemical Ecology 39: 1019-1026.
- [91] Korpi, A., Järnberg, J., Pasanen, A.-L. (2009): Microbial volatile organic compounds. Critical reviews in toxicology 39(2): 139-193.
- [92] Kowalska, J., Krzymińska, J., Tyburski, J. (2022): Yeasts as a potential biological agent in plant disease protection and yield improvement-A short review. – Agriculture 12(9): 1404.
- [93] Kumari, S. M. P., Prabina, B. J. (2019): Protection of tomato, *Lycopersicon esculentum* from wilt pathogen, *Fusarium oxysporum f. sp. lycopersici* by arbuscular mycorrhizal fungi, *Glomus sp.* International Journal of Current Microbiology and Applied Sciences 8: 1368-1378.
- [94] Kumla, J., Nundaeng, S., Suwannarach, N., Lumyong, S. (2020): Evaluation of multifarious plant growth promoting trials of yeast isolated from the soil of assam tea (*Camellia sinensis var. assamica*) plantations in Northern Thailand. – Microorganisms 8(8): 1168.
- [95] Leghari, S. J., Wahocho, N. A., Laghari, G. M., HafeezLaghari, A., MustafaBhabhan, G., HussainTalpur, K., Bhutto, T. A., Wahocho, S. A., Lashari, A. A. (2016): Role of

nitrogen for plant growth and development: A review. – Advances in Environmental Biology 10(9): 209-219.

- [96] Leyval, C., Turnau, K., Haselwandter, K. (1997): Effect of heavy metal pollution on mycorrhizal colonization and function: physiological, ecological and applied aspects. – Mycorrhiza 7: 139-153.
- [97] Li, X., Zeng, R., Liao, H. (2016): Improving crop nutrient efficiency through root architecture modifications. Journal of Integrative Plant Biology 58(3): 193-202.
- [98] Lima, J., Gondim, D. M., Oliveira, J., Oliveira, F. S., Gonçalves, L. R., Viana, F. M. (2013): Use of killer yeast in the management of postharvest papaya anthracnose. – Postharvest Biology and Technology 83: 58-64.
- [99] Liu, L., Li, J., Yue, F., Yan, X., Wang, F., Bloszies, S., Wang, Y. (2018): Effects of arbuscular mycorrhizal inoculation and biochar amendment on maize growth, cadmium uptake and soil cadmium speciation in Cd-contaminated soil. – Chemosphere 194: 495-503.
- [100] Lu, F.-C., Lee, C.-Y., Wang, C.-L. (2015): The influence of arbuscular mycorrhizal fungi inoculation on yam (*Dioscorea spp.*) tuber weights and secondary metabolite content. – Peer J 3: e1266.
- [101] Magallon-Andalon, C., Luna-Solano, G., Ragazzo-Sanchez, J., Calderon-Santoyo, M. (2012): Parasitism and substrate competitions effect of antagonistic yeasts for biocontrol of *Colletotrichum gloeosporioides* in papaya (*Carica papaya* L.) var Maradol. – Mexican Journal of Scientific Research 1: 2-9.
- [102] Marulanda, A., Porcel, R., Barea, J. M., Azcón, R. (2007): Drought tolerance and antioxidant activities in lavender plants colonized by native drought-tolerant or droughtsensitive Glomus species. – Microbial Ecology 54: 543-552.
- [103] Mathur, S., Sharma, M. P., Jajoo, A. (2018): Improved photosynthetic efficacy of maize (*Zea mays*) plants with arbuscular mycorrhizal fungi (AMF) under high temperature stress. – Journal of Photochemistry and Photobiology B: Biology 180: 149-154.
- [104] Mei, L., Yang, X., Cao, H., Zhang, T., Guo, J. (2019): Arbuscular mycorrhizal fungi alter plant and soil C: N: P stoichiometries under warming and nitrogen input in a semiarid meadow of China. – International Journal of Environmental Research and Public Health 16(3): 397.
- [105] Mekki, B., Ahmed, A. G. (2005): Growth, yield and seed quality of soybean (*Glycine max* L.) as affected by organic, biofertilizer and yeast application. Research Journal of Agriculture and Biological Sciences 1(4): 320-324.
- [106] Metwally, R., Al-Amri, S. (2020): Individual and interactive role of *Trichoderma viride* and arbuscular mycorrhizal fungi on growth and pigment content of onion plants. Letters in Applied Microbiology 70(2): 79-86.
- [107] Mirshad, P., Puthur, J. T. (2016): Arbuscular mycorrhizal association enhances drought tolerance potential of promising bioenergy grass (*Saccharum arundinaceum* Retz.). – Environmental Monitoring and Assessment 188: 1-20.
- [108] Mitra, D., Uniyal, N., Panneerselvam, P., Senapati, A., Ganeshamurthy, A. N. (2020): Role of mycorrhiza and its associated bacteria on plant growth promotion and nutrient management in sustainable agriculture. – International Journal of Life Sciences and Applied Sciences 1(1): 1-1.
- [109] Mohamed, H., Saad, A. (2009): The biocontrol of postharvest disease (*Botryodiplodia theobromae*) of guava (*Psidium guajava* L.) by the application of yeast strains. Postharvest Biology and Technology 53(3): 123-130.
- [110] Mondino, P., Casanova, L., Calero, G., Bentancur, O., Alaniz, S. (2012): Zimevit: un biofungicida que combina la acción de una bacteria y una levadura para el control del moho gris de la vid ocasionado por Botrytis cinerea. – Revista Brasileira de Agroecoogia 7(3): 127-134.

- [111] Mostofa, M. G., Li, W., Nguyen, K. H., Fujita, M., Tran, L. S. P. (2018): Strigolactones in plant adaptation to abiotic stresses: An emerging avenue of plant research. – Plant, Cell and Environment 41(10): 2227-2243.
- [112] Mukherjee, S., Sen, S. K. (2015): Exploration of novel rhizospheric yeast isolate as fertilizing soil inoculant for improvement of maize cultivation. – Journal of the Science of Food and Agriculture 95(7): 1491-1499.
- [113] Mukherjee, A., Verma, J. P., Gaurav, A. K., Chouhan, G. K., Patel, J. S., Hesham, A. E.-L. (2020): Yeast a potential bio-agent: future for plant growth and postharvest disease management for sustainable agriculture. – Applied Microbiology and Biotechnology 104: 1497-1510.
- [114] Naik, K., Mishra, S., Srichandan, H., Singh, P. K., Sarangi, P. K. (2019): Plant growth promoting microbes: Potential link to sustainable agriculture and environment. – Biocatalysis and Agricultural Biotechnology 21: 101326.
- [115] Nanjundappa, A., Bagyaraj, D. J., Saxena, A. K., Kumar, M., Chakdar, H. (2019): Interaction between arbuscular mycorrhizal fungi and *Bacillus spp.* in soil enhancing growth of crop plants. – Fungal Biology and Biotechnology 6: 1-10.
- [116] Nassar, A. H., El-Tarabily, K. A., Sivasithamparam, K. (2005): Promotion of plant growth by an auxin-producing isolate of the yeast Williopsis saturnus endophytic in maize (*Zea mays* L.) roots. – Biology and Fertility of Soils 42: 97-108.
- [117] Nelsen, C., Safir, G. (1982): Increased drought tolerance of mycorrhizal onion plants caused by improved phosphorus nutrition. Planta 154: 407-413.
- [118] Nimsi, K., Manjusha, K., Kathiresan, K., Arya, H. (2023): Plant growth-promoting yeasts (PGPY), the latest entrant for use in sustainable agriculture: a review. Journal of Applied Microbiology 134(2): lxac088.
- [119] Nishimura, K., Yamamoto, M., Nakagomi, T., Takiguchi, Y., Naganuma, T., Uzuka, Y. (2002): Biodegradation of triazine herbicides on polyvinylalcohol gel plates by the soil yeast *Lipomyces starkeyi*. Applied Microbiology and Biotechnology 58: 848-852.
- [120] Ortiz, N., Armada, E., Duque, E., Roldán, A., Azcón, R. (2015): Contribution of arbuscular mycorrhizal fungi and/or bacteria to enhancing plant drought tolerance under natural soil conditions: effectiveness of autochthonous or allochthonous strains. – Journal of Plant Physiology 174: 87-96.
- [121] Osakabe, Y., Osakabe, K., Shinozaki, K., Tran, L.-S. P. (2014): Response of plants to water stress. – Frontiers in Plant Science 5: 86.
- [122] Ouledali, S., Ennajeh, M., Ferrandino, A., Khemira, H., Schubert, A., Secchi, F. (2019): Influence of arbuscular mycorrhizal fungi inoculation on the control of stomata functioning by abscisic acid (ABA) in drought-stressed olive plants. – South African Journal of Botany 121: 152-158.
- [123] Pal, A., Pandey, S. (2016): Role of arbuscular mycorrhizal fungi on plant growth and reclamation of barren soil with wheat (*Triticum aestivum* L.) crop. International Journal of Soil Science 12: 25-31.
- [124] Parafati, L., Vitale, A., Restuccia, C., Cirvilleri, G. (2016): The effect of locust bean gum (LBG)-based edible coatings carrying biocontrol yeasts against *Penicillium digitatum* and *Penicillium italicum* causal agents of postharvest decay of mandarin fruit. – Food Microbiology 58: 87-94.
- [125] Parafati, L., Vitale, A., Restuccia, C., Cirvilleri, G. (2017): Performance evaluation of volatile organic compounds by antagonistic yeasts immobilized on hydrogel spheres against gray, green and blue postharvest decays. – Food Microbiology 63: 191-198.
- [126] Parihar, M., Rakshit, A., Meena, V. S., Gupta, V. K., Rana, K., Choudhary, M., Tiwari, G., Mishra, P. K., Pattanayak, A., Bisht, J. K., Jatav, S. S. (2020): The potential of arbuscular mycorrhizal fungi in C cycling: a review. Archives of Microbiology 202: 1581-1596.
- [127] Parniske, M. (2008): Arbuscular mycorrhiza: the mother of plant root endosymbioses. Nature Reviews Microbiology 6(10): 763-775.

- [128] Pavithra, D., Yapa, N. (2018): Arbuscular mycorrhizal fungi inoculation enhances drought stress tolerance of plants. - Groundwater for Sustainable Development 7: 490-494.
- [129] Pedranzani, H., Rodríguez-Rivera, M., Gutiérrez, M., Porcel, R., Hause, B., Ruiz-Lozano, J. M. (2016): Arbuscular mycorrhizal symbiosis regulates physiology and performance of Digitaria eriantha plants subjected to abiotic stresses by modulating antioxidant and jasmonate levels. – Mycorrhiza 26: 141-152.
- [130] Pimenta, R., Silva, J., Coelho, C., Morais, P., Rosa, C., Corrêa Jr, A. (2010): Integrated control of *Penicillium digitatum* by the predacious yeast *Saccharomycopsis crataegensis* and sodium bicarbonate on oranges. - Brazilian Journal of Microbiology 41: 404-410.
- [131] Posta, K., Duc, N. H. (2020): Benefits of arbuscular mycorrhizal fungi application to crop production under water scarcity. Drought Detect Solut. DOI: 10.5772/intechopen.86595.
- [132] Prasad, R., Bhola, D., Akdi, K., Cruz, C., KVSS, S., Tuteja, N., Varma, A. (2017): Introduction to mycorrhiza: historical development. - In: Varma, A., Prasad, R., Tuteja, N. (eds.) Mycorrhiza-function, diversity, State of the Art. Springer, pp. 1-7.
- [133] Rani, B., Madan, S., Pooja, K., Sharma, K., Kumari, N., Kumar, A. (2018): Mitigating the effect of drought stress on yield in wheat (Triticum aestivum) using arbuscular mycorrhiza fungi (Glomus mosseae). - Indian Journal of Agricultural Science 88: 95-100.
- [134] Razakatiana, A. T. E., Trap, J., Baohanta, R. H., Raherimandimby, M., Le Roux, C., Duponnois, R., Ramanankierana, H., Becquer, T. (2020): Benefits of dual inoculation with arbuscular mycorrhizal fungi and rhizobia on Phaseolus vulgaris planted in a lowfertility tropical soil. – Pedobiologia 83: 150685.
- [135] Robinson Boyer, L., Feng, W., Gulbis, N., Hajdu, K., Harrison, R. J., Jeffries, P., Xu, X. (2016): The use of arbuscular mycorrhizal fungi to improve strawberry production in coir substrate. - Frontiers in Plant Science 7: 1237.
- [136] Rosa-Magri, M. M., Avansini, S. H., Lopes-Assad, M. L., Tauk-Tornisielo, S. M., Ceccato-Antonini, S. R. (2012): Release of potassium from rock powder by the yeast *Torulaspora globosa.* – Brazilian Archives of Biology and Technology 55: 577-582.
- [137] Rouphael, Y., Franken, P., Schneider, C., Schwarz, D., Giovannetti, M., Agnolucci, M., De Pascale, S., Bonini, P., Colla, G. (2015): Arbuscular mycorrhizal fungi act as biostimulants in horticultural crops. - Scientia Horticulturae 196: 91-108.
- [138] Ruiz-Lozano, J. M., Aroca, R., Zamarreño, A. M., Molina, S., Andreo-Jiménez, B., Porcel, R., García-Mina, J. M., Ruyter-Spira, C., López-Ráez, J. A. (2016): Arbuscular mycorrhizal symbiosis induces strigolactone biosynthesis under drought and improves drought tolerance in lettuce and tomato. – Plant, Cell & Environment 39(2): 441-452.
- [139] Ruiz-Moyano, S., Hernández, A., Galvan, A. I., Córdoba, M. G., Casquete, R., Serradilla, M. J., Martín, A. (2020): Selection and application of antifungal VOCs-producing yeasts as biocontrol agents of grey mould in fruits. - Food Microbiology 92: 103556.
- [140] Sabia, E., Claps, S., Morone, G., Bruno, A., Sepe, L., Aleandri, R. (2015): Field inoculation of arbuscular mycorrhiza on maize (Zea mays L.) under low inputs: preliminary study on quantitative and qualitative aspects. - Italian Journal of Agronomy 10(1): 30-33.
- [141] Sadhana, B. (2014): Arbuscular Mycorrhizal Fungi (AMF) as a biofertilizer-a review. International Journal of Current Microbiology and Applied Sciences 3(4): 384-400.
- [142] Sbrana, C., Avio, L., Giovannetti, M. (2014): Beneficial mycorrhizal symbionts affecting the production of health-promoting phytochemicals. - Electrophoresis 35(11): 1535-1546.
- [143] Scherm, B., Ortu, G., Muzzu, A., Budroni, M., Arras, G., Migheli, Q. (2003): Biocontrol activity of antagonistic yeasts against Penicillium expansum on apple. - Journal of Plant Pathology 85(3): 205-213.
- [144] Schüssler, A., Schwarzott, D., Walker, C. (2001): A new fungal phylum, the Glomeromycota: phylogeny and evolution. – Mycological Research 105(12): 1413-1421.

- [145] Schüssler, A., Walker, C. (2010): The Glomeromycota: A Species List with New Families and New Gener. CreateSpace Independent Publishing Platform.
- [146] Sebastien, M., Jijakli, M. H. (2014): *Pichia anomala* and *Candida oleophila* in biocontrol of postharvest diseases of fruits: 20 years of fundamental and practical research. – Paper presented at the Post-harvest Pathology: Plant Pathology in the 21st Century, Contributions to the 10th International Congress ICPP 2013.
- [147] Silambarasan, S., Logeswari, P., Cornejo, P., Kannan, V. R. (2019): Evaluation of the production of exopolysaccharide by plant growth promoting yeast *Rhodotorula sp.* strain CAH2 under abiotic stress conditions. – International Journal of Biological Macromolecules 121: 55-62.
- [148] Smith, S. E., Read, D. (2008): Mineral nutrition, toxic element accumulation and water relations of arbuscular mycorrhizal plants. Mycorrhizal Symbiosis 145-VI.
- [149] Smith, S. E., Read, D. J. (2010): Mycorrhizal symbiosis. Academic press.
- [150] Spatafora, J. W., Chang, Y., Benny, G. L., Lazarus, K., Smith, M. E., Berbee, M. L., Bonito, G., Corradi, N., Grigoriev, I., Gryganskyi, A., Gryganskyi, A. (2016): A phylumlevel phylogenetic classification of zygomycete fungi based on genome-scale data. – Mycologia 108(5): 1028-1046.
- [151] Streletskii, R. A., Kachalkin, A. V., Glushakova, A. M., Yurkov, A. M., Demin, V. V. (2019): Yeasts producing zeatin. – PeerJ 7: e6474.
- [152] Subramanian, K., Santhanakrishnan, P., Balasubramanian, P. (2006): Responses of field grown tomato plants to arbuscular mycorrhizal fungal colonization under varying intensities of drought stress. – Scientia Horticulturae 107(3): 245-253.
- [153] Sui, Y., Sun, Z., Zou, Y., Li, W., Jiang, M., Luo, Y., Liao, W., Wang, Y., Gao, X., Liu, J. (2020): The Rlm1 transcription factor in *Candida oleophila* contributes to abiotic stress resistance and biocontrol efficacy against postharvest gray mold of kiwifruit. – Postharvest Biology and Technology 166: 111222.
- [154] Sun, Z., Song, J., Xin, X., Xie, X., Zhao, B. (2018): Arbuscular mycorrhizal fungal 14-3-3 proteins are involved in arbuscule formation and responses to abiotic stresses during AM symbiosis. – Frontiers in Microbiology 9: 91.
- [155] Syamsiyah, J., Herawati, A., Mujiyo (2018): The potential of arbuscular mycorrhizal fungi application on aggregate stability in alfisol soil. – IOP Conference Series: Earth and Environmental Science 142: 012045.
- [156] Symanczik, S., Lehmann, M. F., Wiemken, A., Boller, T., Courty, P.-E. (2018): Effects of two contrasted arbuscular mycorrhizal fungal isolates on nutrient uptake by *Sorghum bicolor* under drought. – Mycorrhiza 28: 779-785.
- [157] Taha, S. R., Seleiman, M. F., Alhammad, B. A., Alkahtani, J., Alwahibi, M. S., Mahdi, A. H. (2020): Activated Yeast extract enhances growth, anatomical structure, and productivity of *Lupinus termis* L. plants under actual salinity conditions. Agronomy 11(1): 74.
- [158] Talaat, N. B., Shawky, B. T. (2014): Protective effects of arbuscular mycorrhizal fungi on wheat (*Triticum aestivum* L.) plants exposed to salinity. – Environmental and Experimental Botany 98: 20-31.
- [159] Tsoata, E., Njock, S. R., Youmbi, E., Nwaga, D. (2015): Early effects of water stress on some biochemical and mineral parameters of mycorrhizal *Vigna subterranea* (L.) Verdc. (Fabaceae) cultivated in Cameroon. – International journal of agronomy and agricultural research 7(2): 21-35.
- [160] Tuladhar, K., Rao, N. S. (1985): Interaction of yeasts and some nitrogen fixing bacteria on nodulation of legumes. – Plant and Soil 84: 287-291.
- [161] Van der Linden, S., Goldberg, M. H. (2020): Alternative meta-analysis of behavioral interventions to promote action on climate change yields different conclusions. – Nature Communications 11(1): 3915.

- [162] Verbruggen, E., Van Der Heijden, M. G., Weedon, J. T., Kowalchuk, G. A., Röling, W. F. (2012): Community assembly, species richness and nestedness of arbuscular mycorrhizal fungi in agricultural soils. Molecular Ecology 21(10): 2341-2353.
- [163] Vero, S., Garmendia, G., González, M. B., Bentancur, O., Wisniewski, M. (2013): Evaluation of yeasts obtained from Antarctic soil samples as biocontrol agents for the management of postharvest diseases of apple (Malus× domestica). – FEMS yeast research 13(2): 189-199.
- [164] Wachowska, U., Głowacka, K. (2014): Antagonistic interactions between Aureobasidium pullulans and Fusarium culmorum, a fungal pathogen of winter wheat. – BioControl 59: 635-645.
- [165] Wang, F. (2017): Occurrence of arbuscular mycorrhizal fungi in mining-impacted sites and their contribution to ecological restoration: Mechanisms and applications. – Critical Reviews in Environmental Science and Technology 47(20): 1901-1957.
- [166] Wang, L., George, T. S., Feng, G. (2023): Concepts and consequences of the hyphosphere core microbiome for arbuscular mycorrhizal fungal fitness and function. – New Phytologist.
- [167] Xin, G., Glawe, D., Doty, S. L. (2009): Characterization of three endophytic, indole-3acetic acid-producing yeasts occurring in Populus trees. – Mycological Research 113(9): 973-980.
- [168] Yadav, P., Yadav, B., Deka, D., Narayan, O. P. (2023): Role of arbuscular mycorrhizas in regulating physiological and molecular aspects of plants under abiotic stress The Role of Growth Regulators and Phytohormones in Overcoming Environmental Stress. – Elsevier, pp. 399-433.
- [169] Yang, G., Liu, N., Lu, W., Wang, S., Kan, H., Zhang, Y., Xu, L., Chen, Y. (2014): The interaction between arbuscular mycorrhizal fungi and soil phosphorus availability influences plant community productivity and ecosystem stability. – Journal of Ecology 102(4): 1072-1082.
- [170] Yooyongwech, S., Samphumphuang, T., Tisarum, R., Theerawitaya, C., Cha-um, S. (2016): Arbuscular mycorrhizal fungi (AMF) improved water deficit tolerance in two different sweet potato genotypes involves osmotic adjustments via soluble sugar and free proline. – Scientia Horticulturae 198: 107-117.
- [171] Youssef, S. M., Abdella, E. M., Al-Elwany, O. A., Alshallash, K. S., Alharbi, K., Ibrahim, M. T., Tawfik, M. M., Abu-Elsaoud, A. M., Elkelish, A. (2022): Integrative application of foliar yeast extract and gibberellic acid improves morpho-physiological responses and nutrient uptake of *Solidago virgaurea* plant in alkaline soil. – Life 12(9): 1405.
- [172] Yurkov, A. M. (2018): Yeasts of the soil-obscure but precious. Yeast 35(5): 369-378.
- [173] Zeng, H., Tan, F., Zhang, Y., Feng, Y., Shu, Y., Wang, J. (2014): Effects of cultivation and return of *Bacillus thuringiensis* (Bt) maize on the diversity of the arbuscular mycorrhizal community in soils and roots of subsequently cultivated conventional maize. – Soil Biology and Biochemistry 75: 254-263.
- [174] Zhang, L., Feng, G., Declerck, S. (2018): Signal beyond nutrient, fructose, exuded by an arbuscular mycorrhizal fungus triggers phytate mineralization by a phosphate solubilizing bacterium. – The ISME Journal 12(10): 2339-2351.
- [175] Zhimo, V., Bhutia, D., Saha, J. (2016): Biological control of post-harvest fruit diseases using antagonistic yeasts in India. Journal of Plant Pathology 98(2): 275-283.
- [176] Zhou, Y. J., Buijs, N. A., Zhu, Z., Qin, J., Siewers, V., Nielsen, J. (2016): Production of fatty acid-derived oleochemicals and biofuels by synthetic yeast cell factories. – Nature Communications 7(1): 11709.